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(54) Title: MAIZE CELLULOSE SYNTHASES AND USES THEREOF

#### (57) Abstract

The invention provides isolated cellulose synthase nucleic acids and their encoded proteins. The present invention provides methods and compositions relating to altering cellulose synthase concentration and/or composition of plants. The invention further provides recombinant expression cassettes, host cells, and transgenic plants.

> Dhugga, et al. S/N 10/080,114

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# Maize Cellulose Synthases and Uses Thereof

## TECHNICAL FIELD

The present invention relates generally to plant molecular biology. More specifically, it relates to nucleic acids and methods for modulating their expression in plants.

## BACKGROUND OF THE INVENTION

Polysaccharides constitute the bulk of the plant cell walls and have been traditionally classified into three categories: cellulose, hemicellulose, and pectin. Fry, S. C. (1988), The growing plant cell wall: Chemical and metabolic analysis. New York: Longman Scientific & Technical. Whereas cellulose is made at the plasma membrane and directly laid down into the cell wall, hemicellulosic and pectic polymers are first made in the Golgi apparatus and then exported to the cell wall by exocytosis. Ray, P.

M., et al., (1976), Ber. Deutsch. Bot. Ges. Bd. 89, 121-146. The variety of chemical linkages in the pectic and hemicellulosic polysaccharides indicates that there must be tens of polysaccharide synthases in the Golgi apparatus. Darvill et al., (1980). The primary cell walls of flowering plants. In The Plant Cell (N. E. Tolbert, ed.), Vol. 1 in Series: The biochemistry of plants: A comprehensive treatise, eds. P.K. Stumpf and E.E. Conn
(New York: Academic Press), pp. 91-162.

Cellulose, by virtue of its ability to form semicrystalline microfibrils, has a very high tensile strength which approaches that of some metals. Niklas, K. J. (1992). Plant Biomechanics: An engineering approach to plant form and function, The University of Chicago Press, pp. 607. Bending strength of the culm of normal and brittle-culm mutants of barley has been found to be directly correlated with the concentration of cellulose in the cell wall. Kokubo, et al., (1989), Plant Physiology 91, 876-882; Kokubo, et al., (1991) Plant Physiology 97, 509-514.

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Even though sugar and polysaccharide compositions of the plant cell walls have been well characterized, very limited progress has been made toward identification of the enzymes involved in polysaccharides formation, the reason being their labile nature and recalcitrance to solubilization by available detergents. Sporadic claims for the identification of cellulose synthase from plant sources have been made over the years. Callaghan, T., and Benziman, M. (1984), Nature 311, 165-167; Okuda, et al., (1993),

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Plant Physiol. 101, 1131-1142. However, these claims have been met with skepticism. Callaghan, T., and Benziman, M. (1985), *Nature* 314, 383-384; Delmer, *et al.*, (1993), Plant Physiol. 103, 307-308. It was only recently that a putative gene for plant cellulose synthase (CelA) was cloned from the developing cotton fibers based on homology to the bacterial gene. Pear, *et al.*, *Proc. Natl. Acad. Sci.* (USA) 93, 12637-12642; Saxena, *et al.*, (1990), *Plant Molecular Biology* 15, 673-684; see also, WO 9818949.

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As brittle snap is a major problem in corn breeding, what is needed in the art are compositions and methods for manipulating cellulose concentration in the cell wall and thereby altering plant stalk quality for improved standability or silage. The present invention provides these and other advantages.

## SUMMARY OF THE INVENTION

Generally, it is the object of the present invention to provide nucleic acids and proteins relating to cellulose synthases. It is an object of the present invention to provide: 1) nucleic acids and proteins relating to maize cellulose synthases; 2) transgenic plants comprising the nucleic acids of the present invention; 3) methods for modulating, in a transgenic plant, the expression of the nucleic acids of the present invention.

Therefore, in one aspect, the present invention relates to an isolated nucleic acid comprising a member selected from the group consisting of (a) a polynucleotide having a specified sequence identity to a polynucleotide encoding a polypeptide of the present invention;; (b) a polynucleotide which is complementary to the polynucleotide of (a); and (c) a polynucleotide comprising a specified number of contiguous nucleotides from a polynucleotide of (a) or (b). The isolated nucleic acid can be DNA or RNA.

In another aspect, the present invention relates to recombinant expression cassettes, comprising a nucleic acid of the present invention operably linked to a promoter. In some embodiments, the nucleic acid is operably linked in antisense orientation to the promoter.

In another aspect, the present invention is directed to a host cell transfected with the recombinant expression cassette.

In a further aspect, the present invention relates to an isolated protein comprising a polypeptide having a specified number of contiguous amino acids encoded by an isolated nucleic acid of the present invention.

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In another aspect, the present invention relates to an isolated nucleic acid comprising a polynucleotide of specified length which selectively hybridizes under stringent conditions to a polynucleotide of the present invention, or a complement thereof. In some embodiments, the isolated nucleic acid is operably linked to a promoter.

In yet another aspect, the present invention relates to an isolated nucleic acid comprising a polynucleotide, the polynucleotide having a specified sequence identity to an identical length of a nucleic acid of the present invention or a complement thereof.

In another aspect, the present invention relates to an isolated nucleic acid comprising a polynucleotide having a sequence of a nucleic acid amplified from a Zea mays nucleic acid library using at least two primers or their complements, one of which selectively hyridizes under stringent conditions to a locus of the nucleic acid comprising the 5' terminal coding region and the other primer selectively hybridizing, under stringent conditions, to a locus of the nucleic acid comprising the 3' terminal coding region, and wherein both primers selectively hybridize within the coding region. In some embodiments, the nucleic acid library is a cDNA library.

In another aspect, the present invention relates to a recombinant expression cassette comprising a nucleic acid, wherein the nucleic acid is operably linked to a promoter. In some embodiments, the present invention relates to a host cell transfected with this recombinant expression cassette. In some embodiments, the present invention relates to a protein of the present invention which is produced from this host cell.

In a further aspect, the present invention relates to a heterologous promoter operably linked to a non-isolated polynucleotide of the present invention, wherein the polypeptide is encoded by a nucleic acid amplified from a nucleic acid library.

In yet another aspect, the present invention relates to a transgenic plant comprising a recombinant expression cassette comprising a plant promoter operably linked to any of the isolated nucleic acids of the present invention. In some embodiments, the transgenic plant is *Zea mays*. The present invention also provides transgenic seed from the transgenic plant.

In a further aspect, the present invention relates to a method of modulating expression of the genes encoding the proteins of the present invention in a plant cell capable of plant regeneration, comprising the steps of (a) transforming a plant cell with a recombinant expression cassette comprising a polynucleotide of the present invention

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operably linked to a promoter; (b) growing the plant cell under plant growing conditions; and (c) inducing expression of the polynucleotide for a time sufficient to modulate expression of the genes in the plant. In some embodiments, the plant is maize. Expression of the genes encoding the proteins of the present invention can be increased or decreased relative to a non-transformed control plant.

## **Definitions**

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Units, prefixes, and symbols may be denoted in their SI accepted form. Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation; amino acid sequences are written left to right in amino to carboxy orientation, respectively. Numeric ranges are inclusive of the numbers defining the range and include each integer within the defined range. Amino acids may be referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUB Biochemical Nomenclature Commission. Nucleotides, likewise, may be referred to by their commonly accepted single-letter codes. Unless otherwise provided for, software, electrical, and electronics terms as used herein are as defined in The New IEEE Standard Dictinary of Electrical and electronics Terms (5th edition, 1993). The terms defined below are more fully defined by reference to the specification as a whole.

By "amplified" is meant the construction of multiple copies of a nucleic acid sequence or multiple copies complementary to the nucleic acid sequence using at least one of the nucleic acid sequences as a template. Amplification systems include the polymerase chain reaction (PCR) system, ligase chain reaction (LCR) system, nucleic acid sequence based amplification (NASBA, Cangene, Mississauga, Ontario), Q-Beta Replicase systems, transcription-based amplification system (TAS), and strand displacement amplification (SDA). See, e.g., Diagnostic Molecular Microbiology: Principles and Applications, D. H. Persing et al., Ed., American Society for Microbiology, Washington, D.C. (1993). The product of amplification is termed an amplicon.

The term "antibody" includes reference to antigen binding forms of antibodies (e.g., Fab, F(ab)<sub>2</sub>). The term "antibody" frequently refers to a polypeptide substantially encoded by an immunoglobulin gene or immunoglobulin genes, or fragments thereof which specifically bind and recognize an analyte (antigen). However, while various antibody fragments can be defined in terms of the digestion of an intact antibody, one of

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skill will appreciate that such fragments may be synthesized *de novo* either chemically or by utilizing recombinant DNA methodology. Thus, the term antibody, as used herein, also includes antibody fragments such as single chain Fv, chimeric antibodies (i.e., comprising constant and variable regions from different species), humanized antibodies (i.e., comprising a complementarity determining region (CDR) from a non-human source) and heteroconjugate antibodies (e.g., bispecific antibodies).

The term "antigen" includes reference to a substance to which an antibody can be generated and/or to which the antibody is specifically immunoreactive. The specific immunoreactive sites within the antigen are known as epitopes or antigenic determinants. These epitopes can be a linear array of monomers in a polymeric composition - such as amino acids in a protein - or consist of or comprise a more complex secondary or tertiary structure. Those of skill will recognize that all immunogens (i.e., substances capable of eliciting an immune response) are antigens; however some antigens, such as haptens, are not immunogens but may be made immunogenic by coupling to a carrier molecule. An antibody immunologically reactive with a particular antigen can be generated *in vivo* or by recombinant methods such as selection of libraries of recombinant antibodies in phage or similar vectors. *See, e.g.*, Huse *et al.*, *Science* 246: 1275-1281 (1989); and Ward, *et al.*, *Nature* 341: 544-546 (1989); and Vaughan *et al.*, *Nature Biotech*. 14: 309-314 (1996).

As used herein, "antisense orientation" includes reference to a duplex polynucleotide sequence which is operably linked to a promoter in an orientation where the antisense strand is transcribed. The antisense strand is sufficiently complementary to an endogenous transcription product such that translation of the endogenous transcription product is often inhibited.

As used herein, "chromosomal region" includes reference to a length of a chromosome which may be measured by reference to the linear segment of DNA which it comprises. The chromosomal region can be defined by reference to two unique DNA sequences, i.e., markers.

The term "conservatively modified variants" applies to both amino acid and nucleic acid sequences. With respect to particular nucleic acid sequences, conservatively modified variants refers to those nucleic acids which encode identical or conservatively modified variants of the amino acid sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given protein.

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For instance, the codons GCA, GCC, GCG and GCU all encode the amino acid alanine. Thus, at every position where an alanine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations are "silent variations" and represent one species of conservatively modified variation. Every nucleic acid sequence herein which encodes a polypeptide also describes every possible silent variation of the nucleic acid. One of ordinary skill will recognize that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine; and UGG, which is ordinarily the only codon for tryptophan) can be modified to yield a functionally identical molecule. Accordingly, each silent variation of a nucleic acid which encodes a polypeptide of the present invention is implicit in each described polypeptide sequence and incorporated herein by reference.

As to amino acid sequences, one of skill will recognize that individual substitutions, deletions or additions to a nucleic acid, peptide, polypeptide, or protein sequence which alters, adds or deletes a single amino acid or a small percentage of amino acids in the encoded sequence is a "conservatively modified variant" where the alteration results in the substitution of an amino acid with a chemically similar amino acid. Thus, any number of amino acid residues selected from the group of integers consisting of from 1 to 15 can be so altered. Thus, for example, 1, 2, 3, 4, 5, 7, or 10 alterations can be made. Conservatively modified variants typically provide similar biological activity as the unmodified polypeptide sequence from which they are derived. For example, substrate specificity, enzyme activity, or ligand/receptor binding is generally at least 30%, 40%, 50%, 60%, 70%, 80%, or 90% of the native protein for it's native substrate. Conservative substitution tables providing functionally similar amino acids are well known in the art.

The following six groups each contain amino acids that are conservative substitutions for one another:

- 1) Alanine (A), Serine (S), Threonine (T);
- 2) Aspartic acid (D), Glutamic acid (E);
- 30 3) Asparagine (N), Glutamine (Q);
  - 4) Arginine (R), Lysine (K):
  - 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V); and
  - 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W).

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See also, Creighton (1984) Proteins W.H. Freeman and Company.

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By "encoding" or "encoded", with respect to a specified nucleic acid, is meant comprising the information for translation into the specified protein. A nucleic acid encoding a protein may comprise non-translated sequences (e.g., introns) within translated regions of the nucleic acid, or may lack such intervening non-translated sequences (e.g., as in cDNA). The information by which a protein is encoded is specified by the use of codons. Typically, the amino acid sequence is encoded by the nucleic acid using the "universal" genetic code. However, variants of the universal code, such as are present in some plant, animal, and fungal mitochondria, the bacterium *Mycoplasma capricolum* (*Proc. Natl. Acad. Sci. (USA)*, 82: 2306-2309 (1985)), or the ciliate *Macronucleus*, may be used when the nucleic acid is expressed using these organisms.

When the nucleic acid is prepared or altered synthetically, advantage can be taken of known codon preferences of the intended host where the nucleic acid is to be expressed. For example, although nucleic acid sequences of the present invention may be expressed in both monocotyledonous and dicotyledonous plant species, sequences can be modified to account for the specific codon preferences and GC content preferences of monocotyledons or dicotyledons as these preferences have been shown to differ (Murray et al. Nucl. Acids Res. 17: 477-498 (1989)). Thus, the maize preferred codon for a particular amino acid may be derived from known gene sequences from maize. Maize codon usage for 28 genes from maize plants are listed in Table 4 of Murray et al., above.

As used herein "full-length sequence" in reference to a specified polynucleotide or its encoded protein means having the entire amino acid sequence of, a native (non-synthetic), endogenous, catalytically active form of the specified protein. Methods to determine whether a sequence is full-length are well known in the art including such exemplary techniques as northern or western blots, primer extension, S1 protection, and ribonuclease protection. See, e.g., *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997). Comparison to known full-length homologous (orthologous and/or paralogous) sequences can also be used to identify full-length sequences of the present invention. Additionally, consensus sequences typically present at the 5' and 3' untranslated regions of mRNA aid in the identification of a polynucleotide as full-length. For example, the consensus sequence ANNNNAUGG,

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where the underlined codon represents the N-terminal methionine, aids in determining whether the polynucleotide has a complete 5' end. Consensus sequences at the 3' end, such as polyadenylation sequences, aid in determining whether the polynucleotide has a complete 3' end.

The term "gene activity" refers to one or more steps involved in gene expression, including transcription, translation, and the functioning of the protein encoded by the gene.

As used herein, "heterologous" in reference to a nucleic acid is a nucleic acid that originates from a foreign species, or, if from the same species, is substantially modified from its native form in composition and/or genomic locus by deliberate human intervention. For example, a promoter operably linked to a heterologous structural gene is from a species different from that from which the structural gene was derived, or, if from the same species, one or both are substantially modified from their original form. A heterologous protein may originate from a foreign species or, if from the same species, is substantially modified from its original form by deliberate human intervention.

By "host cell" is meant a cell which contains a vector and supports the replication and/or expression of the expression vector. Host cells may be prokaryotic cells such as *E. coli*, or eukaryotic cells such as yeast, insect, amphibian, or mammalian cells.

Preferably, host cells are monocotyledonous or dicotyledonous plant cells. A particularly preferred monocotyledonous host cell is a maize host cell.

The term "hybridization complex" includes reference to a duplex nucleic acid structure formed by two single-stranded nucleic acid sequences selectively hybridized with each other.

By "immunologically reactive conditions" or "immunoreactive conditions" is meant conditions which allow an antibody, generated to a particular epitope, to bind to that epitope to a detectably greater degree (e.g., at least 2-fold over background) than the antibody binds to substantially all other epitopes in a reaction mixture comprising the particular epitope. Immunologically reactive conditions are dependent upon the format of the antibody binding reaction and typically are those utilized in immunoassay protocols. See Harlow and Lane, *Antibodies, A Laboratory Manual*, Cold Spring Harbor Publications, New York (1988), for a description of immunoassay formats and conditions.

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The term "introduced" in the context of inserting a nucleic acid into a cell, means "transfection" or "transformation" or "transduction" and includes reference to the incorporation of a nucleic acid into a eukaryotic or prokaryotic cell where the nucleic acid may be incorporated into the genome of the cell (e.g., chromosome, plasmid, plastid or mitochondrial DNA), converted into an autonomous replicon, or transiently expressed (e.g., transfected mRNA).

The terms "isolated" refers to material, such as a nucleic acid or a protein, which is: (1) substantially or essentially free from components which normally accompany or interact with it as found in its naturally occurring environment. The isolated material optionally comprises material not found with the material in its natural environment; or (2) if the material is in its natural environment, the material has been synthetically (nonnaturally) altered by deliberate human intervention to a composition and/or placed at a locus in the cell (e.g., genome or subcellular organelle) not native to a material found in that environment. The alteration to yield the synthetic material can be performed on the material within or removed from its natural state. For example, a naturally occurring nucleic acid becomes an isolated nucleic acid if it is altered, or if it is transcribed from DNA which has been altered, by non-natural, synthetic (i.e., "man-made") methods performed within the cell from which it originates. See, e.g., Compounds and Methods for Site Directed Mutagenesis in Eukaryotic Cells, Kmiec, U.S. Patent No. 5,565,350; In Vivo Homologous Sequence Targeting in Eukaryotic Cells; Zarling et al., PCT/US93/03868. Likewise, a naturally occurring nucleic acid (e.g., a promoter) becomes isolated if it is introduced by non-naturally occurring means to a locus of the genome not native to that nucleic acid. Nucleic acids which are "isolated" as defined herein, are also referred to as "heterologous" nucleic acids.

Unless otherwise stated, the term "cellulose synthase nucleic acid" is a nucleic acid of the present invention and means a nucleic acid comprising a polynucleotide of the present invention (a "cellulose synthase polynucleotide") encoding a cellulose synthase polypeptide. A "cellulose synthase gene" is a gene of the present invention and refers to a non-heterologous genomic form of a full-length cellulose synthase polynucleotide.

As used herein, "localized within the chromosomal region defined by and including" with respect to particular markers includes reference to a contiguous length of a chromosome delimited by and including the stated markers.

As used herein, "marker" includes reference to a locus on a chromosome that serves to identify a unique position on the chromosome. A "polymorphic marker" includes reference to a marker which appears in multiple forms (alleles) such that different forms of the marker, when they are present in a homologous pair, allow transmission of each of the chromosomes in that pair to be followed. A genotype may be defined by use of one or a plurality of markers.

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As used herein, "nucleic acid" includes reference to a deoxyribonucleotide or ribonucleotide polymer in either single- or double-stranded form, and unless otherwise limited, encompasses known analogues having the essential nature of natural nucleotides in that they hybridize to single-stranded nucleic acids in a manner similar to naturally occurring nucleotides (e.g., peptide nucleic acids).

By "nucleic acid library" is meant a collection of isolated DNA or RNA molecules which comprise and substantially represent the entire transcribed fraction of a genome of a specified organism. Construction of exemplary nucleic acid libraries, such as genomic and cDNA libraries, is taught in standard molecular biology references such as Berger and Kimmel, *Guide to Molecular Cloning Techniques, Methods in Enzymology*, Vol. 152, Academic Press, Inc., San Diego, CA (Berger); Sambrook *et al.*, *Molecular Cloning - A Laboratory Manual*, 2nd ed., Vol. 1-3 (1989); and *Current Protocols in Molecular Biology*, F.M. Ausubel *et al.*, Eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc. (1994 Supplement).

As used herein "operably linked" includes reference to a functional linkage between a promoter and a second sequence, wherein the promoter sequence initiates and mediates transcription of the DNA sequence corresponding to the second sequence. Generally, operably linked means that the nucleic acid sequences being linked are contiguous and, where necessary to join two protein coding regions, contiguous and in the same reading frame.

As used herein, the term "plant" includes reference to whole plants, plant parts or organs (e.g., leaves, stems, roots, etc.), plant cells, seeds and progeny of same. Plant cell, as used herein includes, without limitation, cells obtained from or found in: seeds, suspension cultures, embryos, meristematic regions, callus tissue, leaves, roots, shoots, gametophytes, sporophytes, pollen, and microspores. Plant cells can also be understood to include modified cells, such as protoplasts, obtained from the aforementioned tissues.

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The class of plants which can be used in the methods of the invention is generally as broad as the class of higher plants amenable to transformation techniques, including both monocotyledonous and dicotyledonous plants. Particularly preferred plants include maize, soybean, sunflower, sorghum, canola, wheat, alfalfa, cotton, rice, barley and millet.

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As used herein, "polynucleotide" includes reference to a deoxyribopolynucleotide, ribopolynucleotide, or analogs thereof that have the essential nature of a natural ribonucleotide in that they hybridize, under stringent hybridization conditions, to substantially the same nucleotide sequence as naturally occurring nucleotides and/or allow translation into the same amino acid(s) as the naturally occurring nucleotide(s). A polynucleotide can be full-length or a subsequence of a native or heterologous structural or regulatory gene. Unless otherwise indicated, the term includes reference to the specified sequence as well as the complementary sequence thereof. Thus, DNAs or RNAs with backbones modified for stability or for other reasons are "polynucleotides" as that term is intended herein. Moreover, DNAs or RNAs comprising unusual bases, such as inosine, or modified bases, such as tritylated bases, to name just two examples, are polynucleotides as the term is used herein. It will be appreciated that a great variety of modifications have been made to DNA and RNA that serve many useful purposes known to those of skill in the art. The term polynucleotide as it is employed herein embraces such chemically, enzymatically or metabolically modified forms of polynucleotides, as well as the chemical forms of DNA and RNA characteristic of viruses and cells, including among other things, simple and complex cells.

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical analogue of a corresponding naturally occurring amino acid, as well as to naturally occurring amino acid polymers. The essential nature of such analogues of naturally occurring amino acids is that, when incorporated into a protein, that protein is specifically reactive to antibodies elicited to the same protein but consisting entirely of naturally occurring amino acids. The terms "polypeptide", "peptide" and "protein" are also inclusive of modifications including, but not limited to, glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation. Exemplary modifications are described in most basic texts, such as, *Proteins - Structure* 

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and Molecular Properties, 2nd ed., T. E. Creighton, W. H. Freeman and Company, New York (1993). Many detailed reviews are available on this subject, such as, for example, those provided by Wold, F., Post-translational Protein Modifications: Perspectives and Prospects, pp. 1-12 in Posttranslational Covalent Modification of Proteins, B. C. Johnson, Ed., Academic Press, New York (1983); Seifter et al., Meth. Enzymol. 182: 626-646 (1990) and Rattan et al., Protein Synthesis: Posttranslational Modifications and Aging, Ann. N.Y. Acad. Sci. 663: 48-62 (1992). It will be appreciated, as is well known and as noted above, that polypeptides are not always entirely linear. For instance, polypeptides may be branched as a result of ubiquitination, and they may be circular, with or without branching, generally as a result of posttranslation events, including natural processing event and events brought about by human manipulation which do not occur naturally. Circular, branched and branched circular polypeptides may be synthesized by nontranslation natural process and by entirely synthetic methods, as well. Modifications can occur anywhere in a polypeptide, including the peptide backbone, the amino acid sidechains and the amino or carboxyl termini. In fact, blockage of the amino or carboxyl group in a polypeptide, or both, by a covalent modification, is common in naturally occurring and synthetic polypeptides and such modifications may be present in polypeptides of the present invention, as well. For instance, the amino terminal residue of polypeptides made in E. coli or other cells, prior to proteolytic processing, almost invariably will be N-formylmethionine. During post-translational modification of the peptide, a methionine residue at the NH<sub>2</sub>-terminus may be deleted. Accordingly, this invention contemplates the use of both the methionine-containing and the methionine-less amino terminal variants of the protein of the invention. In general, as used herein, the term polypeptide encompasses all such modifications, particularly those that are present in polypeptides synthesized by expressing a polynucleotide in a host cell.

As used herein "promoter" includes reference to a region of DNA upstream from the start of transcription and involved in recognition and binding of RNA polymerase and other proteins to initiate transcription. A "plant promoter" is a promoter capable of initiating transcription in plant cells. Exemplary plant promoters include, but are not limited to, those that are obtained from plants, plant viruses, and bacteria which comprise genes expressed in plant cells such *Agrobacterium* or *Rhizobium*. Examples of promoters under developmental control include promoters that preferentially initiate transcription in certain tissues, such as leaves, roots, or seeds. Such promoters are

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referred to as "tissue preferred". Promoters which initiate transcription only in certain tissue are referred to as "tissue specific". A "cell type" specific promoter primarily drives expression in certain cell types in one or more organs, for example, vascular cells in roots or leaves. An "inducible" promoter is a promoter which is under environmental control. Examples of environmental conditions that may effect transcription by inducible promoters include anaerobic conditions or the presence of light. Tissue specific, tissue preferred, cell type specific, and inducible promoters constitute the class of "non-constitutive" promoters. A "constitutive" promoter is a promoter which is active under most environmental conditions.

The term "cellulose synthase polypeptide" is a polypeptide of the present invention and refers to one or more amino acid sequences, in glycosylated or non-glycosylated form. The term is also inclusive of fragments, variants, homologs, alleles or precursors (e.g., preproproteins or proproteins) thereof. A "cellulose synthase protein" is a protein of the present invention and comprises a cellulose synthase polypeptide.

As used herein "recombinant" includes reference to a cell or vector, that has been modified by the introduction of a heterologous nucleic acid or that the cell is derived from a cell so modified. Thus, for example, recombinant cells express genes that are not found in identical form within the native (non-recombinant) form of the cell or express native genes that are otherwise abnormally expressed, under-expressed or not expressed at all as a result of deliberate human intervention. The term "recombinant" as used herein does not encompass the alteration of the cell or vector by naturally occurring events (e.g., spontaneous mutation, natural transformation/transduction/transposition) such as those occurring without deliberate human intervention.

As used herein, a "recombinant expression cassette" is a nucleic acid construct, generated recombinantly or synthetically, with a series of specified nucleic acid elements which permit transcription of a particular nucleic acid in a host cell. The recombinant expression cassette can be incorporated into a plasmid, chromosome, mitochondrial DNA, plastid DNA, virus, or nucleic acid fragment. Typically, the recombinant expression cassette portion of an expression vector includes, among other sequences, a nucleic acid to be transcribed, and a promoter.

The term "residue" or "amino acid residue" or "amino acid" are used interchangeably herein to refer to an amino acid that is incorporated into a protein,

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polypeptide, or peptide (collectively "protein"). The amino acid may be a naturally occurring amino acid and, unless otherwise limited, may encompass known analogs of natural amino acids that can function in a similar manner as naturally occurring amino acids.

The term "selectively hybridizes" includes reference to hybridization, under stringent hybridization conditions, of a nucleic acid sequence to a specified nucleic acid target sequence to a detectably greater degree (e.g., at least 2-fold over background) than its hybridization to non-target nucleic acid sequences and to the substantial exclusion of non-target nucleic acids. Selectively hybridizing sequences typically have about at least 80% sequence identity, preferably 90% sequence identity, and most preferably 100% sequence identity (i.e., complementary) with each other.

The term "specifically reactive", includes reference to a binding reaction between an antibody and a protein having an epitope recognized by the antigen binding site of the antibody. This binding reaction is determinative of the presence of a protein having the recognized epitope amongst the presence of a heterogeneous population of proteins and other biologics. Thus, under designated immunoassay conditions, the specified antibodies bind to an analyte having the recognized epitope to a substantially greater degree (e.g., at least 2-fold over background) than to substantially all other analytes lacking the epitope which are present in the sample.

The terms "stringent conditions" or "stringent hybridization conditions" includes reference to conditions under which a probe will hybridize to its target sequence, to a detectably greater degree than other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence-dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences can be identified which are 100% complementary to the probe (homologous probing). Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing). Generally, a probe is less than about 1000 nucleotides in length, preferably less than 500 nucleotides in length.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (e.g., 10 to 50 nucleotides) and at least about 60°C for long probes (e.g., greater than 50

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nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C.

Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T<sub>m</sub> can be approximated from the equation of Meinkoth and Wahl, Anal. Biochem., 138:267-284 (1984):  $T_m = 81.5 \, ^{\circ}\text{C} + 16.6 \, (\log M) + 0.41 \, (\% GC) - 10.41 \, (\% GC) + 10.41 \, (\% GC)$ 0.61 (% form) - 500/L; where M is the molarity of monovalent cations, %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The T<sub>m</sub> is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe. T<sub>m</sub> is reduced by about 1 °C for each 1% of mismatching; thus, T<sub>m</sub>, hybridization and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with  $\geq 90\%$  identity are sought, the  $T_m$  can be decreased 10 °C. Generally, stringent conditions are selected to be about 5 °C lower than the thermal melting point (T<sub>m</sub>) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4 °C lower than the thermal melting point (T<sub>m</sub>); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10 °C lower than the thermal melting point (T<sub>m</sub>); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20 °C lower than the thermal melting point  $(T_m)$ . Using the equation, hybridization and wash compositions, and desired T<sub>m</sub>, those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T<sub>m</sub> of less than 45 °C (aqueous solution) or 32 °C (formamide solution) it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive

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guide to the hybridization of nucleic acids is found in Tijssen, Laboratory Techniques in Biochemistry and Molecular Biology--Hybridization with Nucleic Acid Probes, Part I, Chapter 2 "Overview of principles of hybridization and the strategy of nucleic acid probe assays", Elsevier, New York (1993); and Current Protocols in Molecular Biology, Chapter 2, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995).

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As used herein, "transgenic plant" includes reference to a plant which comprises within its genome a heterologous polynucleotide. Generally, the heterologous polynucleotide is stably integrated within the genome such that the polynucleotide is passed on to successive generations. The heterologous polynucleotide may be integrated into the genome alone or as part of a recombinant expression cassette. "Transgenic" is used herein to include any cell, cell line, callus, tissue, plant part or plant, the genotype of which has been altered by the presence of heterologous nucleic acid including those transgenics initially so altered as well as those created by sexual crosses or asexual propagation from the initial transgenic. The term "transgenic" as used herein does not encompass the alteration of the genome (chromosomal or extra-chromosomal) by conventional plant breeding methods or by naturally occurring events such as random cross-fertilization, non-recombinant viral infection, non-recombinant bacterial transformation, non-recombinant transposition, or spontaneous mutation.

As used herein, "vector" includes reference to a nucleic acid used in transfection of a host cell and into which can be inserted a polynucleotide. Vectors are often replicons. Expression vectors permit transcription of a nucleic acid inserted therein.

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

- (a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.
- (b) As used herein, "comparison window" means includes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence may be compared to a reference sequence and wherein the

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portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

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Methods of alignment of sequences for comparison are well-known in the art. Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman, Adv. Appl. Math. 2: 482 (1981); by the homology alignment algorithm of Needleman and Wunsch, J. Mol. Biol. 48: 443 (1970); by the search for similarity method of Pearson and Lipman, Proc. Natl. Acad. Sci. 85: 2444 (1988); by computerized implementations of these algorithms, including, but not limited to: CLUSTAL in the PC/Gene program by Intelligenetics, Mountain View, California, GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, Wisconsin, USA; the CLUSTAL program is well described by Higgins and Sharp, Gene 73: 237-244 (1988); Higgins and Sharp, CABIOS 5: 151-153 (1989); Corpet, et al., Nucleic 20 Acids Research 16: 10881-90 (1988); Huang, et al., Computer Applications in the Biosciences 8: 155-65 (1992), and Pearson, et al., Methods in Molecular Biology 24: 307-331 (1994). The BLAST family of programs which can be used for database similarity searches includes: BLASTN for nucleotide query sequences against nucleotide database sequences; BLASTX for nucleotide query sequences against protein database 25 sequences; BLASTP for protein query sequences against protein database sequences; TBLASTN for protein query sequences against nucleotide database sequences; and TBLASTX for nucleotide query sequences against nucleotide database sequences. See, Current Protocols in Molecular Biology, Chapter 19, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995).

Unless otherwise stated, sequence identity/similarity values provided herein refer to the value obtained using the BLAST 2.0 suite of programs using default parameters. Altschul et al., Nucleic Acids Res. 25:3389-3402 (1997). Software for performing BLAST analyses is publicly available, e.g., through the National Center for Biotechnology

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Information (http://www.ncbi.nlm.nih.gov/). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul et al., supra). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff (1989) Proc. Natl. Acad. Sci. USA 89:10915).

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (*see*, *e.g.*, Karlin & Altschul, *Proc. Nat'l. Acad. Sci. USA* 90:5873-5787 (1993)). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance.

BLAST searches assume that proteins can be modeled as random sequences. However, many real proteins comprise regions of nonrandom sequences which may be homopolymeric tracts, short-period repeats, or regions enriched in one or more amino acids. Such low-complexity regions may be aligned between unrelated proteins even though other regions of the protein are entirely dissimilar. A number of low-complexity filter programs can be employed to reduce such low-complexity alignments. For example, the SEG (Wooten and Federhen, *Comput. Chem.*, 17:149-163 (1993)) and

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XNU (Claverie and States, *Comput. Chem.*, 17:191-201 (1993)) low-complexity filters can be employed alone or in combination.

- (c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences includes reference to the residues in the two sequences which are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g. charge or hydrophobicity) and therefore do not change the functional properties of the molecule. Where sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences which differ by such conservative substitutions are said to have "sequence similarity" or "similarity". Means for making this adjustment are well-known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, e.g., according to the algorithm of Meyers and Miller, Computer Applic. Biol. Sci., 4: 11-17 (1988) e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California, USA).
- (d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity.

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(e) (i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 70% sequence identity, preferably at least 80%, more preferably at least 90% and most preferably at least 95%, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 60%, more preferably at least 70%, 80%, 90%, and most preferably at least 95%.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. However, nucleic acids which do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides which they encode are substantially identical. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is that the polypeptide which the first nucleic acid encodes is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

(e) (ii) The terms "substantial identity" in the context of a peptide indicates that a peptide comprises a sequence with at least 70% sequence identity to a reference sequence, preferably 80%, more preferably 85%, most preferably at least 90% or 95% sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970). An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide is substantially identical to a second peptide, for example, where the two peptides differ only by a conservative substitution. Peptides which are "substantially similar" share sequences as noted above except that residue positions which are not identical may differ by conservative amino acid changes.

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#### Overview

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The present invention provides, among other things, compositions and methods for modulating (i.e., increasing or decreasing) the level of polypeptides of the present invention in plants. In particular, the polypeptides of the present invention can be expressed at developmental stages, in tissues, and/or in quantities which are uncharacteristic of non-recombinantly engineered plants. Thus, the present invention provides utility in such exemplary applications as improvement of stalk quality for improved stand or silage. Further, the present invention provides for an increased concentration of cellulose in the pericarp; hardening the kernel and thus improving its handling ability.

The present invention also provides isolated nucleic acid comprising polynucleotides of sufficient length and complementarity to a gene of the present invention to use as probes or amplification primers in the detection, quantitation, or isolation of gene transcripts. For example, isolated nucleic acids of the present invention can be used as probes in detecting deficiencies in the level of mRNA in screenings for desired transgenic plants, for detecting mutations in the gene (e.g., substitutions, deletions, or additions), for monitoring upregulation of expression or changes in enzyme activity in screening assays of compounds, for detection of any number of allelic variants (polymorphisms) of the gene, or for use as molecular markers in plant breeding programs. The isolated nucleic acids of the present invention can also be used for recombinant expression of their encoded polypeptides, or for use as immunogens in the preparation and/or screening of antibodies. The isolated nucleic acids of the present invention can also be employed for use in sense or antisense suppression of one or more genes of the present invention in a host cell, tissue, or plant. Attachment of chemical agents which bind, intercalate, cleave and/or crosslink to the isolated nucleic acids of the present invention can also be used to modulate transcription or translation.

The present invention also provides isolated proteins comprising a polypeptide of the present invention (e.g., preproenzyme, proenzyme, or enzymes). The present invention also provides proteins comprising at least one epitope from a polypeptide of the present invention. The proteins of the present invention can be employed in assays for enzyme agonists or antagonists of enzyme function, or for use as immunogens or antigens to obtain antibodies specifically immunoreactive with a protein of the present invention. Such antibodies can be used in assays for expression levels, for identifying

and/or isolating nucleic acids of the present invention from expression libraries, or for purification of polypeptides of the present invention.

The isolated nucleic acids and proteins of the present invention can be used over a broad range of plant types, particularly monocots such as the species of the Family

5 Graminiae including Sorghum bicolor and Zea mays. The isolated nucleic acid and proteins of the present invention can also be used in species from the genera: Cucurbita, Rosa, Vitis, Juglans, Fragaria, Lotus, Medicago, Onobrychis, Trifolium, Trigonella, Vigna, Citrus, Linum, Geranium, Manihot, Daucus, Arabidopsis, Brassica, Raphanus, Sinapis, Atropa, Capsicum, Datura, Hyoscyamus, Lycopersicon, Nicotiana, Solanum,

10 Petunia, Digitalis, Majorana, Ciahorium, Helianthus, Lactuca, Bromus, Asparagus, Antirrhinum, Heterocallis, Nemesis, Pelargonium, Panieum, Pennisetum, Ranunculus, Senecio, Salpiglossis, Cucumis, Browaalia, Glycine, Pisum, Phaseolus, Lolium, Oryza, Avena, Hordeum, Secale, Triticum, Bambusa, Dendrocalamus, and Melocanna.

### 15 Nucleic Acids

The present invention provides, among other things, isolated nucleic acids of RNA, DNA, and analogs and/or chimeras thereof, comprising a polynucleotide of the present invention.

A polynucleotide of the present invention is inclusive of:

- 20 (a) a polynucleotide encoding a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58, and conservatively modified and polymorphic variants thereof, including exemplary polynucleotides of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57;
- (b) a polynucleotide which is the product of amplification from a *Zea mays*nucleic acid library using primer pairs which selectively hybridize under stringent conditions to loci within a polynucleotide selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57, wherein the polynucleotide has substantial sequence identity to a polynucleotide selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57:
  - (c) a polynucleotide which selectively hybridizes to a polynucleotide of (a) or (b);
  - (d) a polynucleotide having a specified sequence identity with polynucleotides of (a), (b), or (c);

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- (e) a polynucleotide encoding a protein having a specified number of contiguous amino acids from a prototype polypeptide, wherein the protein is specifically recognized by antisera elicited by presentation of the protein and wherein the protein does not detectably immunoreact to antisera which has been fully immunosorbed with the protein;
  - (f) complementary sequences of polynucleotides of (a), (b), (c), (d), or (e); and
- (g) a polynucleotide comprising at least a specific number of contiguous nucleotides from a polynucleotide of (a), (b), (c), (d), (e), or (f).
- A. Polynucleotides Encoding A Polypeptide of the Present Invention or Conservatively

  Modified or Polymorphic Variants Thereof

As indicated in (a), above, the present invention provides isolated nucleic acids comprising a polynucleotide of the present invention, wherein the polynucleotide encodes a polypeptide of the present invention, or conservatively modified or polymorphic variants thereof. Those of skill in the art will recognize that the degeneracy of the 15 genetic code allows for a plurality of polynucleotides to encode for the identical amino acid sequence. Such "silent variations" can be used, for example, to selectively hybridize and detect allelic variants of polynucleotides of the present invention. Accordingly, the present invention includes polynucleotides of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57, and silent variations of 20 polynucleotides encoding a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58. The present invention further provides isolated nucleic acids comprising polynucleotides encoding conservatively modified variants of a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58. Additionally, the present invention further provides isolated nucleic acids comprising 25 polynucleotides encoding one or more polymorphic (allelic) variants of polypeptides/polynucleotides. Polymorphic variants are frequently used to follow segregation of chromosomal regions in, for example, marker assisted selection methods for crop improvement.

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As indicated in (b), above, the present invention provides an isolated nucleic acid comprising a polynucleotide of the present invention, wherein the polynucleotides are amplified from a Zea mays nucleic acid library. Zea mays lines B73, PHRE1, A632, BMS-P2#10, W23, and Mo17 are known and publicly available. Other publicly known and available maize lines can be obtained from the Maize Genetics Cooperation (Urbana, 5 IL). The nucleic acid library may be a cDNA library, a genomic library, or a library generally constructed from nuclear transcripts at any stage of intron processing. cDNA libraries can be normalized to increase the representation of relatively rare cDNAs. In optional embodiments, the cDNA library is constructed using a full-length cDNA 10 synthesis method. Examples of such methods include Oligo-Capping (Maruyama, K. and Sugano, S. Gene 138: 171-174, 1994), Biotinylated CAP Trapper (Carninci, P., Kvan, C., et al. Genomics 37: 327-336, 1996), and CAP Retention Procedure (Edery, E., Chu, L.L., et al. Molecular and Cellular Biology 15: 3363-3371, 1995). cDNA synthesis is often catalyzed at 50-55°C to prevent formation of RNA secondary 15 structure. Examples of reverse transcriptases that are relatively stable at these temperatures are SuperScript II Reverse Transcriptase (Life Technologies, Inc.), AMV Reverse Transcriptase (Boehringer Mannheim) and RetroAmp Reverse Transcriptase (Epicentre). Rapidly growing tissues, or rapidly dividing cells are preferably used as mRNA sources such as from the elongating internode of corn plants.

The polynucleotides of the present invention include those amplified using the following primer pairs:

SEQ ID NOS: 3 and 4 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 1;

SEQ ID NOS: 7 and 8 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 5; and

SEQ ID NOS: 11 and 12 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 9.

SEQ ID NOS: 15 and 16 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 13.

30 SEQ ID NOS: 19 and 20 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 17;

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SEQ ID NOS: 23 and 24 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 21; and

SEQ ID NOS: 27 and 28 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 25.

SEQ ID NOS: 31 and 32 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 29.

SEQ ID NOS: 35 and 36 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 33;

SEQ ID NOS: 39 and 40 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 37; and

SEQ ID NOS: 43 and 44 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 41.

SEQ ID NOS: 47 and 48 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 45.

SEQ ID NOS: 51 and 52 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 49;

SEQ ID NOS: 55 and 56 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 53; and

SEQ ID NOS: 59 and 60 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 57.

The present invention also provides subsequences of the polynucleotides of the present invention. A variety of subsequences can be obtained using primers which selectively hybridize under stringent conditions to at least two sites within a polynucleotide of the present invention, or to two sites within the nucleic acid which flank and comprise a polynucleotide of the present invention, or to a site within a polynucleotide of the present invention and a site within the nucleic acid which comprises it. Primers are chosen to selectively hybridize, under stringent hybridization conditions, to a polynucleotide of the present invention. Generally, the primers are complementary to a subsequence of the target nucleic acid which they amplify. As those skilled in the art will appreciate, the sites to which the primer pairs will selectively hybridize are chosen such that a single contiguous nucleic acid can be formed under the desired amplification conditions.

In optional embodiments, the primers will be constructed so that they selectively hybridize under stringent conditions to a sequence (or its complement) within the target nucleic acid which comprises the codon encoding the carboxy or amino terminal amino acid residue (i.e., the 3' terminal coding region and 5' terminal coding region, respectively) of the polynucleotides of the present invention. Optionally within these embodiments, the primers will be constructed to selectively hybridize entirely within the coding region of the target polynucleotide of the present invention such that the product of amplification of a cDNA target will consist of the coding region of that cDNA. The primer length in nucleotides is selected from the group of integers consisting of from at 10 least 15 to 50. Thus, the primers can be at least 15, 18, 20, 25, 30, 40, or 50 nucleotides in length. Those of skill will recognize that a lengthened primer sequence can be employed to increase specificity of binding (i.e., annealing) to a target sequence. A non-annealing sequence at the 5'end of a primer (a "tail") can be added, for example, to introduce a cloning site at the terminal ends of the amplicon.

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The amplification products can be translated using expression systems well known to those of skill in the art and as discussed, *infra*. The resulting translation products can be confirmed as polypeptides of the present invention by, for example, assaying for the appropriate catalytic activity (e.g., specific activity and/or substrate specificity), or verifying the presence of one or more linear epitopes which are specific to a polypeptide of the present invention. Methods for protein synthesis from PCR derived templates are known in the art and available commercially. See, e.g., Amersham Life Sciences, Inc, Catalog '97, p.354.

Methods for obtaining 5' and/or 3' ends of a vector insert are well known in the art. See, e.g., RACE (Rapid Amplification of Complementary Ends) as described in Frohman, M. A., in PCR Protocols: A Guide to Methods and Applications, M. A. Innis, D. H. Gelfand, J. J. Sninsky, T. J. White, Eds. (Academic Press, Inc., San Diego, 1990), pp. 28-38.); see also, U.S. Pat. No. 5,470,722, and Current Protocols in Molecular Biology, Unit 15.6, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995); Frohman and Martin, Techniques 1:165 (1989).

C. Polynucleotides Which Selectively Hybridize to a Polynucleotide of (A) or (B)

As indicated in (c), above, the present invention provides isolated nucleic acids comprising polynucleotides of the present invention, wherein the polynucleotides

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selectively hybridize, under selective hybridization conditions, to a polynucleotide of paragraphs (A) or (B) as discussed, above. Thus, the polynucleotides of this embodiment can be used for isolating, detecting, and/or quantifying nucleic acids comprising the polynucleotides of (A) or (B). For example, polynucleotides of the present invention can be used to identify, isolate, or amplify partial or full-length clones in a deposited library. In some embodiments, the polynucleotides are genomic or cDNA sequences isolated or otherwise complementary to a cDNA from a dicot or monocot nucleic acid library. Exemplary species of monocots and dicots include, but are not limited to: corn, canola, soybean, cotton, wheat, sorghum, sunflower, oats, sugar cane, millet, barley, and rice. Optionally, the cDNA library comprises at least 80% full-length sequences, preferably at least 85% or 90% full-length sequences, and more preferably at least 95% full-length sequences. The cDNA libraries can be normalized to increase the representation of rare sequences. Low stringency hybridization conditions are typically, but not exclusively, employed with sequences having a reduced sequence identity relative to complementary sequences. Moderate and high stringency conditions can optionally be employed for sequences of greater identity. Low stringency conditions allow selective hybridization of sequences having about 70% sequence identity and can be employed to identify orthologous or paralogous sequences.

D. Polynucleotides Having a Specific Sequence Identity with the Polynucleotides of (A), (B) or (C)

As indicated in (d), above, the present invention provides isolated nucleic acids comprising polynucleotides of the present invention, wherein the polynucleotides have a specified identity at the nucleotide level to a polynucleotide as disclosed above in paragraphs (A), (B), or (C). The percentage of identity to a reference sequence is at least 60% and, rounded upwards to the nearest integer, can be expressed as an integer selected from the group of integers consisting of from 60 to 99. Thus, for example, the percentage of identity to a reference sequence can be at least 70%, 75%, 80%, 85%, 90%, or 95%.

Optionally, the polynucleotides of this embodiment will share an epitope with a polypeptide encoded by the polynucleotides of (A), (B), or (C). Thus, these polynucleotides encode a first polypeptide which elicits production of antisera comprising antibodies which are specifically reactive to a second polypeptide encoded by

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a polynucleotide of (A), (B), or (C). However, the first polypeptide does not bind to antisera raised against itself when the antisera has been fully immunosorbed with the first polypeptide. Hence, the polynucleotides of this embodiment can be used to generate antibodies for use in, for example, the screening of expression libraries for nucleic acids comprising polynucleotides of (A), (B), or (C), or for purification of, or in immunoassays for, polypeptides encoded by the polynucleotides of (A), (B), or (C). The polynucleotides of this embodiment embrace nucleic acid sequences which can be employed for selective hybridization to a polynucleotide encoding a polypeptide of the present invention.

Screening polypeptides for specific binding to antisera can be conveniently achieved using peptide display libraries. This method involves the screening of large collections of peptides for individual members having the desired function or structure. Antibody screening of peptide display libraries is well known in the art. The displayed peptide sequences can be from 3 to 5000 or more amino acids in length, frequently from 5-100 amino acids long, and often from about 8 to 15 amino acids long. In addition to direct chemical synthetic methods for generating peptide libraries, several recombinant DNA methods have been described. One type involves the display of a peptide sequence on the surface of a bacteriophage or cell. Each bacteriophage or cell contains the nucleotide sequence encoding the particular displayed peptide sequence. Such methods are described in PCT patent publication Nos. 91/17271, 91/18980, 91/19818, and 93/08278. Other systems for generating libraries of peptides have aspects of both in vitro chemical synthesis and recombinant methods. See, PCT Patent publication Nos. 92/05258, 92/14843, and 96/19256. See also, U.S. Patent Nos. 5,658,754; and 5,643,768. Peptide display libraries, vectors, and screening kits are commercially available from such suppliers as Invitrogen (Carlsbad, CA).

E. Polynucleotides Encoding a Protein Having a Subsequence from a Prototype Polypeptide and is Cross-Reactive to the Prototype Polypeptide

As indicated in (e), above, the present invention provides isolated nucleic acids comprising polynucleotides of the present invention, wherein the polynucleotides encode a protein having a subsequence of contiguous amino acids from a prototype polypeptide of the present invention such as are provided in (a), above. The length of contiguous amino acids from the prototype polypeptide is selected from the group of integers

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consisting of from at least 10 to the number of amino acids within the prototype sequence. Thus, for example, the polynucleotide can encode a polypeptide having a subsequence having at least 10, 15, 20, 25, 30, 35, 40, 45, or 50, contiguous amino acids from the prototype polypeptide. Further, the number of such subsequences encoded by a polynucleotide of the instant embodiment can be any integer selected from the group consisting of from 1 to 20, such as 2, 3, 4, or 5. The subsequences can be separated by any integer of nucleotides from 1 to the number of nucleotides in the sequence such as at least 5, 10, 15, 25, 50, 100, or 200 nucleotides.

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The proteins encoded by polynucleotides of this embodiment, when presented as an immunogen, elicit the production of polyclonal antibodies which specifically bind to a prototype polypeptide such as but not limited to, a polypeptide encoded by the polynucleotide of (a) or (b), above. Generally, however, a protein encoded by a polynucleotide of this embodiment does not bind to antisera raised against the prototype polypeptide when the antisera has been fully immunosorbed with the prototype polypeptide. Methods of making and assaying for antibody binding specificity/affinity are well known in the art. Exemplary immunoassay formats include ELISA, competitive immunoassays, radioimmunoassays, Western blots, indirect immunofluorescent assays and the like.

In a preferred assay method, fully immunosorbed and pooled antisera which is elicited to the prototype polypeptide can be used in a competitive binding assay to test the protein. The concentration of the prototype polypeptide required to inhibit 50% of the binding of the antisera to the prototype polypeptide is determined. If the amount of the protein required to inhibit binding is less than twice the amount of the prototype protein, then the protein is said to specifically bind to the antisera elicited to the immunogen. Accordingly, the proteins of the present invention embrace allelic variants, conservatively modified variants, and minor recombinant modifications to a prototype polypeptide.

A polynucleotide of the present invention optionally encodes a protein having a molecular weight as the non-glycosylated protein within 20% of the molecular weight of the full-length non-glycosylated polypeptides of the present invention. Molecular weight can be readily determined by SDS-PAGE under reducing conditions. Preferably, the molecular weight is within 15% of a full length polypeptide of the present invention, more preferably within 10% or 5%, and most preferably within 3%, 2%, or 1% of a full

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length polypeptide of the present invention. Molecular weight determination of a protein can be conveniently performed by SDS-PAGE under denaturing conditions.

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Optionally, the polynucleotides of this embodiment will encode a protein having a specific activity at least 50%, 60%, 80%, or 90% of the native, endogenous (i.e., nonisolated), full-length polypeptide of the present invention. Further, the proteins encoded by polynucleotides of this embodiment will optionally have a substantially similar affinity constant (K<sub>m</sub>) and/or catalytic activity (i.e., the microscopic rate constant, k<sub>cat</sub>) as the native endogenous, full-length protein. Those of skill in the art will recognize that k<sub>cat</sub>/K<sub>m</sub> value determines the specificity for competing substrates and is often referred to as the specificity constant. Proteins of this embodiment can have a k<sub>eat</sub>/K<sub>m</sub> value at least 10% of a non-isolated full-length polypeptide of the present invention as determined using the endogenous substrate of that polypeptide. Optionally, the  $k_{cat}/K_m$  value will be at least 20%, 30%, 40%, 50%, and most preferably at least 60%, 70%, 80%, 90%, or 95% the  $k_{cal}/K_m$  value of the non-isolated, full-length polypeptide of the present invention. Determination of  $k_{cat}$ ,  $K_m$ , and  $k_{cat}/K_m$  can be determined by any number of means well known to those of skill in the art. For example, the initial rates (i.e., the first 5% or less of the reaction) can be determined using rapid mixing and sampling techniques (e.g., continuous-flow, stopped-flow, or rapid quenching techniques), flash photolysis, or relaxation methods (e.g., temperature jumps) in conjunction with such exemplary methods of measuring as spectrophotometry, spectrofluorimetry, nuclear magnetic resonance, or radioactive procedures. Kinetic values are conveniently obtained using a Lineweaver-Burk or Eadie-Hofstee plot.

#### F. Polynucleotides Complementary to the Polynucleotides of (A)-(E)

As indicated in (f), above, the present invention provides isolated nucleic acids comprising polynucleotides complementary to the polynucleotides of paragraphs A-E, above. As those of skill in the art will recognize, complementary sequences base-pair throughout the entirety of their length with the polynucleotides of (A)-(E) (i.e., have 100% sequence identity over their entire length). Complementary bases associate through hydrogen bonding in double stranded nucleic acids. For example, the following base pairs are complementary: guanine and cytosine; adenine and thymine; and adenine and uracil.

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G. Polynucleotides Which are Subsequences of the Polynucleotides of (A)-(F)

As indicated in (g), above, the present invention provides isolated nucleic acids comprising polynucleotides which comprise at least 15 contiguous bases from the polynucleotides of (A) through (F) as discussed above. The length of the polynucleotide is given as an integer selected from the group consisting of from at least 15 to the length of the nucleic acid sequence from which the polynucleotide is a subsequence of. Thus, for example, polynucleotides of the present invention are inclusive of polynucleotides comprising at least 15, 20, 25, 30, 40, 50, 60, 75, or 100 contiguous nucleotides in length from the polynucleotides of (A)-(F). Optionally, the number of such subsequences encoded by a polynucleotide of the instant embodiment can be any integer selected from the group consisting of from 1 to 20, such as 2, 3, 4, or 5. The subsequences can be separated by any integer of nucleotides from 1 to the number of nucleotides in the sequence such as at least 5, 10, 15, 25, 50, 100, or 200 nucleotides.

The subsequences of the present invention can comprise structural characteristics of the sequence from which it is derived. Alternatively, the subsequences can lack certain structural characteristics of the larger sequence from which it is derived. For example, a subsequence from a polynucleotide encoding a polypeptide having at least one linear epitope in common with a prototype polypeptide sequence as provided in (a), above, may encode an epitope in common with the prototype sequence. Alternatively, the subsequence may not encode an epitope in common with the prototype sequence but can be used to isolate the larger sequence by, for example, nucleic acid hybridization with the sequence from which it's derived. Subsequences can be used to modulate or detect gene expression by introducing into the subsequences compounds which bind, intercalate, cleave and/or crosslink to nucleic acids. Exemplary compounds include acridine, psoralen, phenanthroline, naphthoquinone, daunomycin or chloroethylaminoaryl conjugates.

#### **Construction of Nucleic Acids**

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The isolated nucleic acids of the present invention can be made using (a) standard recombinant methods, (b) synthetic techniques, or combinations thereof. In some embodiments, the polynucleotides of the present invention will be cloned, amplified, or otherwise constructed from a monocot. In preferred embodiments the monocot is Zea mays.

The nucleic acids may conveniently comprise sequences in addition to a polynucleotide of the present invention. For example, a multi-cloning site comprising one or more endonuclease restriction sites may be inserted into the nucleic acid to aid in isolation of the polynucleotide. Also, translatable sequences may be inserted to aid in the isolation of the translated polynucleotide of the present invention. For example, a hexa-histidine marker sequence provides a convenient means to purify the proteins of the present invention. A polynucleotide of the present invention can be attached to a vector, adapter, or linker for cloning and/or expression of a polynucleotide of the present invention. Additional sequences may be added to such cloning and/or expression sequences to optimize their function in cloning and/or expression, to aid in isolation of the polynucleotide, or to improve the introduction of the polynucleotide into a cell. Typically, the length of a nucleic acid of the present invention less the length of its polynucleotide of the present invention is less than 20 kilobase pairs, often less than 15 kb, and frequently less than 10 kb. Use of cloning vectors, expression vectors, adapters, and linkers is well known and extensivley described in the art. For a description of various nucleic acids see, for example, Stratagene Cloning Systems, Catalogs 1995, 1996, 1997 (La Jolla, CA); and, Amersham Life Sciences, Inc, Catalog '97 (Arlington Heights, IL).

## 20 A. Recombinant Methods for Constructing Nucleic Acids

The isolated nucleic acid compositions of this invention, such as RNA, cDNA, genomic DNA, or a hybrid thereof, can be obtained from plant biological sources using any number of cloning methodologies known to those of skill in the art. In some embodiments, oligonucleotide probes which selectively hybridize, under stringent conditions, to the polynucleotides of the present invention are used to identify the desired sequence in a cDNA or genomic DNA library. While isolation of RNA, and construction of cDNA and genomic libraries is well known to those of ordinary skill in the art, the following highlights some of the methods employed.

## 30 A1. mRNA Isolation and Purification

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Total RNA from plant cells comprises such nucleic acids as mitochondrial RNA, chloroplastic RNA, rRNA, tRNA, hnRNA and mRNA. Total RNA preparation typically involves lysis of cells and removal of proteins, followed by precipitation of nucleic

acids. Extraction of total RNA from plant cells can be accomplished by a variety of means. Frequently, extraction buffers include a strong detergent such as SDS and an organic denaturant such as guanidinium isothiocyanate, guanidine hydrochloride or phenol. Following total RNA isolation, poly(A)<sup>+</sup> mRNA is typically purified from the remainder RNA using oligo(dT) cellulose. Exemplary total RNA and mRNA isolation protocols are described in *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997); and, *Current Protocols in Molecular Biology*, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995). Total RNA and mRNA isolation kits are commercially available from vendors such as Stratagene (La Jolla, CA), Clonetech (Palo Alto, CA), Pharmacia (Piscataway, NJ), and 5'-3' (Paoli, PA). See also, U.S. Patent Nos. 5,614,391; and, 5,459,253. The mRNA can be fractionated into populations with size ranges of about 0.5, 1.0, 1.5, 2.0, 2.5 or 3.0 kb. The cDNA synthesized for each of these fractions can be size selected to the same size range as its mRNA prior to vector insertion. This method helps eliminate truncated cDNA formed by incompletely reverse transcribed mRNA.

#### A2. Construction of a cDNA Library

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Construction of a cDNA library generally entails five steps. First, first strand cDNA synthesis is initiated from a poly(A)<sup>+</sup> mRNA template using a poly(dT) primer or random hexanucleotides. Second, the resultant RNA-DNA hybrid is converted into double stranded cDNA, typically by a combination of RNAse H and DNA polymerase I (or Klenow fragment). Third, the termini of the double stranded cDNA are ligated to adaptors. Ligation of the adaptors will produce cohesive ends for cloning. Fourth, size selection of the double stranded cDNA eliminates excess adaptors and primer fragments, and eliminates partial cDNA molecules due to degradation of mRNAs or the failure of reverse transcriptase to synthesize complete first strands. Fifth, the cDNAs are ligated into cloning vectors and packaged. cDNA synthesis protocols are well known to the skilled artisan and are described in such standard references as: *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997); and, *Current Protocols in Molecular Biology*, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995). cDNA synthesis kits are available from a variety of commercial vendors such as Stratagene or Pharmacia.

A number of cDNA synthesis protocols have been described which provide substantially pure full-length cDNA libraries. Substantially pure full-length cDNA libraries are constructed to comprise at least 90%, and more preferably at least 93% or 95% full-length inserts amongst clones containing inserts. The length of insert in such libraries can be from 0 to 8, 9, 10, 11, 12, 13, or more kilobase pairs. Vectors to accommodate inserts of these sizes are known in the art and available commercially. See, e.g., Stratagene's lambda ZAP Express (cDNA cloning vector with 0 to 12 kb cloning capacity).

An exemplary method of constructing a greater than 95% pure full-length cDNA library is described by Carninci *et al.*, *Genomics*, 37:327-336 (1996). In that protocol, the cap-structure of eukaryotic mRNA is chemically labeled with biotin. By using streptavidin-coated magnetic beads, only the full-length first-strand cDNA/mRNA hybrids are selectively recovered after RNase I treatment. The method provides a high yield library with an unbiased representation of the starting mRNA population. Other methods for producing full-length libraries are known in the art. See, e.g., Edery *et al.*, *Mol. Cell Biol.*,15(6):3363-3371 (1995); and, PCT Application WO 96/34981.

#### A3. Normalized or Subtracted cDNA Libraries

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A non-normalized cDNA library represents the mRNA population of the tissue it was made from. Since unique clones are out-numbered by clones derived from highly expressed genes their isolation can be laborious. Normalization of a cDNA library is the process of creating a library in which each clone is more equally represented.

A number of approaches to normalize cDNA libraries are known in the art. One approach is based on hybridization to genomic DNA. The frequency of each hybridized cDNA in the resulting normalized library would be proportional to that of each corresponding gene in the genomic DNA. Another approach is based on kinetics. If cDNA reannealing follows second-order kinetics, rarer species anneal less rapidly and the remaining single-stranded fraction of cDNA becomes progressively more normalized during the course of the hybridization. Specific loss of any species of cDNA, regardless of its abundance, does not occur at any Cot value. Construction of normalized libraries is described in Ko, *Nucl. Acids. Res.*, 18(19):5705-5711 (1990); Patanjali *et al.*, *Proc. Natl. Acad. U.S.A.*, 88:1943-1947 (1991); U.S. Patents 5,482,685, and 5,637,685. In an exemplary method described by Soares *et al.*, normalization resulted in reduction of

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the abundance of clones from a range of four orders of magnitude to a narrow range of only 1 order of magnitude. *Proc. Natl. Acad. Sci. USA*, 91:9228-9232 (1994).

Subtracted cDNA libraries are another means to increase the proportion of less abundant cDNA species. In this procedure, cDNA prepared from one pool of mRNA is depleted of sequences present in a second pool of mRNA by hybridization. The cDNA:mRNA hybrids are removed and the remaining un-hybridized cDNA pool is enriched for sequences unique to that pool. See, *Foote et al.* in, *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997); Kho and Zarbl, *Technique*, 3(2):58-63 (1991); Sive and St. John, *Nucl. Acids Res.*, 16(22):10937 (1988); *Current Protocols in Molecular Biology*, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995); and, Swaroop *et al.*, *Nucl. Acids Res.*, 19)8):1954 (1991). cDNA subtraction kits are commercially available. See, e.g., PCR-Select (Clontech).

#### 15 A4. Construction of a Genomic Library

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To construct genomic libraries, large segments of genomic DNA are generated by random fragmentation, e.g. using restriction endonucleases, and are ligated with vector DNA to form concatemers that can be packaged into the appropriate vector. Methodologies to accomplish these ends, and sequencing methods to verify the sequence 20 of nucleic acids are well known in the art. Examples of appropriate molecular biological techniques and instructions sufficient to direct persons of skill through many construction, cloning, and screening methodologies are found in Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor Laboratory Vols. 1-3 (1989), Methods in Enzymology, Vol. 152: Guide to Molecular Cloning 25 Techniques, Berger and Kimmel, Eds., San Diego: Academic Press, Inc. (1987), Current Protocols in Molecular Biology, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995); Plant Molecular Biology: A Laboratory Manual, Clark, Ed., Springer-Verlag, Berlin (1997). Kits for construction of genomic libraries are also commercially available.

# A5. Nucleic Acid Screening and Isolation Methods

The cDNA or genomic library can be screened using a probe based upon the sequence of a polynucleotide of the present invention such as those disclosed herein.

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Probes may be used to hybridize with genomic DNA or cDNA sequences to isolate homologous genes in the same or different plant species. Those of skill in the art will appreciate that various degrees of stringency of hybridization can be employed in the assay; and either the hybridization or the wash medium can be stringent. As the conditions for hybridization become more stringent, there must be a greater degree of complementarity between the probe and the target for duplex formation to occur. The degree of stringency can be controlled by temperature, ionic strength, pH and the presence of a partially denaturing solvent such as formamide. For example, the stringency of hybridization is conveniently varied by changing the polarity of the reactant solution through manipulation of the concentration of formamide within the range of 0% to 50%. The degree of complementarity (sequence identity) required for detectable binding will vary in accordance with the stringency of the hybridization medium and/or wash medium. The degree of complementarity will optimally be 100 percent; however, it should be understood that minor sequence variations in the probes and primers may be compensated for by reducing the stringency of the hybridization and/or wash medium.

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The nucleic acids of interest can also be amplified from nucleic acid samples using amplification techniques. For instance, polymerase chain reaction (PCR) technology can be used to amplify the sequences of polynucleotides of the present invention and related genes directly from genomic DNA or cDNA libraries. PCR and other *in vitro* amplification methods may also be useful, for example, to clone nucleic acid sequences that code for proteins to be expressed, to make nucleic acids to use as probes for detecting the presence of the desired mRNA in samples, for nucleic acid sequencing, or for other purposes. Examples of techniques sufficient to direct persons of skill through *in vitro* amplification methods are found in Berger, Sambrook, and Ausubel, as well as Mullis *et al.*, U.S. Patent No. 4,683,202 (1987); and, *PCR Protocols A Guide to Methods and Applications*, Innis *et al.*, Eds., Academic Press Inc., San Diego, CA (1990). Commercially available kits for genomic PCR amplification are known in the art. See, e.g., Advantage-GC Genomic PCR Kit (Clontech). The T4 gene 32 protein (Boehringer Mannheim) can be used to improve yield of long PCR products.

PCR-based screening methods have also been described. Wilfinger et al. describe a PCR-based method in which the longest cDNA is identified in the first step so that incomplete clones can be eliminated from study. *BioTechniques*, 22(3): 481-486 (1997). In that method, a primer pair is synthesized with one primer annealing to the 5'

end of the sense strand of the desired cDNA and the other primer to the vector. Clones are pooled to allow large-scale screening. By this procedure, the longest possible clone is identified amongst candidate clones. Further, the PCR product is used solely as a diagnostic for the presence of the desired cDNA and does not utilize the PCR product itself. Such methods are particularly effective in combination with a full-length cDNA construction methodology, above.

# B. Synthetic Methods for Constructing Nucleic Acids

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The isolated nucleic acids of the present invention can also be prepared by direct 10 chemical synthesis by methods such as the phosphotriester method of Narang et al., Meth. Enzymol. 68: 90-99 (1979); the phosphodiester method of Brown et al., Meth. Enzymol. 68: 109-151 (1979); the diethylphosphoramidite method of Beaucage et al., Tetra. Lett. 22: 1859-1862 (1981); the solid phase phosphoramidite triester method described by Beaucage and Caruthers, Tetra. Letts. 22(20): 1859-1862 (1981), e.g., using an automated synthesizer, e.g., as described in Needham-VanDevanter et al., 15 Nucleic Acids Res., 12: 6159-6168 (1984); and, the solid support method of U.S. Patent No. 4,458,066. Chemical synthesis generally produces a single stranded oligonucleotide. This may be converted into double stranded DNA by hybridization with a complementary sequence, or by polymerization with a DNA polymerase using the single strand as a template. One of skill will recognize that while chemical synthesis of 20 DNA is limited to sequences of about 100 bases, longer sequences may be obtained by the ligation of shorter sequences.

# **Recombinant Expression Cassettes**

The present invention further provides recombinant expression cassettes comprising a nucleic acid of the present invention. A nucleic acid sequence coding for the desired polynucleotide of the present invention, for example a cDNA or a genomic sequence encoding a full length polypeptide of the present invention, can be used to construct a recombinant expression cassette which can be introduced into the desired host cell. A recombinant expression cassette will typically comprise a polynucleotide of the present invention operably linked to transcriptional initiation regulatory sequences which will direct the transcription of the polynucleotide in the intended host cell, such as tissues of a transformed plant.

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For example, plant expression vectors may include (1) a cloned plant gene under the transcriptional control of 5' and 3' regulatory sequences and (2) a dominant selectable marker. Such plant expression vectors may also contain, if desired, a promoter regulatory region (e.g., one conferring inducible or constitutive, environmentally- or developmentally-regulated, or cell- or tissue-specific/selective expression), a transcription initiation start site, a ribosome binding site, an RNA processing signal, a transcription termination site, and/or a polyadenylation signal.

A plant promoter fragment can be employed which will direct expression of a polynucleotide of the present invention in all tissues of a regenerated plant. Such promoters are referred to herein as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include the cauliflower mosaic virus (CaMV) 35S transcription initiation region, the 1'- or 2'- promoter derived from T-DNA of *Agrobacterium tumefaciens*, the ubiquitin 1 promoter, the Smas promoter, the cinnamyl alcohol dehydrogenase promoter (U.S. Patent No. 5,683,439), the *Nos* promoter, the pEmu promoter, the rubisco promoter, the GRP1-8 promoter, the actin promoter, the F3.7 promoter, and other transcription initiation regions from various plant genes known to those of skill.

Alternatively, the plant promoter can direct expression of a polynucleotide of the present invention in a specific tissue or may be otherwise under more precise environmental or developmental control. Such promoters are referred to here as "inducible" promoters. Environmental conditions that may effect transcription by inducible promoters include pathogen attack, anaerobic conditions, or the presence of light. Examples of inducible promoters are the Adh1 promoter which is inducible by hypoxia or cold stress, the Hsp70 promoter which is inducible by heat stress, and the PPDK promoter which is inducible by light.

Examples of promoters under developmental control include promoters that initiate transcription only, or preferentially, in certain tissues, such as leaves, roots, fruit, seeds, or flowers. The operation of a promoter may also vary depending on its location in the genome. Thus, an inducible promoter may become fully or partially constitutive in certain locations.

Both heterologous and non-heterologous (i.e., endogenous) promoters can be employed to direct expression of the nucleic acids of the present invention. These

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promoters can also be used, for example, in recombinant expression cassettes to drive expression of antisense nucleic acids to reduce, increase, or alter concentration and/or composition of the proteins of the present invention in a desired tissue. Thus, in some embodiments, the nucleic acid construct will comprise a promoter functional in a plant cell, such as in *Zea mays*, operably linked to a polynucleotide of the present invention. Promoters useful in these embodiments include the endogenous promoters driving expression of a polypeptide of the present invention.

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In some embodiments, isolated nucleic acids which serve as promoter or enhancer elements can be introduced in the appropriate position (generally upstream) of a non-heterologous form of a polynucleotide of the present invention so as to up or down regulate expression of a polynucleotide of the present invention. For example, endogenous promoters can be altered *in vivo* by mutation, deletion, and/or substitution (see, Kmiec, U.S. Patent 5,565,350; Zarling *et al.*, PCT/US93/03868), or isolated promoters can be introduced into a plant cell in the proper orientation and distance from a gene of the present invention so as to control the expression of the gene. Gene expression can be modulated under conditions suitable for plant growth so as to alter the total concentration and/or alter the composition of the polypeptides of the present invention in plant cell. Thus, the present invention provides compositions, and methods for making, heterologous promoters and/or enhancers operably linked to a native, endogenous (i.e., non-heterologous) form of a polynucleotide of the present invention.

Methods for identifying promoters with a particular expression pattern, in terms of, e.g., tissue type, cell type, stage of development, and/or environmental conditions, are well known in the art. See, e.g., *The Maize Handbook*, Chapters 114-115, Freeling and Walbot, Eds., Springer, New York (1994); *Corn and Corn Improvement*, 3<sup>rd</sup> edition, Chapter 6, Sprague and Dudley, Eds., American Society of Agronomy, Madison, Wisconsin (1988). A typical step in promoter isolation methods is identification of gene products that are expressed with some degree of specificity in the target tissue. Amongst the range of methodologies are: differential hybridization to cDNA libraries; subtractive hybridization; differential display; differential 2-D protein gel electrophoresis; DNA probe arrays; and isolation of proteins known to be expressed with some specificity in the target tissue. Such methods are well known to those of skill in the art. Commercially available products for identifying promoters are known in the art such as Clontech's (Palo Alto, CA) Universal GenomeWalker Kit.

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For the protein-based methods, it is helpful to obtain the amino acid sequence for at least a portion of the identified protein, and then to use the protein sequence as the basis for preparing a nucleic acid that can be used as a probe to identify either genomic DNA directly, or preferably, to identify a cDNA clone from a library prepared from the target tissue. Once such a cDNA clone has been identified, that sequence can be used to identify the sequence at the 5' end of the transcript of the indicated gene. For differential hybridization, subtractive hybridization and differential display, the nucleic acid sequence identified as enriched in the target tissue is used to identify the sequence at the 5' end of the transcript of the indicated gene. Once such sequences are identified, starting either from protein sequences or nucleic acid sequences, any of these sequences identified as being from the gene transcript can be used to screen a genomic library prepared from the target organism. Methods for identifying and confirming the transcriptional start site are well known in the art.

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In the process of isolating promoters expressed under particular environmental conditions or stresses, or in specific tissues, or at particular developmental stages, a number of genes are identified that are expressed under the desired circumstances, in the desired tissue, or at the desired stage. Further analysis will reveal expression of each particular gene in one or more other tissues of the plant. One can identify a promoter with activity in the desired tissue or condition but that do not have activity in any other common tissue.

To identify the promoter sequence, the 5' portions of the clones described here are analyzed for sequences characteristic of promoter sequences. For instance, promoter sequence elements include the TATA box consensus sequence (TATAAT), which is usually an AT-rich stretch of 5-10 bp located approximately 20 to 40 base pairs upstream of the transcription start site. Identification of the TATA box is well known in the art. For example, one way to predict the location of this element is to identify the transcription start site using standard RNA-mapping techniques such as primer extension, S1 analysis, and/or RNase protection. To confirm the presence of the AT-rich sequence, a structure-function analysis can be performed involving mutagenesis of the putative region and quantification of the mutation's effect on expression of a linked downstream reporter gene. See, e.g., *The Maize Handbook*, Chapter 114, Freeling and Walbot, Eds., Springer, New York, (1994).

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In plants, further upstream from the TATA box, at positions -80 to -100, there is typically a promoter element (i.e., the CAAT box) with a series of adenines surrounding the trinucleotide G (or T) N G. J. Messing et al., in Genetic Engineering in Plants, Kosage, Meredith and Hollaender, Eds., pp. 221-227 1983. In maize, there is no well conserved CAAT box but there are several short, conserved protein-binding motifs upstream of the TATA box. These include motifs for the trans-acting transcription factors involved in light regulation, anaerobic induction, hormonal regulation, or anthocyanin biosynthesis, as appropriate for each gene.

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Once promoter and/or gene sequences are known, a region of suitable size is selected from the genomic DNA that is 5' to the transcriptional start, or the translational start site, and such sequences are then linked to a coding sequence. If the transcriptional start site is used as the point of fusion, any of a number of possible 5' untranslated regions can be used in between the transcriptional start site and the partial coding sequence. If the translational start site at the 3' end of the specific promoter is used, then it is linked directly to the methionine start codon of a coding sequence.

If polypeptide expression is desired, it is generally desirable to include a polyadenylation region at the 3'-end of a polynucleotide coding region. The polyadenylation region can be derived from the natural gene, from a variety of other plant genes, or from T-DNA. The 3' end sequence to be added can be derived from, for example, the nopaline synthase or octopine synthase genes, or alternatively from another plant gene, or less preferably from any other eukaryotic gene.

An intron sequence can be added to the 5' untranslated region or the coding sequence of the partial coding sequence to increase the amount of the mature message that accumulates in the cytosol. Inclusion of a spliceable intron in the transcription unit in both plant and animal expression constructs has been shown to increase gene expression at both the mRNA and protein levels up to 1000-fold. Buchman and Berg, *Mol. Cell Biol.* 8: 4395-4405 (1988); Callis *et al.*, *Genes Dev.* 1: 1183-1200 (1987). Such intron enhancement of gene expression is typically greatest when placed near the 5' end of the transcription unit. Use of maize introns Adh1-S intron 1, 2, and 6, the Bronze-1 intron are known in the art. See generally, *The Maize Handbook*, Chapter 116, Freeling and Walbot, Eds., Springer, New York (1994).

The vector comprising the sequences from a polynucleotide of the present invention will typically comprise a marker gene which confers a selectable phenotype on

plant cells. Usually, the selectable marker gene will encode antibiotic resistance, with suitable genes including genes coding for resistance to the antibiotic spectinomycin (e.g., the aada gene), the streptomycin phosphotransferase (SPT) gene coding for streptomycin resistance, the neomycin phosphotransferase (NPTII) gene encoding kanamycin or geneticin resistance, the hygromycin phosphotransferase (HPT) gene coding for hygromycin resistance, genes coding for resistance to herbicides which act to inhibit the action of acetolactate synthase (ALS), in particular the sulfonylurea-type herbicides (e.g., the acetolactate synthase (ALS) gene containing mutations leading to such resistance in particular the S4 and/or Hra mutations), genes coding for resistance to herbicides which act to inhibit action of glutamine synthase, such as phosphinothricin or basta (e.g., the *bar* gene), or other such genes known in the art. The *bar* gene encodes resistance to the herbicide basta, the *nptII* gene encodes resistance to the antibiotics kanamycin and geneticin, and the ALS gene encodes resistance to the herbicide chlorsulfuron.

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Typical vectors useful for expression of genes in higher plants are well known in the art and include vectors derived from the tumor-inducing (Ti) plasmid of Agrobacterium tumefaciens described by Rogers et al., Meth. In Enzymol., 153:253-277 (1987). These vectors are plant integrating vectors in that on transformation, the vectors integrate a portion of vector DNA into the genome of the host plant. Exemplary A.

20 tumefaciens vectors useful herein are plasmids pKYLX6 and pKYLX7 of Schardl et al., Gene, 61:1-11 (1987) and Berger et al., Proc. Natl. Acad. Sci. U.S.A., 86:8402-8406 (1989). Another useful vector herein is plasmid pBI101.2 that is available from Clontech Laboratories, Inc. (Palo Alto, CA).

A polynucleotide of the present invention can be expressed in either sense or antisense orientation as desired. It will be appreciated that control of gene expression in either sense or anti-sense orientation can have a direct impact on the observable plant characteristics. Antisense technology can be conveniently used to gene expression in plants. To accomplish this, a nucleic acid segment from the desired gene is cloned and operably linked to a promoter such that the anti-sense strand of RNA will be transcribed. The construct is then transformed into plants and the antisense strand of RNA is produced. In plant cells, it has been shown that antisense RNA inhibits gene expression by preventing the accumulation of mRNA which encodes the enzyme of interest, see,

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e.g., Sheehy et al., Proc. Nat'l. Acad. Sci. (USA) 85: 8805-8809 (1988); and Hiatt et al., U.S. Patent No. 4,801,340.

Another method of suppression is sense suppression. Introduction of nucleic acid configured in the sense orientation has been shown to be an effective means by which to block the transcription of target genes. For an example of the use of this method to modulate expression of endogenous genes see, Napoli *et al.*, *The Plant Cell* 2: 279-289 (1990) and U.S. Patent No. 5,034,323.

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Catalytic RNA molecules or ribozymes can also be used to inhibit expression of plant genes. It is possible to design ribozymes that specifically pair with virtually any target RNA and cleave the phosphodiester backbone at a specific location, thereby functionally inactivating the target RNA. In carrying out this cleavage, the ribozyme is not itself altered, and is thus capable of recycling and cleaving other molecules, making it a true enzyme. The inclusion of ribozyme sequences within antisense RNAs confers RNA-cleaving activity upon them, thereby increasing the activity of the constructs. The design and use of target RNA-specific ribozymes is described in Haseloff *et al.*, *Nature* 334: 585-591 (1988).

A variety of cross-linking agents, alkylating agents and radical generating species as pendant groups on polynucleotides of the present invention can be used to bind, label, detect, and/or cleave nucleic acids. For example, Vlassov, V. V., et al., Nucleic Acids Res (1986) 14:4065-4076, describe covalent bonding of a single-stranded DNA fragment with alkylating derivatives of nucleotides complementary to target sequences. A report of similar work by the same group is that by Knorre, D. G., et al., Biochimie (1985) 67:785-789. Iverson and Dervan also showed sequence-specific cleavage of singlestranded DNA mediated by incorporation of a modified nucleotide which was capable of activating cleavage (J Am Chem Soc (1987) 109:1241-1243). Meyer, R. B., et al., J Am Chem Soc (1989) 111:8517-8519, effect covalent crosslinking to a target nucleotide using an alkylating agent complementary to the single-stranded target nucleotide sequence. A photoactivated crosslinking to single-stranded oligonucleotides mediated by psoralen was disclosed by Lee, B. L., et al., Biochemistry (1988) 27:3197-3203. Use of crosslinking in triple-helix forming probes was also disclosed by Home, et al., J Am Chem Soc (1990) 112:2435-2437. Use of N4, N4-ethanocytosine as an alkylating agent to crosslink to single-stranded oligonucleotides has also been described by Webb and Matteucci, J Am Chem Soc (1986) 108:2764-2765; Nucleic Acids Res (1986) 14:7661-7674; Feteritz

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et al., J. Am. Chem. Soc. 113:4000 (1991). Various compounds to bind, detect, label, and/or cleave nucleic acids are known in the art. See, for example, U.S. Patent Nos. 5,543,507; 5,672,593; 5,484,908; 5,256,648; and, 5,681941.

#### 5 Proteins

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The isolated proteins of the present invention comprise a polypeptide having at least 10 amino acids encoded by any one of the polynucleotides of the present invention as discussed more fully, above, or polypeptides which are conservatively modified variants thereof. The proteins of the present invention or variants thereof can comprise any number of contiguous amino acid residues from a polypeptide of the present invention, wherein that number is selected from the group of integers consisting of from 10 to the number of residues in a full-length polypeptide of the present invention. Optionally, this subsequence of contiguous amino acids is at least 15, 20, 25, 30, 35, or 40 amino acids in length, often at least 50, 60, 70, 80, or 90 amino acids in length. Further, the number of such subsequences can be any integer selected from the group consisting of from 1 to 20, such as 2, 3, 4, or 5.

As those of skill will appreciate, the present invention includes catalytically active polypeptides of the present invention (i.e., enzymes). Catalytically active polypeptides have a specific activity of at least 20%, 30%, or 40%, and preferably at least 50%, 60%, or 70%, and most preferably at least 80%, 90%, or 95% that of the native (nonsynthetic), endogenous polypeptide. Further, the substrate specificity ( $k_{cat}/K_m$ ) is optionally substantially similar to the native (non-synthetic), endogenous polypeptide. Typically, the  $K_m$  will be at least 30%, 40%, or 50%, that of the native (non-synthetic), endogenous polypeptide; and more preferably at least 60%, 70%, 80%, or 90%. Methods of assaying and quantifying measures of enzymatic activity and substrate specificity ( $k_{cat}/K_m$ ), are well known to those of skill in the art.

Generally, the proteins of the present invention will, when presented as an immunogen, elicit production of an antibody specifically reactive to a polypeptide of the present invention. Further, the proteins of the present invention will not bind to antisera raised against a polypeptide of the present invention which has been fully immunosorbed with the same polypeptide. Immunoassays for determining binding are well known to those of skill in the art. A preferred immunoassay is a competitive immunoassay as discussed, *infra*. Thus, the proteins of the present invention can be employed as

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immunogens for constructing antibodies immunoreactive to a protein of the present invention for such exemplary utilities as immunoassays or protein purification techniques.

#### 5 Expression of Proteins in Host Cells

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Using the nucleic acids of the present invention, one may express a protein of the present invention in a recombinantly engineered cell such as bacteria, yeast, insect, mammalian, or preferably plant cells. The cells produce the protein in a non-natural condition (e.g., in quantity, composition, location, and/or time), because they have been genetically altered through human intervention to do so.

It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of a nucleic acid encoding a protein of the present invention. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of isolated nucleic acids encoding a protein of the present invention will typically be achieved by operably linking, for example, the DNA or cDNA to a promoter (which is either constitutive or inducible), followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding a protein of the present invention. To obtain high level expression of a cloned gene, it is desirable to construct expression vectors which contain, at the minimum, a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator. One of skill would recognize that modifications can be made to a protein of the present invention without diminishing its biological activity. Some modifications may be made to facilitate the cloning, expression, or incorporation of the targeting molecule into a fusion protein. Such modifications are well known to those of skill in the art and include, for example, a methionine added at the amino terminus to provide an initiation site, or additional amino acids (e.g., poly His) placed on either terminus to create conveniently located purification sequences. Restriction sites or termination codons can also be introduced.

#### A. Expression in Prokaryotes

Prokaryotic cells may be used as hosts for expression. Prokaryotes most frequently are represented by various strains of *E. coli*; however, other microbial strains may also be used. Commonly used prokaryotic control sequences which are defined herein to include promoters for transcription initiation, optionally with an operator, along with ribosome binding site sequences, include such commonly used promoters as the beta lactamase (penicillinase) and lactose (lac) promoter systems (Chang et al., Nature 198:1056 (1977)), the tryptophan (trp) promoter system (Goeddel et al., Nucleic Acids Res. 8:4057 (1980)) and the lambda derived P L promoter and N-gene ribosome binding site (Shimatake *et al.*, Nature 292:128 (1981)). The inclusion of selection markers in DNA vectors transfected in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol.

The vector is selected to allow introduction into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA. Expression systems for expressing a protein of the present invention are available using Bacillus sp. and Salmonella (Palva, et al., Gene 22: 229-235 (1983); Mosbach, et al., Nature 302: 543-545 (1983)).

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## B. Expression in Eukaryotes

A variety of eukaryotic expression systems such as yeast, insect cell lines, plant and mammalian cells, are known to those of skill in the art. As explained briefly below, a of the present invention can be expressed in these eukaryotic systems. In some embodiments, transformed/transfected plant cells, as discussed *infra*, are employed as expression systems for production of the proteins of the instant invention.

Synthesis of heterologous proteins in yeast is well known. Sherman, F., et al., Methods in Yeast Genetics, Cold Spring Harbor Laboratory (1982) is a well recognized work describing the various methods available to produce the protein in yeast. Two widely utilized yeast for production of eukaryotic proteins are Saccharomyces cerevisiae and Pichia pastoris. Vectors, strains, and protocols for expression in Saccharomyces and Pichia are known in the art and available from commercial suppliers (e.g., Invitrogen). Suitable vectors usually have expression control sequences, such as

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promoters, including 3-phosphoglycerate kinase or alcohol oxidase, and an origin of replication, termination sequences and the like as desired.

A protein of the present invention, once expressed, can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassay of other standard immunoassay techniques.

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The sequences encoding proteins of the present invention can also be ligated to various expression vectors for use in transfecting cell cultures of, for instance, mammalian, insect, or plant origin. Illustrative of cell cultures useful for the production of the peptides are mammalian cells. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. A number of suitable host cell lines capable of expressing intact proteins have been developed in the art, and include the HEK293, BHK21, and CHO cell lines. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter (e.g., the CMV promoter, a HSV tk promoter or pgk (phosphoglycerate kinase) promoter), an enhancer (Queen et al., Immunol. Rev. 89: 49 (1986)), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites (e.g., an SV40 large T Ag poly A addition site), and transcriptional terminator sequences. Other animal cells useful for production of proteins of the present invention are available, for instance, from the American Type Culture Collection Catalogue of Cell Lines and Hybridomas (7th edition, 1992).

Appropriate vectors for expressing proteins of the present invention in insect cells are usually derived from the SF9 baculovirus. Suitable insect cell lines include mosquito larvae, silkworm, armyworm, moth and *Drosophila* cell lines such as a Schneider cell line (See Schneider, *J. Embryol. Exp. Morphol.* 27: 353-365 (1987).

As with yeast, when higher animal or plant host cells are employed, polyadenlyation or transcription terminator sequences are typically incorporated into the vector. An example of a terminator sequence is the polyadenlyation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VP1 intron from SV40 (Sprague, et al., J. Virol. 45: 773-781 (1983)). Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus type-vectors. Saveria-Campo, M., Bovine Papilloma Virus DNA a

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Eukaryotic Cloning Vector in *DNA Cloning Vol. II a Practical Approach*, D.M. Glover, Ed., IRL Press, Arlington, Virginia pp. 213-238 (1985).

# **Transfection/Transformation of Cells**

The method of transformation/transfection is not critical to the instant invention; various methods of transformation or transfection are currently available. As newer methods are available to transform crops or other host cells they may be directly applied. Accordingly, a wide variety of methods have been developed to insert a DNA sequence into the genome of a host cell to obtain the transcription and/or translation of the sequence to effect phenotypic changes in the organism. Thus, any method which provides for efficient transformation/transfection may be employed.

#### A. Plant Transformation

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A DNA sequence coding for the desired polynucleotide of the present invention, for example a cDNA or a genomic sequence encoding a full length protein, will be used to construct a recombinant expression cassette which can be introduced into the desired plant.

Isolated nucleic acid acids of the present invention can be introduced into plants according techniques known in the art. Generally, recombinant expression cassettes as described above and suitable for transformation of plant cells are prepared. Techniques for transforming a wide variety of higher plant species are well known and described in the technical, scientific, and patent literature. See, for example, Weising *et al.*, *Ann. Rev. Genet.* 22: 421-477 (1988). For example, the DNA construct may be introduced directly into the genomic DNA of the plant cell using techniques such as electroporation, PEG poration, particle bombardment, silicon fiber delivery, or microinjection of plant cell protoplasts or embryogenic callus. See e.g., Tomes, *et al.*, Direct DNA Transfer into Intact Plant Cells via Microprojectile Bombardment, pp. 197-213 in Plant Cell, Tissue and Organ Culture, Fundamental Methods, (eds. O.L. Gamborg and G.C. Phillips, Springer-Verlag Berlin Heidelberg New York, 1995). Alternatively, the DNA constructs may be combined with suitable T-DNA flanking regions and introduced into a conventional *Agrobacterium tumefaciens* host vector. The virulence functions of the *Agrobacterium tumefaciens* host will direct the insertion of the construct and adjacent

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marker into the plant cell DNA when the cell is infected by the bacteria. See, U.S. Patent No. 5,591,616.

The introduction of DNA constructs using polyethylene glycol precipitation is described in Paszkowski *et al.*, *Embo J.* 3: 2717-2722 (1984). Electroporation techniques are described in Fromm *et al.*, *Proc. Natl. Acad. Sci.* 82: 5824 (1985). Ballistic transformation techniques are described in Klein *et al.*, *Nature* 327: 70-73 (1987).

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Agrobacterium tumefaciens-meditated transformation techniques are well described in the scientific literature. See, for example Horsch et al., Science 233: 496-498 (1984), and Fraley et al., Proc. Natl. Acad. Sci. 80: 4803 (1983). Although Agrobacterium is useful primarily in dicots, certain monocots can be transformed by Agrobacterium. For instance, Agrobacterium transformation of maize is described in U.S. Patent No. 5,550,318.

Other methods of transfection or transformation include (1) Agrobacterium

15 rhizogenes-mediated transformation (see, e.g., Lichtenstein and Fuller In: Genetic Engineering, vol. 6, PWJ Rigby, Ed., London, Academic Press, 1987; and Lichtenstein, C. P., and Draper, J., In: DNA Cloning, Vol. II, D. M. Glover, Ed., Oxford, IRI Press, 1985), Application PCT/US87/02512 (WO 88/02405 published Apr. 7, 1988) describes the use of A. rhizogenes strain A4 and its Ri plasmid along with A. tumefaciens vectors pARC8 or pARC16 (2) liposome-mediated DNA uptake (see, e.g., Freeman et al., Plant Cell Physiol. 25: 1353, 1984), (3) the vortexing method (see, e.g., Kindle, Proc. Natl. Acad. Sci., USA 87: 1228, (1990).

DNA can also be introduced into plants by direct DNA transfer into pollen as described by Zhou *et al.*, Methods in Enzymology, 101:433 (1983); D. Hess, Intern Rev. Cytol., 107:367 (1987); Luo *et al.*, Plane Mol. Biol. Reporter, 6:165 (1988). Expression of polypeptide coding genes can be obtained by injection of the DNA into reproductive organs of a plant as described by Pena *et al.*, Nature, 325.:274 (1987). DNA can also be injected directly into the cells of immature embryos and the rehydration of desiccated embryos as described by Neuhaus *et al.*, Theor. Appl. Genet., 75:30 (1987); and Benbrook *et al.*, in Proceedings Bio Expo 1986, Butterworth, Stoneham, Mass., pp. 27-54 (1986). A variety of plant viruses that can be employed as vectors are known in the art and include cauliflower mosaic virus (CaMV), geminivirus, brome mosaic virus, and tobacco mosaic virus.

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#### B. Transfection of Prokaryotes, Lower Eukaryotes, and Animal Cells

Animal and lower eukaryotic (e.g., yeast) host cells are competent or rendered competent for transfection by various means. There are several well-known methods of introducing DNA into animal cells. These include: calcium phosphate precipitation, fusion of the recipient cells with bacterial protoplasts containing the DNA, treatment of the recipient cells with liposomes containing the DNA, DEAE dextran, electroporation, biolistics, and micro-injection of the DNA directly into the cells. The transfected cells are cultured by means well known in the art. Kuchler, R.J., *Biochemical Methods in Cell Culture and Virology*, Dowden, Hutchinson and Ross, Inc. (1977).

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#### **Synthesis of Proteins**

The proteins of the present invention can be constructed using non-cellular synthetic methods. Solid phase synthesis of proteins of less than about 50 amino acids in length may be accomplished by attaching the C-terminal amino acid of the sequence to an insoluble support followed by sequential addition of the remaining amino acids in the sequence. Techniques for solid phase synthesis are described by Barany and Merrifield, Solid-Phase Peptide Synthesis, pp. 3-284 in *The Peptides: Analysis, Synthesis, Biology.* Vol. 2: Special Methods in Peptide Synthesis, Part A.; Merrifield, et al., J. Am. Chem. Soc. 85: 2149-2156 (1963), and Stewart et al., Solid Phase Peptide Synthesis, 2nd ed., Pierce Chem. Co., Rockford, Ill. (1984). Proteins of greater length may be synthesized by condensation of the amino and carboxy termini of shorter fragments. Methods of forming peptide bonds by activation of a carboxy terminal end (e.g., by the use of the coupling reagent N,N'-dicycylohexylcarbodiimide)) is known to those of skill.

# 25 **Purification of Proteins**

The proteins of the present invention may be purified by standard techniques well known to those of skill in the art. Recombinantly produced proteins of the present invention can be directly expressed or expressed as a fusion protein. The recombinant protein is purified by a combination of cell lysis (e.g., sonication, French press) and affinity chromatography. For fusion products, subsequent digestion of the fusion protein with an appropriate proteolytic enzyme releases the desired recombinant protein.

The proteins of this invention, recombinant or synthetic, may be purified to substantial purity by standard techniques well known in the art, including detergent

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solubilization, selective precipitation with such substances as ammonium sulfate, column chromatography, immunopurification methods, and others. See, for instance, R. Scopes, Protein Purification: Principles and Practice, Springer-Verlag: New York (1982); Deutscher, Guide to Protein Purification, Academic Press (1990). For example, antibodies may be raised to the proteins as described herein. Purification from E. coli can be achieved following procedures described in U.S. Patent No. 4,511,503. The protein may then be isolated from cells expressing the protein and further purified by standard protein chemistry techniques as described herein. Detection of the expressed protein is achieved by methods known in the art and include, for example, radioimmunoassays, Western blotting techniques or immunoprecipitation.

## **Transgenic Plant Regeneration**

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Transformed plant cells which are derived by any of the above transformation techniques can be cultured to regenerate a whole plant which possesses the transformed genotype. Such regeneration techniques often rely on manipulation of certain phytohormones in a tissue culture growth medium, typically relying on a biocide and/or herbicide marker which has been introduced together with a polynucleotide of the present invention. For transformation and regeneration of maize see, Gordon-Kamm et al., The Plant Cell, 2:603-618 (1990).

20 Plants cells transformed with a plant expression vector can be regenerated, e.g., from single cells, callus tissue or leaf discs according to standard plant tissue culture techniques. It is well known in the art that various cells, tissues, and organs from almost any plant can be successfully cultured to regenerate an entire plant. Plant regeneration from cultured protoplasts is described in Evans et al., Protoplasts Isolation and Culture, Handbook of Plant Cell Culture, Macmillilan Publishing Company, New York, pp. 124-176 (1983); and Binding, Regeneration of Plants, Plant Protoplasts, CRC Press, Boca Raton, pp. 21-73 (1985).

The regeneration of plants containing the foreign gene introduced by Agrobacterium from leaf explants can be achieved as described by Horsch et al., Science, 227:1229-1231 (1985). In this procedure, transformants are grown in the presence of a selection agent and in a medium that induces the regeneration of shoots in the plant species being transformed as described by Fraley et al., Proc. Natl. Acad. Sci. U.S.A., 80:4803 (1983). This procedure typically produces shoots within two to four

weeks and these transformant shoots are then transferred to an appropriate root-inducing medium containing the selective agent and an antibiotic to prevent bacterial growth.

Transgenic plants of the present invention may be fertile or sterile.

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Regeneration can also be obtained from plant callus, explants, organs, or parts thereof. Such regeneration techniques are described generally in Klee *et al.*, *Ann. Rev. of Plant Phys.* 38: 467-486 (1987). The regeneration of plants from either single plant protoplasts or various explants is well known in the art. See, for example, Methods for Plant Molecular Biology, A. Weissbach and H. Weissbach, eds., Academic Press, Inc., San Diego, Calif. (1988). This regeneration and growth process includes the steps of selection of transformant cells and shoots, rooting the transformant shoots and growth of the plantlets in soil. For maize cell culture and regeneration see generally, *The Maize Handbook*, Freeling and Walbot, Eds., Springer, New York (1994); *Corn and Corn Improvement*, 3<sup>rd</sup> edition, Sprague and Dudley Eds., American Society of Agronomy, Madison, Wisconsin (1988).

One of skill will recognize that after the recombinant expression cassette is stably incorporated in transgenic plants and confirmed to be operable, it can be introduced into other plants by sexual crossing. Any of a number of standard breeding techniques can be used, depending upon the species to be crossed.

In vegetatively propagated crops, mature transgenic plants can be propagated by the taking of cuttings or by tissue culture techniques to produce multiple identical plants. Selection of desirable transgenics is made and new varieties are obtained and propagated vegetatively for commercial use. In seed propagated crops, mature transgenic plants can be self-crossed to produce a homozygous inbred plant. The inbred plant produces seed containing the newly introduced heterologous nucleic acid. These seeds can be grown to produce plants that would produce the selected phenotype.

Parts obtained from the regenerated plant, such as flowers, seeds, leaves, branches, fruit, and the like are included in the invention, provided that these parts comprise cells comprising the isolated nucleic acid of the present invention. Progeny and variants, and mutants of the regenerated plants are also included within the scope of the invention, provided that these parts comprise the introduced nucleic acid sequences.

Transgenic plants expressing the selectable marker can be screened for transmission of the nucleic acid of the present invention by, for example, standard immunoblot and DNA detection techniques. Transgenic lines are also typically evaluated

on levels of expression of the heterologous nucleic acid. Expression at the RNA level can be determined initially to identify and quantitate expression-positive plants. Standard techniques for RNA analysis can be employed and include PCR amplification assays using oligonucleotide primers designed to amplify only the heterologous RNA templates and solution hybridization assays using heterologous nucleic acid-specific probes. The RNA-positive plants can then analyzed for protein expression by Western immunoblot analysis using the specifically reactive antibodies of the present invention. In addition, in situ hybridization and immunocytochemistry according to standard protocols can be done using heterologous nucleic acid specific polynucleotide probes and antibodies, respectively, to localize sites of expression within transgenic tissue. Generally, a number of transgenic lines are usually screened for the incorporated nucleic acid to identify and select plants with the most appropriate expression profiles.

A preferred embodiment is a transgenic plant that is homozygous for the added heterologous nucleic acid; i.e., a transgenic plant that contains two added nucleic acid sequences, one gene at the same locus on each chromosome of a chromosome pair. A homozygous transgenic plant can be obtained by sexually mating (selfing) a heterozygous transgenic plant that contains a single added heterologous nucleic acid, germinating some of the seed produced and analyzing the resulting plants produced for altered expression of a polynucleotide of the present invention relative to a control plant (i.e., native, non-transgenic). Back-crossing to a parental plant and out-crossing with a non-transgenic plant are also contemplated.

# Modulating Polypeptide Levels and/or Composition

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The present invention further provides a method for modulating (i.e., increasing or decreasing) the concentration or composition of the polypeptides of the present invention in a plant or part thereof. Modulation can be effected by increasing or decreasing the concentration and/or the composition (i.e., the ratio of the polypeptides of the present invention) in a plant. The method comprises transforming a plant cell with a recombinant expression cassette comprising a polynucleotide of the present invention as described above to obtain a transformed plant cell, growing the transformed plant cell under plant forming conditions, and inducing expression of a polynucleotide of the present invention in the plant for a time sufficient to modulate concentration and/or composition in the plant or plant part.

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In some embodiments, the content and/or composition of polypeptides of the present invention in a plant may be modulated by altering, in vivo or in vitro, the promoter of a non-isolated gene of the present invention to up- or down-regulate gene expression. In some embodiments, the coding regions of native genes of the present invention can be altered via substitution, addition, insertion, or deletion to decrease activity of the encoded enzyme. See, e.g., Kmiec, U.S. Patent 5,565,350; Zarling et al., PCT/US93/03868. And in some embodiments, an isolated nucleic acid (e.g., a vector) comprising a promoter sequence is transfected into a plant cell. Subsequently, a plant cell comprising the promoter operably linked to a polynucleotide of the present invention is selected for by means known to those of skill in the art such as, but not limited to, Southern blot, DNA sequencing, or PCR analysis using primers specific to the promoter and to the gene and detecting amplicons produced therefrom. A plant or plant part altered or modified by the foregoing embodiments is grown under plant forming conditions for a time sufficient to modulate the concentration and/or composition of polypeptides of the present invention in the plant. Plant forming conditions are well known in the art and discussed briefly, above.

In general, concentration or composition is increased or decreased by at least 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, or 90% relative to a native control plant, plant part, or cell lacking the aforementioned recombinant expression cassette. Modulation in the present invention may occur during and/or subsequent to growth of the plant to the desired stage of development. Modulating nucleic acid expression temporally and/or in particular tissues can be controlled by employing the appropriate promoter operably linked to a polynucleotide of the present invention in, for example, sense or antisense orientation as discussed in greater detail, *above*. Induction of expression of a polynucleotide of the present invention can also be controlled by exogenous administration of an effective amount of inducing compound. Inducible promoters and inducing compounds which activate expression from these promoters are well known in the art. In preferred embodiments, the polypeptides of the present invention are modulated in monocots, particularly maize.

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The present invention provides a method of genotyping a plant comprising a polynucleotide of the present invention. Preferably, the plant is a monocot, such as maize or sorghum. Genotyping provides a means of distinguishing homologs of a chromosome pair and can be used to differentiate segregants in a plant population.

5 Molecular marker methods can be used for phylogenetic studies, characterizing genetic relationships among crop varieties, identifying crosses or somatic hybrids, localizing chromosomal segments affecting monogenic traits, map based cloning, and the study of quantitative inheritance. See, e.g., *Plant Molecular Biology: A Laboratory Manual*, Chapter 7, Clark, Ed., Springer-Verlag, Berlin (1997). For molecular marker methods, see generally, The DNA Revolution by Andrew H. Paterson 1996 (Chapter 2) in: Genome Mapping in Plants (ed. Andrew H. Paterson) by Academic Press/R. G. Landis Company, Austin, Texas, pp.7-21.

The particular method of genotyping in the present invention may employ any number of molecular marker analytic techniques such as, but not limited to, restriction fragment length polymorphisms (RFLPs). RFLPs are the product of allelic differences between DNA restriction fragments caused by nucleotide sequence variability. As is well known to those of skill in the art, RFLPs are typically detected by extraction of genomic DNA and digestion with a restriction enzyme. Generally, the resulting fragments are separated according to size and hybridized with a probe; single copy probes are preferred. Restriction fragments from homologous chromosomes are revealed. Differences in fragment size among alleles represent an RFLP. Thus, the present invention further provides a means to follow segregation of a gene or nucleic acid of the present invention as well as chromosomal sequences genetically linked to these genes or nucleic acids using such techniques as RFLP analysis. Linked chromosomal sequences are within 50 centiMorgans (cM), often within 40 or 30 cM, preferably within 20 or 10 cM, more preferably within 5, 3, 2, or 1 cM of a gene of the present invention.

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In the present invention, the nucleic acid probes employed for molecular marker mapping of plant nuclear genomes selectively hybridize, under selective hybridization conditions, to a gene encoding a polynucleotide of the present invention. In preferred embodiments, the probes are selected from polynucleotides of the present invention. Typically, these probes are cDNA probes or *Pst I* genomic clones. The length of the probes is discussed in greater detail, above, but are typically at least 15 bases in length,

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more preferably at least 20, 25, 30, 35, 40, or 50 bases in length. Generally, however, the probes are less than about 1 kilobase in length. Preferably, the probes are single copy probes that hybridize to a unique locus in a haploid chromosome complement. Some exemplary restriction enzymes employed in RFLP mapping are *EcoRI*, *EcoRv*, and *SstI*. As used herein the term "restriction enzyme" includes reference to a composition that recognizes and, alone or in conjunction with another composition, cleaves at a specific nucleotide sequence.

The method of detecting an RFLP comprises the steps of (a) digesting genomic DNA of a plant with a restriction enzyme; (b) hybridizing a nucleic acid probe, under selective hybridization conditions, to a sequence of a polynucleotide of the present of said genomic DNA; (c) detecting therefrom a RFLP. Other methods of differentiating polymorphic (allelic) variants of polynucleotides of the present invention can be had by utilizing molecular marker techniques well known to those of skill in the art including such techniques as: 1) single stranded conformation analysis (SSCP); 2) denaturing gradient gel electrophoresis (DGGE); 3) RNase protection assays; 4) allele-specific oligonucleotides (ASOs); 5) the use of proteins which recognize nucleotide mismatches, such as the E. coli mutS protein; and 6) allele-specific PCR. Other approaches based on the detection of mismatches between the two complementary DNA strands include clamped denaturing gel electrophoresis (CDGE); heteroduplex analysis (HA); and chemical mismatch cleavage (CMC). Exemplary polymorphic variants are provided in Table I, above. Thus, the present invention further provides a method of genotyping comprising the steps of contacting, under stringent hybridization conditions, a sample suspected of comprising a polynucleotide of the present invention with a nucleic acid probe. Generally, the sample is a plant sample; preferably, a sample suspected of comprising a maize polynucleotide of the present invention (e.g., gene, mRNA). The nucleic acid probe selectively hybridizes, under stringent conditions, to a subsequence of a polynucleotide of the present invention comprising a polymorphic marker. Selective hybridization of the nucleic acid probe to the polymorphic marker nucleic acid sequence yields a hybridization complex. Detection of the hybridization complex indicates the presence of that polymorphic marker in the sample. In preferred embodiments, the nucleic acid probe comprises a polynucleotide of the present invention.

## **UTR's and Codon Preference**

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In general, translational efficiency has been found to be regulated by specific sequence elements in the 5' non-coding or untranslated region (5' UTR) of the RNA. Positive sequence motifs include translational initiation consensus sequences (Kozak, Nucleic Acids Res. 15:8125 (1987)) and the 7-methylguanosine cap structure (Drummond et al., Nucleic Acids Res. 13:7375 (1985)). Negative elements include stable intramolecular 5' UTR stem-loop structures (Muesing et al., Cell 48:691 (1987)) and AUG sequences or short open reading frames preceded by an appropriate AUG in the 5' UTR (Kozak, above, Rao et al., Mol. and Cell. Biol. 8:284 (1988)). Accordingly, the present invention provides 5' and/or 3' UTR regions for modulation of translation of heterologous coding sequences.

Further, the polypeptide-encoding segments of the polynucleotides of the present invention can be modified to alter codon usage. Altered codon usage can be employed to alter translational efficiency and/or to optimize the coding sequence for expression in a desired host or to optimize the codon usage in a heterologous sequence for expression in maize. Codon usage in the coding regions of the polynucleotides of the present invention 15 can be analyzed statistically using commercially available software packages such as "Codon Preference" available from the University of Wisconsin Genetics Computer Group (see Devereaux et al., Nucleic Acids Res. 12: 387-395 (1984)) or MacVector 4.1 (Eastman Kodak Co., New Haven, Conn.). Thus, the present invention provides a 20 codon usage frequency characteristic of the coding region of at least one of the polynucleotides of the present invention. The number of polynucleotides that can be used to determine a codon usage frequency can be any integer from 1 to the number of polynucleotides of the present invention as provided herein. Optionally, the polynucleotides will be full-length sequences. An exemplary number of sequences for 25 statistical analysis can be at least 1, 5, 10, 20, 50, or 100.

# **Sequence Shuffling**

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The present invention provides methods for sequence shuffling using polynucleotides of the present invention, and compositions resulting therefrom.

Sequence shuffling is described in PCT publication No. 96/19256. See also, Zhang, J.-H., et al. Proc. Natl. Acad. Sci. USA 94:4504-4509 (1997). Generally, sequence shuffling provides a means for generating libraries of polynucleotides having a desired characteristic which can be selected or screened for. Libraries of recombinant

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polynucleotides are generated from a population of related sequence polynucleotides which comprise sequence regions which have substantial sequence identity and can be homologously recombined in vitro or in vivo. The population of sequence-recombined polynucleotides comprises a subpopulation of polynucleotides which possess desired or advantageous characteristics and which can be selected by a suitable selection or screening method. The characteristics can be any property or attribute capable of being selected for or detected in a screening system, and may include properties of: an encoded protein, a transcriptional element, a sequence controlling transcription, RNA processing. RNA stability, chromatin conformation, translation, or other expression property of a gene or transgene, a replicative element, a protein-binding element, or the like, such as any feature which confers a selectable or detectable property. In some embodiments, the selected characteristic will be a decreased K<sub>m</sub> and/or increased K<sub>cu</sub> over the wild-type protein as provided herein. In other embodiments, a protein or polynculeotide generated from sequence shuffling will have a ligand binding affinity greater than the non-shuffled wild-type polynucleotide. The increase in such properties can be at least 110%, 120%, 130%, 140% or at least 150% of the wild-type value.

#### Generic and Consensus Sequences

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Polynuclotides and polypeptides of the present invention further include those having: (a) a generic sequence of at least two homologous polynucleotides or polypeptides, respectively, of the present invention; and, (b) a consensus sequence of at least three homologous polynucleotides or polypeptides, respectively, of the present invention. The generic sequence of the present invention comprises each species of polypeptide or polynucleotide embraced by the generic polypeptide or polynucleotide, sequence, respectively. The individual species encompassed by a polynucleotide having an amino acid or nucleic acid consensus sequence can be used to generate antibodies or produce nucleic acid probes or primers to screen for homologs in other species, genera, families, orders, classes, phyla, or kingdoms. For example, a polynucleotide having a consensus sequences from a gene family of *Zea mays* can be used to generate antibody or nucleic acid probes or primers to other *Gramineae* species such as wheat, rice, or sorghum. Alternatively, a polynucleotide having a consensus sequence generated from orthologous genes can be used to identify or isolate orthologs of other taxa. Typically, a polynucleotide having a consensus sequence will be at least 9, 10, 15, 20, 25, 30, or 40

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amino acids in length, or 20, 30, 40, 50, 100, or 150 nucleotides in length. As those of skill in the art are aware, a conservative amino acid substitution can be used for amino acids which differ amongst aligned sequence but are from the same conservative substitution group as discussed above. Optionally, no more than 1 or 2 conservative amino acids are substituted for each 10 amino acid length of consensus sequence.

Similar sequences used for generation of a consensus or generic sequence include any number and combination of allelic variants of the same gene, orthologous, or paralogous sequences as provided herein. Optionally, similar sequences used in generating a consensus or generic sequence are identified using the BLAST algorithm's smallest sum probability (P(N)). Various suppliers of sequence-analysis software are listed in chapter 7 of Current Protocols in Molecular Biology, F.M. Ausubel et al., Eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc. (Supplement 30). A polynucleotide sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.1, more preferably less than about 0.01, or 0.001, and most preferably less than about 0.0001, or 0.00001. Similar polynucleotides can be aligned and a consensus or generic sequence generated using multiple sequence alignment software available from a number of commercial suppliers such as the Genetics Computer Group's (Madison, WI) PILEUP software, Vector NTI's (North Bethesda, MD) ALIGNX, or Genecode's (Ann Arbor, MI) SEQUENCHER. Conveniently, default parameters of such software can be used to generate consensus or generic sequences.

#### **Detection of Nucleic Acids**

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The present invention further provides methods for detecting a polynucleotide of the present invention in a nucleic acid sample suspected of comprising a polynucleotide of the present invention, such as a plant cell lysate, particularly a lysate of corn. In some embodiments, a gene of the present invention or portion thereof can be amplified prior to the step of contacting the nucleic acid sample with a polynucleotide of the present invention. The nucleic acid sample is contacted with the polynucleotide to form a hybridization complex. The polynucleotide hybridizes under stringent conditions to a gene encoding a polypeptide of the present invention. Formation of the hybridization complex is used to detect a gene encoding a polypeptide of the present invention in the

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nucleic acid sample. Those of skill will appreciate that an isolated nucleic acid comprising a polynucleotide of the present invention should lack cross-hybridizing sequences in common with non-target genes that would yield a false positive result.

Detection of the hybridization complex can be achieved using any number of well known methods. For example, the nucleic acid sample, or a portion thereof, may be assayed by hybridization formats including but not limited to, solution phase, solid phase, mixed phase, or in situ hybridization assays. Briefly, in solution (or liquid) phase hybridizations, both the target nucleic acid and the probe or primer are free to interact in the reaction mixture. In solid phase hybridization assays, probes or primers are typically linked to a solid support where they are available for hybridization with target nucleic in solution. In mixed phase, nucleic acid intermediates in solution hybridize to target nucleic acids in solution as well as to a nucleic acid linked to a solid support. In in situ hybridization, the target nucleic acid is liberated from its cellular surroundings in such as to be available for hybridization within the cell while preserving the cellular morphology for subsequent interpretation and analysis. The following articles provide an overview of the various hybridization assay formats: Singer et al., Biotechniques 4(3): 230-250 (1986); Haase et al., Methods in Virology, Vol. VII, pp. 189-226 (1984); Wilkinson, The theory and practice of in situ hybridization in: In situ Hybridization, D.G. Wilkinson, Ed., IRL Press, Oxford University Press, Oxford; and Nucleic Acid Hybridization: A Practical Approach, Hames, B.D. and Higgins, S.J., Eds., IRL Press (1987).

# Nucleic Acid Labels and Detection Methods

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The means by which nucleic acids of the present invention are labeled is not a critical aspect of the present invention and can be accomplished by any number of methods currently known or later developed. Detectable labels suitable for use in the present invention include any composition detectable by spectroscopic, radioisotopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means. Useful labels in the present invention include biotin for staining with labeled streptavidin conjugate, magnetic beads, fluorescent dyes (e.g., fluorescein, texas red, rhodamine, green fluorescent protein, and the like), radiolabels (e.g., <sup>3</sup>H, <sup>125</sup>I, <sup>35</sup>S, <sup>14</sup>C, or <sup>32</sup>P), enzymes (e.g., horse radish peroxidase, alkaline phosphatase and others commonly used

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in an ELISA), and colorimetric labels such as colloidal gold or colored glass or plastic (e.g., polystyrene, polypropylene, latex, etc.) beads.

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Nucleic acids of the present invention can be labeled by any one of several methods typically used to detect the presence of hybridized nucleic acids. One common method of detection is the use of autoradiography using probes labeled with <sup>3</sup>H, <sup>125</sup>I, <sup>35</sup>S, <sup>14</sup>C, or <sup>32</sup>P, or the like. The choice of radio-active isotope depends on research preferences due to ease of synthesis, stability, and half lives of the selected isotopes. Other labels include ligands which bind to antibodies labeled with fluorophores, chemiluminescent agents, and enzymes. Alternatively, probes can be conjugated directly with labels such as fluorophores, chemiluminescent agents or enzymes. The choice of label depends on sensitivity required, ease of conjugation with the probe, stability requirements, and available instrumentation. Labeling the nucleic acids of the present invention is readily achieved such as by the use of labeled PCR primers.

In some embodiments, the label is simultaneously incorporated during the amplification step in the preparation of the nucleic acids. Thus, for example, polymerase chain reaction (PCR) with labeled primers or labeled nucleotides will provide a labeled amplification product. In another embodiment, transcription amplification using a labeled nucleotide (e.g., fluorescein-labeled UTP and/or CTP) incorporates a label into the transcribed nucleic acids.

Non-radioactive probes are often labeled by indirect means. For example, a ligand molecule is covalently bound to the probe. The ligand then binds to an anti-ligand molecule which is either inherently detectable or covalently bound to a detectable signal system, such as an enzyme, a fluorophore, or a chemiluminescent compound. Enzymes of interest as labels will primarily be hydrolases, such as phosphatases, esterases and glycosidases, or oxidoreductases, particularly peroxidases. Fluorescent compounds include fluorescein and its derivatives, rhodamine and its derivatives, dansyl, umbelliferone, etc. Chemiluminescers include luciferin, and 2,3-dihydrophthalazinediones, *e.g.*, luminol. Ligands and anti-ligands may be varied widely. Where a ligand has a natural anti-ligand, namely ligands such as biotin, thyroxine, and cortisol, it can be used in conjunction with its labeled, naturally occurring anti-ligands. Alternatively, any haptenic or antigenic compound can be used in combination with an antibody.

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Probes can also be labeled by direct conjugation with a label. For example, cloned DNA probes have been coupled directly to horseradish peroxidase or alkaline phosphatase, (Renz. M., and Kurz, K., A Colorimetric Method for DNA Hybridization, Nucl. Acids Res. 12: 3435-3444 (1984)) and synthetic oligonucleotides have been coupled directly with alkaline phosphatase (Jablonski, E., et al., Preparation of Oligodeoxynucleotide-Alkaline Phosphatase Conjugates and Their Use as Hybridization Probes, Nuc. Acids. Res. 14: 6115-6128 (1986); and Li P., et al., Enzyme-linked Synthetic Oligonucleotide probes: Non-Radioactive Detection of Enterotoxigenic Escherichia Coli in Faeca Specimens, Nucl. Acids Res. 15: 5275-5287 (1987)).

Means of detecting such labels are well known to those of skill in the art. Thus, for example, radiolabels may be detected using photographic film or scintillation counters, fluorescent markers may be detected using a photodetector to detect emitted light. Enzymatic labels are typically detected by providing the enzyme with a substrate and detecting the reaction product produced by the action of the enzyme on the substrate, and colorimetric labels are detected by simply visualizing the colored label.

#### **Antibodies to Proteins**

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Antibodies can be raised to a protein of the present invention, including individual, allelic, strain, or species variants, and fragments thereof, both in their naturally occurring (full-length) forms and in recombinant forms. Additionally, antibodies are raised to these proteins in either their native configurations or in non-native configurations. Anti-idiotypic antibodies can also be generated. Many methods of making antibodies are known to persons of skill. The following discussion is presented as a general overview of the techniques available; however, one of skill will recognize that many variations upon the following methods are known.

A number of immunogens are used to produce antibodies specifically reactive with a protein of the present invention. An isolated recombinant, synthetic, or native polynucleotide of the present invention are the preferred immunogens (antigen) for the production of monoclonal or polyclonal antibodies. Those of skill will readily understand that the proteins of the present invention are typically denatured, and optionally reduced, prior to formation of antibodies for screening expression libraries or other assays in which a putative protein of the present invention is expressed or

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denatured in a non-native secondary, tertiary, or quartenary structure. Non-isolated polypeptides of the present invention can be used either in pure or impure form.

The protein of the present invention is then injected into an animal capable of producing antibodies. Either monoclonal or polyclonal antibodies can be generated for subsequent use in immunoassays to measure the presence and quantity of the protein of the present invention. Methods of producing polyclonal antibodies are known to those of skill in the art. In brief, an immunogen (antigen), preferably a purified protein, a protein coupled to an appropriate carrier (e.g., GST, keyhole limpet hemanocyanin, etc.), or a protein incorporated into an immunization vector such as a recombinant vaccinia virus (see, U.S. Patent No. 4,722,848) is mixed with an adjuvant and animals are immunized with the mixture. The animal's immune response to the immunogen preparation is monitored by taking test bleeds and determining the titer of reactivity to the protein of interest. When appropriately high titers of antibody to the immunogen are obtained, blood is collected from the animal and antisera are prepared. Further fractionation of the antisera to enrich for antibodies reactive to the protein is performed where desired (See, e.g., Coligan, Current Protocols in Immunology, Wiley/Greene, NY (1991); and Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor Press, NY (1989)).

Antibodies, including binding fragments and single chain recombinant versions thereof, against predetermined fragments of a protein of the present invention are raised by immunizing animals, e.g., with conjugates of the fragments with carrier proteins as described above. Typically, the immunogen of interest is a protein of at least about 5 amino acids, more typically the protein is 10 amino acids in length, preferably, 15 amino acids in length and more preferably the protein is 20 amino acids in length or greater. The peptides are typically coupled to a carrier protein (e.g., as a fusion protein), or are recombinantly expressed in an immunization vector. Antigenic determinants on peptides to which antibodies bind are typically 3 to 10 amino acids in length.

Monoclonal antibodies are prepared from cells secreting the desired antibody. Monoclonals antibodies are screened for binding to a protein from which the immunogen was derived. Specific monoclonal and polyclonal antibodies will usually have an antibody binding site with an affinity constant for its cognate monovalent antigen at least between 10<sup>6</sup>-10<sup>7</sup>, usually at least 10<sup>8</sup>, preferably at least 10<sup>9</sup>, more preferably at least 10<sup>10</sup>, and most preferably at least 10<sup>11</sup> liters/mole.

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In some instances, it is desirable to prepare monoclonal antibodies from various mammalian hosts, such as mice, rodents, primates, humans, etc. Description of techniques for preparing such monoclonal antibodies are found in, e.g., Basic and Clinical Immunology, 4th ed., Stites et al., Eds., Lange Medical Publications, Los Altos, CA, and references cited therein; Harlow and Lane, Supra; Goding, Monoclonal 5 Antibodies: Principles and Practice, 2nd ed., Academic Press, New York, NY (1986); and Kohler and Milstein, Nature 256: 495-497 (1975). Summarized briefly, this method proceeds by injecting an animal with an immunogen comprising a protein of the present invention. The animal is then sacrificed and cells taken from its spleen, which are fused with myeloma cells. The result is a hybrid cell or "hybridoma" that is capable of 10 reproducing in vitro. The population of hybridomas is then screened to isolate individual clones, each of which secrete a single antibody species to the immunogen. In this manner, the individual antibody species obtained are the products of immortalized and cloned single B cells from the immune animal generated in response to a specific site recognized on the immunogenic substance.

Other suitable techniques involve selection of libraries of recombinant antibodies in phage or similar vectors (see, e.g., Huse et al., Science 246: 1275-1281 (1989); and Ward, et al., Nature 341: 544-546 (1989); and Vaughan et al., Nature Biotechnology, 14: 309-314 (1996)). Alternatively, high avidity human monoclonal antibodies can be obtained from transgenic mice comprising fragments of the unrearranged human heavy and light chain Ig loci (i.e., minilocus transgenic mice). Fishwild et al., Nature Biotech., 14: 845-851 (1996). Also, recombinant immunoglobulins may be produced. See, Cabilly, U.S. Patent No. 4,816,567; and Queen et al., Proc. Nat'l Acad. Sci. 86: 10029-10033 (1989).

The antibodies of this invention are also used for affinity chromatography in isolating proteins of the present invention. Columns are prepared, e.g., with the antibodies linked to a solid support, e.g., particles, such as agarose, Sephadex, or the like, where a cell lysate is passed through the column, washed, and treated with increasing concentrations of a mild denaturant, whereby purified protein are released.

The antibodies can be used to screen expression libraries for particular expression products such as normal or abnormal protein. Usually the antibodies in such a procedure are labeled with a moiety allowing easy detection of presence of antigen by antibody binding.

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Antibodies raised against a protein of the present invention can also be used to raise anti-idiotypic antibodies. These are useful for detecting or diagnosing various pathological conditions related to the presence of the respective antigens.

Frequently, the proteins and antibodies of the present invention will be labeled by joining, either covalently or non-covalently, a substance which provides for a detectable signal. A wide variety of labels and conjugation techniques are known and are reported extensively in both the scientific and patent literature. Suitable labels include radionucleotides, enzymes, substrates, cofactors, inhibitors, fluorescent moieties, chemiluminescent moieties, magnetic particles, and the like.

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# **Protein Immunoassays**

Means of detecting the proteins of the present invention are not critical aspects of the present invention. In a preferred embodiment, the proteins are detected and/or quantified using any of a number of well recognized immunological binding assays (see, e.g., U.S. Patents 4,366,241; 4,376,110; 4,517,288; and 4,837,168). For a review of 15 the general immunoassays, see also Methods in Cell Biology, Vol. 37: Antibodies in Cell Biology, Asai, Ed., Academic Press, Inc. New York (1993); Basic and Clinical Immunology 7th Edition, Stites & Terr, Eds. (1991). Moreover, the immunoassays of the present invention can be performed in any of several configurations, e.g., those reviewed in Enzyme Immunoassay, Maggio, Ed., CRC Press, Boca Raton, Florida 20 (1980); Tijan, Practice and Theory of Enzyme Immunoassays, Laboratory Techniques in Biochemistry and Molecular Biology, Elsevier Science Publishers B.V., Amsterdam (1985); Harlow and Lane, above; Immunoassay: A Practical Guide, Chan, Ed., Academic Press, Orlando, FL (1987); Principles and Practice of Immunoassaysm, Price and Newman Eds., Stockton Press, NY (1991); and Non-isotopic Immunoassays, Ngo, 25 Ed., Plenum Press, NY (1988). Immunological binding assays (or immunoassays) typically utilize a "capture agent" to specifically bind to and often immobilize the analyte (in this case, a protein of the present invention). The capture agent is a moiety that specifically binds to the analyte. In a preferred embodiment, the capture agent is an 30 antibody that specifically binds a protein(s) of the present invention. The antibody may be produced by any of a number of means known to those of skill in the art as described herein.

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Immunoassays also often utilize a labeling agent to specifically bind to and label the binding complex formed by the capture agent and the analyte. The labeling agent may itself be one of the moieties comprising the antibody/analyte complex. Thus, the labeling agent may be a labeled protein of the present invention or a labeled antibody specifically reactive to a protein of the present invention. Alternatively, the labeling agent may be a third moiety, such as another antibody, that specifically binds to the antibody/protein complex.

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In a preferred embodiment, the labeling agent is a second antibody bearing a label. Alternatively, the second antibody may lack a label, but it may, in turn, be bound by a labeled third antibody specific to antibodies of the species from which the second antibody is derived. The second can be modified with a detectable moiety, such as biotin, to which a third labeled molecule can specifically bind, such as enzyme-labeled streptavidin.

Other proteins capable of specifically binding immunoglobulin constant regions, such as protein A or protein G may also be used as the label agent. These proteins are normal constituents of the cell walls of streptococcal bacteria. They exhibit a strong non-immunogenic reactivity with immunoglobulin constant regions from a variety of species (See, generally Kronval, et al., J. Immunol. 111: 1401-1406 (1973), and Akerstrom, et al., J. Immunol. 135: 2589-2542 (1985)).

Throughout the assays, incubation and/or washing steps may be required after each combination of reagents. Incubation steps can vary from about 5 seconds to several hours, preferably from about 5 minutes to about 24 hours. However, the incubation time will depend upon the assay format, analyte, volume of solution, concentrations, and the like. Usually, the assays will be carried out at ambient temperature, although they can be conducted over a range of temperatures, such as 10°C to 40°C.

While the details of the immunoassays of the present invention may vary with the particular format employed, the method of detecting a protein of the present invention in a biological sample generally comprises the steps of contacting the biological sample with an antibody which specifically reacts, under immunologically reactive conditions, to a protein of the present invention. The antibody is allowed to bind to the protein under immunologically reactive conditions, and the presence of the bound antibody is detected directly or indirectly.

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# A. Non-Competitive Assay Formats

Immunoassays for detecting proteins of the present invention include competitive and noncompetitive formats. Noncompetitive immunoassays are assays in which the amount of captured analyte (i.e., a protein of the present invention) is directly measured. In one preferred "sandwich" assay, for example, the capture agent (e.g., an antibody specifically reactive, under immunoreactive conditions, to a protein of the present invention) can be bound directly to a solid substrate where they are immobilized. These immobilized antibodies then capture the protein present in the test sample. The protein thus immobilized is then bound by a labeling agent, such as a second antibody bearing a label. Alternatively, the second antibody may lack a label, but it may, in turn, be bound by a labeled third antibody specific to antibodies of the species from which the second antibody is derived. The second can be modified with a detectable moiety, such as biotin, to which a third labeled molecule can specifically bind, such as enzyme-labeled streptavidin.

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# B. Competitive Assay Formats

In competitive assays, the amount of analyte present in the sample is measured indirectly by measuring the amount of an added (exogenous) analyte (e.g., a protein of the present invention) displaced (or competed away) from a capture agent (e.g., an antibody specifically reactive, under immunoreactive conditions, to the protein) by the analyte present in the sample. In one competitive assay, a known amount of analyte is added to the sample and the sample is then contacted with a capture agent that specifically binds a protein of the present invention. The amount of protein bound to the capture agent is inversely proportional to the concentration of analyte present in the sample.

In a particularly preferred embodiment, the antibody is immobilized on a solid substrate. The amount of protein bound to the antibody may be determined either by measuring the amount of protein present in a protein/antibody complex, or alternatively by measuring the amount of remaining uncomplexed protein. The amount of protein may be detected by providing a labeled protein.

A hapten inhibition assay is another preferred competitive assay. In this assay a known analyte, (such as a protein of the present invention) is immobilized on a solid substrate. A known amount of antibody specifically reactive, under immunoreactive

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conditions, to the protein is added to the sample, and the sample is then contacted with the immobilized protein. In this case, the amount of antibody bound to the immobilized protein is inversely proportional to the amount of protein present in the sample. Again, the amount of immobilized antibody may be detected by detecting either the immobilized fraction of antibody or the fraction of the antibody that remains in solution. Detection may be direct where the antibody is labeled or indirect by the subsequent addition of a labeled moiety that specifically binds to the antibody as described above.

# C. Generation of pooled antisera for use in immunoassays

A protein that specifically binds to or that is specifically immunoreactive with an antibody generated against a defined immunogen is determined in an immunoassay. The immunoassay uses a polyclonal antiserum which is raised to a polypeptide of the present invention (i.e., the immunogenic polypeptide). This antiserum is selected to have low crossreactivity against other proteins and any such crossreactivity is removed by immunoabsorbtion prior to use in the immunoassay (e.g., by immunosorbtion of the antisera with a protein of different substrate specificity (e.g., a different enzyme) and/or a protein with the same substrate specificity but of a different form).

In order to produce antisera for use in an immunoassay, a polypeptide of the present invention is isolated as described herein. For example, recombinant protein can be produced in a mammalian or other eukaryotic cell line. An inbred strain of mice is immunized with the protein using a standard adjuvant, such as Freund's adjuvant, and a standard mouse immunization protocol (see Harlow and Lane, above). Alternatively, a synthetic polypeptide derived from the sequences disclosed herein and conjugated to a carrier protein is used as an immunogen. Polyclonal sera are collected and titered against the immunogenic polypeptide in an immunoassay, for example, a solid phase immunoassay with the immunogen immobilized on a solid support. Polyclonal antisera with a titer of 10<sup>4</sup> or greater are selected and tested for their cross reactivity against polypeptides of different forms or substrate specificity, using a competitive binding immunoassay such as the one described in Harlow and Lane, above, at pages 570-573. Preferably, two or more distinct forms of polypeptides are used in this determination. These distinct types of polypeptides are used as competitors to identify antibodies which are specifically bound by the polypeptide being assayed for. The competitive

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polypeptides can be produced as recombinant proteins and isolated using standard molecular biology and protein chemistry techniques as described herein.

Immunoassays in the competitive binding format are used for crossreactivity determinations. For example, the immunogenic polypeptide is immobilized to a solid support. Proteins added to the assay compete with the binding of the antisera to the immobilized antigen. The ability of the above proteins to compete with the binding of the antisera to the immobilized protein is compared to the immunogenic polypeptide. The percent crossreactivity for the above proteins is calculated, using standard calculations. Those antisera with less than 10% crossreactivity with a distinct form of a polypeptide are selected and pooled. The cross-reacting antibodies are then removed from the pooled antisera by immunoabsorbtion with a distinct form of a polypeptide.

The immunoabsorbed and pooled antisera are then used in a competitive binding immunoassay as described herein to compare a second "target" polypeptide to the immunogenic polypeptide. In order to make this comparison, the two polypeptides are each assayed at a wide range of concentrations and the amount of each polypeptide required to inhibit 50% of the binding of the antisera to the immobilized protein is determined using standard techniques. If the amount of the target polypeptide required is less than twice the amount of the immunogenic polypeptide that is required, then the target polypeptide is said to specifically bind to an antibody generated to the immunogenic protein. As a final determination of specificity, the pooled antisera is fully immunosorbed with the immunogenic polypeptide until no binding to the polypeptide used in the immunosorbtion is detectable. The fully immunosorbed antisera is then tested for reactivity with the test polypeptide. If no reactivity is observed, then the test polypeptide is specifically bound by the antisera elicited by the immunogenic protein.

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# D. Other Assay Formats

In a particularly preferred embodiment, Western blot (immunoblot) analysis is used to detect and quantify the presence of protein of the present invention in the sample. The technique generally comprises separating sample proteins by gel electrophoresis on the basis of molecular weight, transferring the separated proteins to a suitable solid support, (such as a nitrocellulose filter, a nylon filter, or derivatized nylon filter), and incubating the sample with the antibodies that specifically bind a protein of the present invention. The antibodies specifically bind to the protein on the solid support. These

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antibodies may be directly labeled or alternatively may be subsequently detected using labeled antibodies (e.g., labeled sheep anti-mouse antibodies) that specifically bind to the antibodies.

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## E. Quantification of Proteins.

The proteins of the present invention may be detected and quantified by any of a number of means well known to those of skill in the art. These include analytic biochemical methods such as electrophoresis, capillary electrophoresis, high performance liquid chromatography (HPLC), thin layer chromatography (TLC), hyperdiffusion chromatography, and the like, and various immunological methods such as fluid or gel precipitin reactions, immunodiffusion (single or double), immunoelectrophoresis, radioimmunoassays (RIAs), enzyme-linked immunosorbent assays (ELISAs), immunofluorescent assays, and the like.

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# F. Reduction of Non-Specific Binding

One of skill will appreciate that it is often desirable to reduce non-specific binding in immunoassays and during analyte purification. Where the assay involves an antigen, antibody, or other capture agent immobilized on a solid substrate, it is desirable to minimize the amount of non-specific binding to the substrate. Means of reducing such non-specific binding are well known to those of skill in the art. Typically, this involves coating the substrate with a proteinaceous composition. In particular, protein compositions such as bovine serum albumin (BSA), nonfat powdered milk, and gelatin are widely used.

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## G. Immunoassay Labels

The labeling agent can be, e.g., a monoclonal antibody, a polyclonal antibody, a binding protein or complex, or a polymer such as an affinity matrix, carbohydrate or lipid. Detectable labels suitable for use in the present invention include any composition detectable by spectroscopic, radioisotopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means. Detection may proceed by any known method, such as immunoblotting, western analysis, gel-mobility shift assays, fluorescent in situ hybridization analysis (FISH), tracking of radioactive or

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bioluminescent markers, nuclear magnetic resonance, electron paramagnetic resonance, stopped-flow spectroscopy, column chromatography, capillary electrophoresis, or other methods which track a molecule based upon an alteration in size and/or charge. The particular label or detectable group used in the assay is not a critical aspect of the invention. The detectable group can be any material having a detectable physical or chemical property. Such detectable labels have been well-developed in the field of immunoassays and, in general, any label useful in such methods can be applied to the present invention. Thus, a label is any composition detectable by spectroscopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means.

10 Useful labels in the present invention include magnetic beads, fluorescent dyes, radiolabels, enzymes, and colorimetric labels or colored glass or plastic beads, as discussed for nucleic acid labels, above.

The label may be coupled directly or indirectly to the desired component of the assay according to methods well known in the art. As indicated above, a wide variety of labels may be used, with the choice of label depending on the sensitivity required, ease of conjugation of the compound, stability requirements, available instrumentation, and disposal provisions.

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Non-radioactive labels are often attached by indirect means. Generally, a ligand molecule (e.g., biotin) is covalently bound to the molecule. The ligand then binds to an anti-ligand (e.g., streptavidin) molecule which is either inherently detectable or covalently bound to a signal system, such as a detectable enzyme, a fluorescent compound, or a chemiluminescent compound. A number of ligands and anti-ligands can be used. Where a ligand has a natural anti-ligand, for example, biotin, thyroxine, and cortisol, it can be used in conjunction with the labeled, naturally occurring anti-ligands. Alternatively, any haptenic or antigenic compound can be used in combination with an antibody.

The molecules can also be conjugated directly to signal generating compounds, e.g., by conjugation with an enzyme or fluorophore. Enzymes of interest as labels will primarily be hydrolases, particularly phosphatases, esterases and glycosidases, or oxidoreductases, particularly peroxidases. Fluorescent compounds include fluorescein and its derivatives, rhodamine and its derivatives, dansyl, umbelliferone, etc. Chemiluminescent compounds include luciferin, and 2,3-dihydrophthalazinediones, e.g.,

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luminol. For a review of various labeling or signal producing systems which may be used, see, U.S. Patent No. 4,391,904, which is incorporated herein by reference.

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Means of detecting labels are well known to those of skill in the art. Thus, for example, where the label is a radioactive label, means for detection include a scintillation counter or photographic film as in autoradiography. Where the label is a fluorescent label, it may be detected by exciting the fluorochrome with the appropriate wavelength of light and detecting the resulting fluorescence, *e.g.*, by microscopy, visual inspection, via photographic film, by the use of electronic detectors such as charge coupled devices (CCDs) or photomultipliers and the like. Similarly, enzymatic labels may be detected by providing appropriate substrates for the enzyme and detecting the resulting reaction product. Finally, simple colorimetric labels may be detected simply by observing the color associated with the label. Thus, in various dipstick assays, conjugated gold often appears pink, while various conjugated beads appear the color of the bead.

Some assay formats do not require the use of labeled components. For instance, agglutination assays can be used to detect the presence of the target antibodies. In this case, antigen-coated particles are agglutinated by samples comprising the target antibodies. In this format, none of the components need be labeled and the presence of the target antibody is detected by simple visual inspection.

# 20 Assays for Compounds that Modulate Enzymatic Activity or Expression

The present invention also provides means for identifying compounds that bind to (e.g., substrates), and/or increase or decrease (i.e., modulate) the enzymatic activity of, catalytically active polypeptides of the present invention. The method comprises contacting a polypeptide of the present invention with a compound whose ability to bind to or modulate enzyme activity is to be determined. The polypeptide employed will have at least 20%, preferably at least 30% or 40%, more preferably at least 50% or 60%, and most preferably at least 70% or 80% of the specific activity of the native, full-length polypeptide of the present invention (e.g., enzyme). Generally, the polypeptide will be present in a range sufficient to determine the effect of the compound, typically about 1 nM to  $10~\mu M$ . Likewise, the compound will be present in a concentration of from about 1 nM to  $10~\mu M$ . Those of skill will understand that such factors as enzyme concentration, ligand concentrations (i.e., substrates, products, inhibitors, activators), pH, ionic strength, and temperature will be controlled so as to obtain useful kinetic data

and determine the presence of absence of a compound that binds or modulates polypeptide activity. Methods of measuring enzyme kinetics is well known in the art. See, e.g., Segel, *Biochemical Calculations*, 2<sup>nd</sup> ed., John Wiley and Sons, New York (1976).

Although the present invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

## 10 Example 1

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This example describes the construction cDNA libraries.

## **Total RNA Isolation**

Inc. Gaithersburg, MD) using a modification of the guanidine isothiocyanate/acid-phenol procedure described by Chomczynski and Sacchi (Chomczynski, P., and Sacchi, N. Anal. Biochem. 162, 156 (1987)). In brief, plant tissue samples were pulverized in liquid nitrogen before the addition of the TRIzol Reagent, and then were further homogenized with a mortar and pestle. Addition of chloroform followed by centrifugation was conducted for separation of an aqueous phase and an organic phase. The total RNA was recovered by precipitation with isopropyl alcohol from the aqueous phase.

## Poly(A) + RNA Isolation

The selection of poly(A)+ RNA from total RNA was performed using PolyATact system (Promega Corporation. Madison, WI). In brief, biotinylated oligo(dT) primers were used to hybridize to the 3' poly(A) tails on mRNA. The hybrids were captured using streptavidin coupled to paramagnetic particles and a magnetic separation stand. The mRNA was washed at high stringent condition and eluted by RNase-free deionized water.

## cDNA Library Construction

cDNA synthesis was performed and unidirectional cDNA libraries were constructed using the SuperScript Plasmid System (Life Technology Inc. Gaithersburg, MD). The first stand of cDNA was synthesized by priming an oligo(dT) primer containing a Not I site. The reaction was catalyzed by SuperScript Reverse

Transcriptase II at 45°C. The second strand of cDNA was labeled with alpha-32P-dCTP and a portion of the reaction was analyzed by agarose gel electrophoresis to determine cDNA sizes. cDNA molecules smaller than 500 base pairs and unligated adapters were removed by Sephacryl-S400 chromatography. The selected cDNA molecules were ligated into pSPORT1 vector in between of Not I and Sal I sites.

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#### Example 2

This example describes cDNA sequencing and library subtraction.

## **Sequencing Template Preparation**

Individual colonies were picked and DNA was prepared either by PCR with M13 forward primers and M13 reverse primers, or by plasmid isolation. All the cDNA clones were sequenced using M13 reverse primers.

### **Q-bot Subtraction Procedure**

cDNA libraries subjected to the subtraction procedure were plated out on 22 x 22 cm<sup>2</sup> agar plate at density of about 3,000 colonies per plate. The plates were incubated in a 37°C incubator for 12-24 hours. Colonies were picked into 384-well plates by a robot colony picker, Q-bot (GENETIX Limited). These plates were incubated overnight at 37°C.

Once sufficient colonies were picked, they were pinned onto  $22 \times 22 \text{ cm}^2$  nylon membranes using Q-bot. Each membrane contained 9,216 colonies or 36,864 colonies. These membranes were placed onto agar plate with appropriate antibiotic. The plates were incubated at  $37^{\circ}\text{C}$  for overnight.

After colonies were recovered on the second day, these filters were placed on filter paper prewetted with denaturing solution for four minutes, then were incubated on top of a boiling water bath for additional four minutes. The filters were then placed on filter paper prewetted with neutralizing solution for four minutes. After excess solution was removed by placing the filters on dry filter papers for one minute, the colony side of the filters were place into Proteinase K solution, incubated at 37°C for 40-50 minutes.

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The filters were placed on dry filter papers to dry overnight. DNA was then crosslinked to nylon membrane by UV light treatment.

Colony hybridization was conducted as described by Sambrook, J., Fritsch, E.F. and Maniatis, T., (in Molecular Cloning: A laboratory Manual, 2<sup>nd</sup> Edition). The following probes were used in colony hybridization:

- 1. First strand cDNA from the same tissue as the library was made from to remove the most redundant clones.
- 2. 48-192 most redundant cDNA clones from the same library based on previous sequencing data.
- 10 3. 192 most redundant cDNA clones in the entire corn partial sequence database.
  - 4. A Sal-A20 oligo nucleotide: TCG ACC CAC GCG TCC GAA AAA AAA AAA AAA AAA AAA AAA, removes clones containing a poly A tail but no cDNA.
  - 5. cDNA clones derived from rRNA.

The image of the autoradiography was scanned into computer and the signal intensity and cold colony addresses of each colony was analyzed. Re-arraying of cold-colonies from 384 well plates to 96 well plates was conducted using Q-bot.

### Example 3

This example describes identification of the gene from a computer homology 20 Gene identities were determined by conducting BLAST (Basic Local search. Alignment Search Tool; Altschul, S. F., et al., (1993) J. Mol. Biol. 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/) searches under default parameters for similarity to sequences contained in the BLAST "nr" database (comprising all non-redundant GenBank CDS translations, sequences derived from the 3-dimensional structure 25 Brookhaven Protein Data Bank, the last major release of the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The cDNA sequences were analyzed for similarity to all publicly available DNA sequences contained in the "nr" database using the BLASTN algorithm. The DNA sequences were translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the 30 "nr" database using the BLASTX algorithm (Gish, W. and States, D. J. (1993) Nature Genetics 3:266-272) provided by the NCBI. In some cases, the sequencing data from two or more clones containing overlapping segments of DNA were used to construct contiguous DNA sequences.

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The above examples are provided to illustrate the invention but not to limit its scope. Other variants of the invention will be readily apparent to one of ordinary skill in the art and are encompassed by the appended claims. All publications, patents, and patent applications cited herein are hereby incorporated by reference.

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## WHAT IS CLAIMED IS:

- 1. An isolated nucleic acid comprising a member selected from the group consisting of:
- 5 (a) a polynucleotide having at least 80% sequence identity, as determined by the BLAST 2.0 algorithm under default parameters, to a polynucleotide encoding a polypeptide selected from the group consisting of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58
  - (b) a polynucleotide encoding a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58
  - (c) a polynucleotide amplified from a *Zea mays* nucleic acid library using primers which selectively hybridize, under stringent hybridization conditions, to loci within a polynucleotide selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57
- 15 (d) a polynucleotide which selectively hybridizes, under stringent hybridization conditions and a wash in 2X SSC at 50°C, to a polynucleotide of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57
  - (e) a polynucleotide of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57
- 20 (f) a polynucleotide which is complementary to a polynucleotide of (a), (b), (c), (d), or (e); and
  - (g) a polynucleotide comprising at least 25 contiguous nucleotides from a polynucleotide of (a), (b), (c), (d), (e), or (f).
- 25 2. A recombinant expression cassette, comprising a member of claim 1 operably linked, in sense or anti-sense orientation, to a promoter.
  - 3. A host cell comprising the recombinant expression cassette of claim 2.
- 30 4. A transgenic plant comprising a recombinant expression cassette of claim 2.
  - 5. The transgenic plant of claim 4, wherein the plant is a monocot.

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- 6. The transgenic plant of claim 4, wherein the plant is selected from the group consisting of: maize, soybean, sunflower, sorghum, canola, wheat, alfalfa, cotton, rice, barley, and millet.
- 5 7. A transgenic seed from the transgenic plant of claim 4.
  - 8. A method of modulating the level of cellulose synthase in a plant cell capable of plant regeneration, comprising:
    - (a) transforming the plant cell with a recombinant expression cassette comprising a cellulose synthase polynucleotide of claim 1 operably linked to a promoter;
    - (b) culturing the transformed plant cell; and

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- (c) inducing expression of said polynucleotide for a time sufficient to modulate the level of cellulose synthase in said transformed plant cell.
- 15 9. The method of claim 8, wherein a plant is regenerated from the transformed plant cell.
- 10. The method of claim 9, wherein the plant is selected from the group consisting of maize, soybean, sunflower, sorghum, canola, wheat, alfalfa, cotton, rice, barley, and millet.
  - 11. The method of claim 8, wherein the promoter is a tissue-preferred promoter.
  - 12. The method of claim 8, wherein the level of cellulose synthase is increased.
  - 13. The method of claim 8 wherein the cell cycle polynucleotide is amplified from a Zea mays nucleic acid library using primers which selectively hybridize, under stringent hybridization conditions, to loci within a polynucleotide selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57.
  - 14. The method of claim 8 wherein the cell cycle gene is selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57.

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- 15. An isolated protein comprising a member selected from the group consisting of:
  - (a) a polypeptide of at least 20 contiguous amino acids from a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58
  - (b) a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58
  - (c) a polypeptide having at least 80% sequence identity to, and having at least one linear epitope in common with, a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58, wherein said sequence identity is determined using BLAST 2.0 under default parameters; and,
- 10 (d) a polypeptide encoded by a member of claim 1.

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#### SEQUENCE LISTING

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- 4 -

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- 6 -

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Gin	Phe	Pro	Gln	Δτα	Dhe	Acn	Glv	Tla	Aen	Ara	Uic	7 cn	7 ~~	Tire	7.7.2
			Gln Val	565					570					575	
Asn	Arg	Asn	Val 580	565 Val	Phe	Phe	Asp	Ile 585	570 Asn	Met	Lys	Gly	Leu 590	575 Asp	Gly
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Trp Tyr Gly Tyr Gly Gly Gly Leu Lys Phe Leu Glu Arg Phe Ser Tyr
            825 830
Ile Asn Ser Ile Val Tyr Pro Trp Thr Ser Ile Pro Leu Leu Ala Tyr
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Cys Thr Leu Pro Ala Ile Cys Leu Leu Thr Gly Lys Phe Ile Thr Pro
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Asn Ala Ile Asn Asn Gly Tyr Glu Ser Trp Gly Pro Leu Phe Gly Lys
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Ser Ile Leu Leu Ala Ser Ile Phe Ser Leu Leu Trp Val Arg Ile Asp
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												att Ile				1027
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-												atg Met 275		_	_	1171
												cgt Arg				1219
												gtt Val				1267
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WO 00/09706

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											ctt Leu					2083
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att Ile 615	aac Asn	ttg Leu	aga Arg	ggt Gly	ctt Leu 620	gat Asp	ggc Gly	atc Ile	caa Gln	gga Gly 625	cca Pro	gtt Val	tat Tyr	gtc Val	gga Gly 630	2227
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Thr	Gly	Cys	Val	Phe 635	Asn	Arg	Thr	Ala	Leu 640	Tyr	Gļy	Tyr	Glu	Pro 645	Pro	
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_	cat His 680		_	_				_				_	_			2419
	gga Gly	_	_		-			_	-						-	2467
	caa Gln	_	_	_		_	-					_			-	2515
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			_	_	_	_		-					_		gly aaa	2803
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		Arg													acc Thr	2899
	Tyr	_		_		Ile	_				Tyr			_	ccc Pro 870	2947

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410

405

- 15 -

Phe Val Lys Lys Tyr Asn Ile Glu Pro Arg Ala Pro Glu Trp Tyr Phe 425 420 Ser Gln Lys Ile Asp Tyr Leu Lys Asp Lys Val His Pro Ser Phe Val 440 Lys Asp Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg 455 Val Asn Gly Leu Val Ala Lys Ala Gln Lys Val Pro Glu Glu Gly Trp 470 475 Ile Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn Thr Xaa Asp His 490 485 Pro Gly Met Ile Gln Val Phe Leu Gly His Ser Gly Gly Leu Asp Thr 505 500 Glu Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg 520 Pro Gly Phe Gln His His Lys Lys Ala Gly Ala Met Asn Ala Leu Val 535 Arg Val Ser Ala Val Leu Thr Asn Gly Gln Tyr Met Leu Asn Leu Asp 550 555 Cys Asp His Tyr Ile Asn Asn Ser Lys Ala Leu Arg Glu Ala Met Cys 565 570 Phe Leu Met Asp Pro Asn Leu Gly Arg Ser Val Cys Tyr Val Gln Phe 580 585 Pro Gln Arg Phe Asp Gly Ile Asp Arg Asn Asp Arg Tyr Ala Asn Arg 605 600 Asn Thr Val Phe Phe Asp Ile Asn Leu Arg Gly Leu Asp Gly Ile Gln 615 620 Gly Pro Val Tyr Val Gly Thr Gly Cys Val Phe Asn Arg Thr Ala Leu 630 Tyr Gly Tyr Glu Pro Pro Ile Lys Gln Lys Lys Gly Gly Phe Leu Ser 650 Ser Leu Cys Gly Gly Arg Lys Lys Ala Ser Lys Ser Lys Gly Ser 665 Asp Lys Lys Ser Gln Lys His Val Asp Ser Ser Val Pro Val Phe 675 680 Asn Leu Glu Asp Ile Glu Glu Gly Val Glu Gly Ala Gly Phe Asp Asp 695 Glu Lys Ser Leu Leu Met Ser Gln Met Ser Leu Glu Lys Arg Phe Gly 710 715 Gln Ser Ala Ala Phe Val Ala Ser Thr Leu Met Glu Tyr Gly Gly Val 730 Pro Gln Ser Ala Thr Pro Glu Ser Leu Leu Lys Glu Ala Ile His Val 745 Ile Ser Cys Gly Tyr Glu Asp Lys Thr Glu Trp Gly Thr Glu Ile Gly 760 Trp Ile Tyr Gly Ser Val Thr Glu Asp Ile Leu Thr Gly Phe Lys Met 775 His Ala Arg Gly Trp Arg Ser Ile Tyr Cys Met Pro Lys Arg Pro Ala 790 795 Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp Arg Leu Asn Gln Val 810 Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Leu Phe Ser Arg His Cys 825 Pro Leu Trp Tyr Gly Tyr Gly Gly Arg Leu Lys Phe Leu Glu Arg Phe 845 840 Ala Tyr Ile Asn Thr Thr Ile Tyr Pro Leu Thr Ser Ile Pro Leu Leu 855 860 Ile Tyr Cys Ile Leu Pro Ala Ile Cys Leu Leu Thr Gly Lys Phe Ile 870 875

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		tac Tyr 270														1049
		cag Gln		-	-	_				-	-	-				1097
		gta Val														1145
	-	cag Gln									_					1193
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		_				-		Leu	_	_					tca Ser	1529
		Lys					Met					Glu			aaa Lys	1577

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caa Gln	ttt Phe	eca Pro	Gln	aga Arg	ttt Phe	gat Asp	ggc Gly 595	· Ile	gac Asp	ttg Leu	cac His	gat Asp 600	Arg	tat Tyr	gct Ala	2009
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aac Ası	att n Ile	gt: Vai	t at: 1 Il: 65!	e Ly:	g ago s Sei	c tgo	tgt GCys	ggt Gly 660	y Arg	a agg	g aag g Ly:	g aaa s Lya	a aag s Ly: 66	s As	c aag n Lys	2201
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atc Ile					_			_		_					2873
ttc Phe															2921
gtt Val				_			_			-			-		2969

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				att Ile												3065
				gac Asp 960												3113
_				ccg Pro			_		-						-	3161
			Ile	tct Ser				Asn					Ser			3209
		Phe		aag Lys			Phe					Ile				3257
	Pro			aag Lys		Leu					Asn					3305
				tgg Trp 104	Ser					Ser					Leu	3353
				gat Asp 5					Pro					Ala		3401
_		_	Cys	ggc	_			ctga	tcga	g ac	agtg	actc	tta	tttg	aag	3453
agg	rctica	atc	aaga	tata	ממ מ	cata	atat	a aa	tacc	taaa	gao	acta	gat	ggga	attect	3513
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cca	gcag	cgt	aaga	tgtg	aa t	tttg	aagt	t tt	gtta	.tgcg	tgo : ata	agtt	cat	tact	ttagag gttatt	3693 3753
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                                                                  178
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                                                                  226
 Met Ala Ala Asn Lys Gly Met Val Ala Gly Ser His Asn Arg Asn Glu
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                                                                  274
 Phe Val Met Ile Arg His Asp Gly Asp Ala Pro Val Pro Ala Lys Pro
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 Thr Lys Ser Ala Asn Gly Gln Val Cys Gln Ile Cys Gly Asp Thr Val
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	_	-		_	tgc Cys	_		-		_	_	_				466
					gat Asp											514
	•				aag Lys					_						562
			_	-	gct Ala	-	_				-	_		-		610
					cgc Arg 150											658
		_	_		cct Pro	_	_				_					706
					agc Ser											754
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		Glu			agg Arg		Lys					Met			gtg Val	850
	Asn					Ala					Glu				tca Ser 240	898
		-	-	_	Gln	_	-	_	_	Āla	_			_	agc Ser	946
				Ile					Lev					Ile	gta Val	994
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tgt Cys 305	gag Glu	gtc Val	tgg Trp	ttt Phe	gcc Ala 310	ttg Leu	tcc Ser	tgg Trp	ctt Leu	cta Leu 315	gat Asp	cag Gln	ttc Phe	cca Pro	aaa Lys 320	1138
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gtc Val	agt Ser	aca Thr 355	gtg Val	gat Asp	cca Pro	ttg Leu	aag Lys 360	gaa Glu	cct Pro	cca Pro	ctg Leu	atc Ile 365	aca Thr	gcc Ala	aac Asn	1282
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gat Asp	tac Tyr	cto Leu 435	Lys	gac Asp	aaa Lys	att Ile	caa Gln 440	Pro	tca Ser	ttt Phe	gtt Val	aag Lys 445	Glu	aga Arg	cga	1522
gca Ala	a ato a Met 450	Lys	g aga s Arg	gag Glu	r tat ı Tyr	gaa Glu 455	ı Glu	tto Phe	aaa Lys	ata : Ile	aga Arg	a ato g Ile O	aat Asn	geo Ala	ctt Leu	1570
gti Va: 46!	l Ala	c aaa a Lys	a gca s Ala	a caç a Glr	aaa Lys 470	val	g cct L Pro	gaa Glu	a gag ı Glı	999 Gly 479	r Trj	g acc p Thr	atg Met	gct Ala	gat Asp 480	1618
gg; Gl;	a act	z ge	t tgg a Tr	g cct Pro 48	o Gly	g aat y Asi	aac n Ası	e eet	t agg Arg 490	g Asp	ca Hi	t cct s Pro	ggo Gly	ato Met 499	: Ile	1666
Ca Gl:	g gtg n Va	g tt l Ph	c ttg e Le: 50	ı Gl	g cad y His	c ag s Se	t gg r Gl	gg: y Gl: 50	y Lei	t gad u Asp	ac o Th	t gat r Asp	gga Gly 510	Ası	gaa n Glu	1714

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		_		gtc Val		_		-	_	_	_				_	1762
		_	_	gct Ala		_	_		_	_		_	_		_	1810
	_			ggt Gly	_						_	_	_			1858
		-	-	aaa Lys 565	-			_	-	_	_		_	_	-	1906
	_			agg Arg			_		_					_		1954
_		_	_	ttg Leu		_	_		-					_		2002
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_	-		_	aag Lys	_		_			_						2242
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	_		-	agg Arg		_			_			_				2338
		-		acc Thr 725	Phe	_									Thr	2386
aac	cca	gct	tct	cta	ctg	aag	gaa	gct	atc	cat	gtt	atc	agc	tgt	ggg	2434

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Asn	Pro	Ala	Ser 740	Leu	Leu	Lys	Glu	Ala 745	Ile	His	Val	Ile	Ser 750	Cys	Gly	
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		_									atc Ile 860				_	2770
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tctgcatcca agttatgcct ctgtttatta gcttcttcgg tgccggtgct gctgcagaca atcatggagc ctttctacct tgcttgtagt gctggccagc agcgtaaatt gtgaattctg catttttta tacgtggtgt ttattgttt agagtaaatt atcatttgtt tgaggtaact attcacacga actatatggc aatgctgtta tttaaaa    <210 > 14	3568 3628 3688
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	530	)				535	5				540	)			r Ala
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				56	5				57	0				57	
			58	0				58	5				59	0	g Phe
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ago Ser	caç Glı	g at	t aa e As 27	n Pro	a tat o Tyi	agg r Arg	g ato	g ati	e Ile	c att	att E Ile	t cgg e Arg	g Cti g Lei 28	ı Va.	g gtt l Val	989
tt <u>e</u> Lev	g ggg	y tt y Ph 28	e Ph	c tte e Ph	c cad	c tac	c cg c Ar	g Va	g atq 1 Me	g cat t His	t cc	g gtg o Val 29!	L As:	t ga n As	t gca p Ala	1037
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agt Sei	tgi Cyi	gg s Gl: 76	у Туз	gaa Glu	a gad u Asp	c aag o Lys	aca Thi	c Asp	tgg Tr	g 996 o Gly	a aa y Ly	a gag s Glu 77!	ı Ile	t ggd e Gly	tgg Y Trp	2477
ato Ilo	e Ty	t gg r Gl	a tca y Sei	a gti r Vai	t aca	a gaa r Gl	a gat ı Ası	t att	cta e Le	a act	t gg r Gl	t tto y Pho	c aaq	g ato	g cat t His	2525

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aaa q Lys ( 1035				Gly		Gln			Thr		Thr					3293
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1 Leu Glu Ala Val 65 Cys	<pre>&lt;2 &lt;2 &lt;4 Glu Val Gln Pro 50 Cys Pro Thr</pre>	211> 212> 213> 400> Ala Val Asn 35 Gly Arg Gln Gly Trp	1086 PRT Zea  18 Ser  Ile 20 Gly Gly Asp Cys Asp 100	Ala 5 Arg Gln Asp Cys Lys 85 Glu	Gly Arg Val Pro Tyr 70 Thr	Asp Cys Phe 55 Glu Arg	Gly Gln 40 Val Tyr Tyr Asp	Asp 25 Ile Ala Glu Lys Gly 105 Gln	10 Pro Cys Cys Arg Arg 90 Val	Gly Gly Asn Arg 75 Leu Asp	Pro Asp Glu 60 Glu Lys Asp	Lys Asp 45 Cys Gly Gly Leu Glu	Pro 30 Val Ala Thr Cys Asp 110 Ser	Pro Gly Phe Gln Gln 95 Asn	Leu Pro Asn 80 Arg	
1 Leu Glu Ala Val 65 Cys Val	<pre>&lt;2 &lt;2 &lt;2 &lt;4 Glu Val Gln Pro 50 Cys Pro Thr Asn Gly</pre>	211> 212> 213> 400> Ala Val Asn 35 Gly Arg Gln Gly Trp 115 His	1086 PRT Zea  18 Ser Ile 20 Gly Gly Asp Cys Asp 100 Asp	Mays Ala 5 Arg Gln Asp Cys Lys 85 Glu Gly	Gly Arg Val Pro Tyr 70 Thr Glu His	Asp Cys Phe 55 Glu Arg Glu Asp	Gly Gln 40 Val Tyr Tyr Asp Ser 120 Arg	Asp 25 Ile Ala Glu Lys Gly 105 Gln	10 Pro Cys Cys Arg Arg 90 Val	Gly Asn Arg 75 Leu Asp Val	Pro Asp Glu 60 Glu Lys Asp Ala	Lys Asp 45 Cys Gly Gly Leu Glu 125 Asn	Pro 30 Val Ala Thr Cys Asp 110 Ser	Phe Gln Gln 95 Asn	Arg Leu Pro Asn 80 Arg	
1 Leu Glu Ala Val 65 Cys Val Phe Tyr	<pre>&lt;2 &lt;2 &lt;2 &lt;4 Glu Val Gln Pro 50 Cys Pro Thr Asn Gly 130</pre>	211> 212> 213> 400> Ala Val Asn 35 Gly Arg Gln Gly Trp 115 His	1086 PRT Zea  18 Ser  Ile 20 Gly Gly Asp Cys Asp 100 Asp Met	Mays Ala 5 Arg Gln Asp Cys Lys 85 Glu Gly Ser	Gly Arg Val Pro Tyr 70 Thr Glu His Tyr Asn	Asp Cys Phe 55 Glu Arg Glu Asp	Gly Gln 40 Val Tyr Tyr Asp Ser 120 Arg	Asp 25 11e Ala Glu Lys Gly 105 Gln	10 Pro Cys Cys Arg Arg 90 Val Ser	Gly Asn Arg 75 Leu Asp Val Asp	Pro Asp Glu 60 Glu Lys Asp Ala Pro 140 Leu	Lys Asp 45 Cys Gly Gly Leu Glu 125 Asn	Pro 30 Val Ala Thr Cys Asp 110 Ser	Phe Gln Gln 95 Asn Met	Arg Leu Pro Asn 80 Arg Glu Leu Pro Gln	
1 Leu Glu Ala Val 65 Cys Val Phe Tyr Gln 145	<pre>&lt;2 &lt;2 &lt;2 &lt;4 Glu Val Gln Pro 50 Cys Pro Thr Asn Gly 130 Ala</pre>	211> 212> 213> 400> Ala Val Asn 35 Gly Arg Gln Gly Trp 115 His	1086 PRT Zea  18 Ser  Ile 20 Gly Gly Asp Cys Asp 100 Asp Met Gln	Mays Ala 5 Arg Gln Asp Cys Lys 85 Glu Gly Ser Leu	Gly Arg Val Pro Tyr 70 Thr Glu His Tyr Asn 150	Asp Cys Phe 55 Glu Arg Glu Asp Gly 135 Pro	Gly Gln 40 Val Tyr Tyr Asp Ser 120 Arg	Asp 25 11e Ala Glu Lys Gly 105 Gln Gly Val	10 Pro Cys Cys Arg 90 Val Ser Gly	Gly Gly Asn Arg 75 Leu Asp Val Asp	Pro Asp Glu 60 Glu Lys Asp Ala Pro 140 Leu	Lys Asp 45 Cys Gly Gly Leu Glu 125 Asn	Pro 30 Val Ala Thr Cys Asp 110 Ser Gly	Phe Gln Gln Gln 95 Asn Met	Arg Leu Pro Asn 80 Arg Glu Leu Pro	

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Met Gly Gly Gly Lys Arg Ile His Pro Leu Pro Tyr Ala Asp Pro 185 Ser Leu Pro Val Gln Pro Arg Ser Met Asp Pro Ser Lys Asp Leu Ala 200 Ala Tyr Gly Tyr Gly Ser Val Ala Trp Lys Glu Arg Met Glu Asn Trp 215 220 Lys Gln Arg Gln Glu Arg Met His Gln Thr Gly Asn Asp Gly Gly Gly 230 235 Asp Asp Gly Asp Asp Ala Asp Leu Pro Leu Met Asp Glu Ala Arg Gln 245 250 Gln Leu Ser Arg Lys Ile Pro Leu Pro Ser Ser Gln Ile Asn Pro Tyr 265 Arg Met Ile Ile Ile Arg Leu Val Val Leu Gly Phe Phe His 280 Tyr Arg Val Met His Pro Val Asn Asp Ala Phe Ala Leu Trp Leu Ile 295 300 Ser Val Ile Cys Glu Ile Trp Phe Ala Met Ser Trp Ile Leu Asp Gln 310 315 Phe Pro Lys Trp Phe Pro Ile Glu Arg Glu Thr Tyr Leu Asp Arg Leu 325 330 Ser Leu Arg Phe Asp Lys Glu Gly Gln Pro Ser Gln Leu Ala Pro Ile 345 Asp Phe Phe Val Ser Thr Val Asp Pro Leu Lys Glu Pro Pro Leu Val 360 Thr Thr Asn Thr Val Leu Ser Ile Leu Ser Val Asp Tyr Pro Val Asp 375 380 Lys Val Ser Cys Tyr Val Ser Asp Asp Gly Ala Ala Met Leu Thr Phe 390 395 Glu Ala Leu Ser Glu Thr Ser Glu Phe Ala Lys Lys Trp Val Pro Phe Cys Lys Arg Tyr Asn Ile Glu Pro Arg Ala Pro Glu Trp Tyr Phe Gln 420 425 Gln Lys Ile Asp Tyr Leu Lys Asp Lys Val Ala Ala Asn Phe Val Arg 440 Glu Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Ile 455 Asn Ala Leu Val Ala Lys Ala Gln Lys Val Pro Glu Glu Gly Trp Thr 470 475 Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn Val Arg Asp His Pro 490 Gly Met Ile Gln Val Phe Leu Gly Gln Ser Gly Gly Leu Asp Cys Glu Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro 520 Gly Tyr Asn His His Lys Lys Ala Gly Ala Met Asn Ala Leu Val Arg 535 Val Ser Ala Val Leu Thr Asn Ala Pro Tyr Leu Leu Asn Leu Asp Cys 550 555 Asp His Tyr Ile Asn Asn Ser Lys Ala Ile Lys Glu Ala Met Cys Phe 570 Met Met Asp Pro Leu Leu Gly Lys Lys Val Cys Tyr Val Gln Phe Pro 585 Gln Arg Phe Asp Gly Ile Asp Arg His Asp Arg Tyr Ala Asn Arg Asn 600 Val Val Phe Phe Asp Ile Asn Met Lys Gly Leu Asp Gly Ile Gln Gly 615 620 Pro Ile Tyr Val Gly Thr Gly Cys Val Phe Arg Arg Gln Ala Leu Tyr 630 635

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Gly Tyr Asp Ala Pro Lys Thr Lys Lys Pro Pro Ser Arg Thr Cys Asn 650 Cys Trp Pro Lys Trp Cys Phe Cys Cys Cys Cys Phe Gly Asn Arg Lys 665 660 Gln Lys Lys Thr Thr Lys Pro Lys Thr Glu Lys Lys Lys Leu Leu Phe 680 Phe Lys Lys Glu Glu Asn Gln Ser Pro Ala Tyr Ala Leu Gly Glu Ile 695 700 Asp Glu Ala Ala Pro Gly Ala Glu Asn Glu Lys Ala Gly Ile Val Asn 710 715 Gln Gln Lys Leu Glu Lys Lys Phe Gly Gln Ser Ser Val Phe Val Thr 730 725 Ser Thr Leu Leu Glu Asn Gly Gly Thr Leu Lys Ser Ala Ser Pro Ala 745 740 Ser Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Asp 760 765 Lys Thr Asp Trp Gly Lys Glu Ile Gly Trp Ile Tyr Gly Ser Val Thr 775 Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys His Gly Trp Arg Ser 795 790 Ile Tyr Cys Ile Pro Lys Arg Val Ala Phe Lys Gly Ser Ala Pro Leu 810 Asn Leu Ser Asp Arg Leu His Gln Val Leu Arg Trp Ala Leu Gly Ser 825 820 Ile Glu Ile Phe Phe Ser Asn His Cys Pro Leu Trp Tyr Gly Tyr Gly 840 Gly Gly Leu Lys Phe Leu Glu Arg Phe Ser Tyr Ile Asn Ser Ile Val 860 855 Tyr Pro Trp Thr Ser Ile Pro Leu Leu Ala Tyr Cys Thr Leu Pro Ala 870 875 880 Ile Cys Leu Leu Thr Gly Lys Phe Ile Thr Pro Glu Leu Asn Asn Val Ala Ser Leu Trp Phe Met Ser Leu Phe Ile Cys Ile Phe Ala Thr Ser 905 Ile Leu Glu Met Arg Trp Ser Gly Val Gly Ile Asp Asp Trp Trp Arg 915 920 Asn Glu Gln Phe Trp Val Ile Gly Gly Val Ser Ser His Leu Phe Ala 935 Val Phe Gln Gly Leu Leu Lys Val Ile Ala Gly Val Asp Thr Ser Phe 950 955 Thr Val Thr Ser Lys Gly Gly Asp Asp Glu Glu Phe Ser Glu Leu Tyr 965 970 Thr Phe Lys Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Leu Leu Leu 985 Leu Asn Phe Ile Gly Val Val Ala Gly Val Ser Asn Ala Ile Asn Asn 1000 Gly Tyr Glu Ser Trp Gly Pro Leu Phe Gly Lys Leu Phe Phe Ala Phe 1015 1020 Trp Val Ile Val His Leu Tyr Pro Phe Leu Lys Gly Leu Val Gly Arg 1035 1030 Gln Asn Arg Thr Pro Thr Ile Val Ile Val Trp Ser Ile Leu Leu Ala 1045 1050 Ser Ile Phe Ser Leu Leu Trp Val Arg Ile Asp Pro Phe Leu Ala Lys 1060 1065 Asp Asp Gly Pro Leu Leu Glu Glu Cys Gly Leu Asp Cys Asn 1080 1075

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<211> 25 <212> DNA <213> Zea mays <400> 19 atggaggcga gcgccgggct ggtgg <210> 20 <211> 25 <212> DNA <213> Zea mays <400> 20 ctagttgcaa tccaaaccac actcc 25 <210> 21 <211> 3725 <212> DNA <213> Zea mays <220> <221> CDS <222> (179)...(3398) <400> 21 gcagcagcag caccaccact gcgcggcatt gcagcgagca agcgggaggg atctggggca 60 120 tggtggcggt cgctgccgct gccgctcgga tctagagggc cgcacgggct gattgccctc 178 cgccggcctc gtcggtgtcg gtggagtgtg aatcggtgtg tgtaggagga gcgcggag 226 atg gcg gcc aac aag ggg atg gtg gca ggc tct cac aac cgc aac gag Met Ala Ala Asn Lys Gly Met Val Ala Gly Ser His Asn Arg Asn Glu 5 10 274 tte qte atq ate eqe cae qae qqe gae geg cet gte eeg get aag eec Phe Val Met Ile Arg His Asp Gly Asp Ala Pro Val Pro Ala Lys Pro 25 322 acg aag agt gcg aat ggg cag gtc tgc cag att tgt ggc gac act gtt Thr Lys Ser Ala Asn Gly Gln Val Cys Gln Ile Cys Gly Asp Thr Val 35 qqc qtt tca qcc act qqt qat qtc ttt gtt gcc tgc aat gag tgt gcc 370 Gly Val Ser Ala Thr Gly Asp Val Phe Val Ala Cys Asn Glu Cys Ala 50 55 418 tte eet gte tge ege eet tge tat gag tae gag ege aag gaa ggg aac Phe Pro Val Cys Arg Pro Cys Tyr Glu Tyr Glu Arg Lys Glu Gly Asn 65 70 466 caa tgc tgc cct cag tgc aag act aga tac aag aga cag aaa ggt agc Gln Cys Cys Pro Gln Cys Lys Thr Arg Tyr Lys Arg Gln Lys Gly Ser 85 514 Pro Arg Val His Gly Asp Asp Glu Glu Asp Val Asp Asp Leu Asp 105

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aat Asn	gaa Glu	ttc Phe 115	aac Asn	tat Tyr	aag Lys	Gln	ggc Gly 120	aat Asn	gj aaa	aag Lys	ggc ggc	cca Pro 125	gag Glu	tgg Trp	cag Gln	562
ctt Leu	caa Gln 130	gga Gly	gat Asp	gac Asp	gct Ala	gat Asp 135	ctg Leu	tct Ser	tca Ser	tct Ser	gct Ala 140	cgc Arg	cat His	gac Asp	cca Pro	610
cac His 145	cat His	cgg Arg	att Ile	cca Pro	cgc Arg 150	ctt Leu	aca Thr	agt Ser	gga Gly	caa Gln 155	cag Gln	ata Ile	tct Ser	gga Gly	gag Glu 160	658
atc Ile	cct Pro	gat Asp	gca Ala	tcc Ser 165	cct Pro	gac Asp	cgt Arg.	cat His	tct Ser 170	atc Ile	cgc Arg	agt Ser	cca Pro	aca Thr 175	tcg Ser	706
agc Ser	tat Tyr	gtt Val	gat Asp 180	cca Pro	agc Ser	gtt Val	cca Pro	gtt Val 185	cct Pro	gtg Val	agg Arg	att Ile	gtg Val 190	gac Asp	ccc Pro	754
tcg Ser	aag Lys	gac Asp 195	ttg Leu	aat Asn	tcc Ser	tat Tyr	999 900	Leu	aat Asn	agt Ser	gtt Val	gac Asp 205	tgg Trp	aag Lys	gaa Glu	802
aga Arg	gtt Val 210	Glu	agc Ser	tgg Trp	agg Arg	gtt Val 215	aaa Lys	cag Gln	gac Asp	aaa Lys	aat Asn 220	Met	ttg Leu	caa Gln	gtg Val	850
act Thr 225	Asn	aaa Lys	tat Tyr	cca Pro	gag Glu 230	Ala	aga Arg	gga Gly	gac Asp	atg Met 235	Glu	ggg Gly	act Thr	ggc	tca Ser 240	898
aat Asn	gga Gly	gaa Glu	ı gat ı Asp	atg Met 245	Gln	atg Met	gtt Val	gat Asp	gat Asp 250	Ala	cgc Arg	cta Leu	cct Pro	ttg Leu 255	agc Ser	946
cgc Arg	att JIle	gtg Val	g cca L Pro 260	Il∈	tcc Ser	tca Ser	aac Asr	cag Glr 265	ı Lev	aac Asr	ctt Lei	tac ı Tyr	e cgg Arg 270	Ile	gta Val	994
ato Ile	c att	cto Let 27	Arg	ctt Lei	ato 1 Ile	ato lle	cto Lev 280	a Cys	tto Phe	tto Phe	tto Phe	c caa e Glr 285	туг	cgt Arg	atc Ile	1042
agt Sei	cat r His 290	s Pro	a gtg o Val	g egt L Arg	aat g Asi	get n Ala 299	ту:	gga r Gl	a ttg y Lei	g tgg ı Tr]	g cta o Lei 30	u Val	a tct L Sei	gtt Val	atc l Ile	1090
tg: Cy: 30:	s Gl	g gt u Va	c tgg l Trj	g tt:	t gco e Ala 31	a Lei	g to 1 Se:	c tgg r Trj	g cti p Lei	t cta 1 Lem 31	u As	t cag p Gli	g tto n Phe	e cca	a aaa b Lys 320	1138
tg:	g ta p Ty	t cc r Pr	a ato	c aa e As: 32	n Ar	t gag g Gl	g ac	a ta r Ty	t cto r Leo 33	u As	c ag p Ar	g cti g Lei	t gca u Ala	a ttg a Lei 33!	g agg u Arg 5	1186
ta Ty	t ga r As	t ag p Ar	a ga g Gl	g gg u Gl	a ga y Gl	g cc u Pr	a tc o Se	a ca r Gl	g ct n Le	g gc u Al	t cc a Pr	c at	t ga e As	t gte p Va	c ttt l Phe	1234

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			340					345					350				
-	_			_		-	_	_			_			gcc Ala		=	1282
	-	-				_		_			_	•		gtg Val		:	
_		_		_	_			_	_	-				tct Ser		:	1378
	-		-	_		_	_	_		_			_	aag Lys 415	_	=	1426
			_		_	_		-				_		aaa Lys		•	1474
-		_	_	-		_	_				_	_	_	aga Arg	_	:	1522
														gcc Ala		:	1570
									-				_	gct Ala	-	:	1618
										_				atg Met 495		:	1666
						-				_		_		aat Asn	_	:	1714
		_		-		_		_	-	_	_			ttt Phe	_	:	1762
		_	_	_		_	_		_	_		_	_	tct Ser	_		1810
														cat His			1858
		-	_		_		_	_	-	_	_		_	atg Met 575	_		1906

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	gct Ala															1954
	ggc															2002
	gat Asp 610			_				_			_			_		2050
	gga Gly															2098
	gtt Val	_		_	_	_	_	_				_	_	_	_	2146
	tgt Cys															2194
	cgt Arg									_			_			2242
	gac Asp 690															2290
	atg Met		_			_			_							2338
	att Ile	_				_										2386
	cca Pro	-			_	_	_	_			_		-			2434
	gag Glu		Lys												ggt Gly	2482
	gtt Val 770	Thr		_		-					_	His			ggc	2530
						Met					Cys				Ser 800	2578
_						-	_			-					gct Ala	2626

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				805					810					815		
ctt Leu																2674
												gct Ala 845				2722
												gcc Ala				2770
												att Ile				2818
												gcc Ala				2866
												ggc				2914
			Asn									acc Thr 925				2962
		Ala										gct Ala				3010
											Glu	gat Asp				3058
					Phe					Leu		atc Ile			Thr	3106
				Ile					Met					Ser	tat Tyr	3154
gcc Ala	att Ile	aac Asr 99	ı Ser	ggc Gly	tac Tyr	caa Glr	tcc Ser 100	Trp	ggt Gly	ccg Pro	cto Lev	ttt Phe	Gly	aag Lys	ctg Leu	3202
		e Sei					Lev					) Phe			ggt Gly	3250
	ı Met					ı Arg					≥ Va				s tcc Ser 1040	3298

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																2245
atc			Ala S					Leu 1					Ile 2			3346
ttc Phe		Ser 1					Ala .					Gln				3394
aac Asn	t gc	tgato	ccag	att	gtga	ctc	ttat	ctga	ag a	ggct	cagc	c aa	agat	ctgc		3448
tctg atca catt	catc tgga tttt	ca a	gtta tttc acgt	tgcc tacc ggtg	t ct t tg t tt	gttt cttg attg	atta tagt tttt	gct gct aga	tctt ggcc gtaa	cgg agc att	tgcc agcg	ggtg taaa	ct g tt g	ctgc tgaa	atgga agaca ttctg taact	3508 3568 3628 3688 3725
		10>														
		11> 12>														
		13>		mays	;											
				_												
Met 1		00> Ala		Lys 5	Gly	Met	Val	Ala	Gly 10	Ser	His	Asn	Arg	Asn 15	Glu	
	Val	Met	Ile		His	Asp	Gly	Asp		Pro	Val	Pro	Ala		Pro	
_,	_		20	•	<b>0</b> 1	a1 -	17- 7	25	<b>71</b> -	T1.	C	C3	30	Thr	Val	
Thr	Lys	Ser 35	Ala	Asn	GLY	GIN	va1 40	Cys	GIN	пе	Cys	45	ASD	1111	Val	
_	50	Ser				55					60					
	Pro	Val	Cys	Arg	Pro 70	Cys	Tyr	Glu	Tyr	Glu 75	Arg	Lys	Glu	Gly	Asn 80	
65 Gln	Cys	Суз	Pro	Gln 85		Lys	Thr	Arg	Tyr 90	-	Arg	Gln	Lys	Gly 95		
Pro	Arg	Val		Gly	Asp	Asp	Glu		Glu	Asp	Val	Asp		Leu	Asp	
Acn	Glu	Phe	100 Asn	Tvr	Lvs	Gln	Glv	105 Asn	Glv	Lvs	Glv	Pro	110 Glu	Trp	Gln	
		115					120					125				
	130	Gly				135					140					
		Arg	Ile	Pro		Leu	Thr	Ser	Gly			Ile	Ser	Gly	Glu 160	
145 Ile		Asp	Ala	Ser 165	150 Pro	Asp	Arg	His	Ser 170	155 Ile		Ser	Pro	Thr 175		
Ser	Tyr	Val	Asp		Ser	Val	Pro	Val 185		Val	Arg	Ile	Val 190		Pro	
Ser	Lys	Asp 195		Asn	Ser	Tyr	Gly 200	Leu	Asn	Ser	Val	Asp 205		Lys	Glu	
Arg	Val 210	Glu	Ser	Trp	Arg	Val 215			Asp	Lys	Asn 220		Leu	Gln	Val	
Thr		Lys	Tyr	Pro	Glu		Arg	Gly	Asp			Gly	Thr	Gly		
225		<b>03</b>	n	3.6 - A	230		17-1	7 ~	7 am	235		T.eur	Dro	T.e.	240 Ser	
Asn	GIY	Glu	Asp	Met 245		met	val	Asp	250		arg	TEU	FIO	255		
Arg	Ile	Val	Pro 260	Ile	Ser	Ser	Asn	Gln 265		Asn	Leu	Tyr	Arg 270		Val	

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Ile	Ile	Leu	Arg	Leu	Ile	Ile	Leu	Cys	Phe	Phe	Phe	Gln	Tyr	Arg	Ile
		275					280					285			
	290					295					300			Val	
	Glu	Val	Trp	Phe		Leu	Ser	Trp	Leu	Leu 315	Asp	GIn	Phe	Pro :	Lуs 320
305	ጥ፣፣፦	Dro	Tla	λen	310 Arg	Glu	Thr	ጥvr	Leu		Ara	Leu	Ala	Leu .	
ırp	IYL	FIO	110	325	y	<u> </u>		-1-	330		5			335	
_			340					345					350	Val	
		355					360					365		Ala	
	370					375					380			Val	
385					390					395				Ser	400
				405					410					Lys 415	
			420					425					430	Lys	
		435					440					445		Arg	
	450					455					460			Ala	
465					470					475				Ala	480
Gly	Thr	Ala	Trp	Pro 485		Asn	Asn	Pro	Arg 490		His	Pro	Gly	Met 495	Ile
			500					505					510	Asn	
		515					520	H				525		Phe	
	530					535	;				540			Ser	
545					550	)				555				His	560
				565					570	)				Met 575	
			580	)				585	;				590		
		595	5				600	)				605		Val	
	610	)				615	5				620	)			Tyr
		Thi	Gly	Cys			a Ası	1 Arg	g Glr	n Ala 635		Tyr	Gly	Tyr	Asp 640
625		I.e.	ı Thi	- Gli	630 1 Ala		. Lei	ı Glı	ı Pro			. Val	. Val	Lys	Ser
				645	5				650	)				655	
			660	)				66	5				670	1	
		67	5				686	0				685	5		Met
	690	)				69	5				700	)			Val
705	5				71	0				71	5				720
Ph€	e Ile	e Ala	a Se	r Th: 72!		e Me	t Th	r Gl	n Gl <sup>-</sup> 73		y Ile	e Pro	o Pro	735	Thr

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Asn Pro Ala Ser Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys Gly 745 Tyr Glu Asp Lys Thr Glu Trp Gly Lys Glu Ile Gly Trp Ile Tyr Gly 760 Ser Val Thr Glu Asp Ile Leu Thr Gly Phe Lys Met His Ala Arg Gly 775 Trp Gln Ser Ile Tyr Cys Met Pro Pro Arg Pro Cys Phe Lys Gly Ser 790 795 Ala Pro Ile Asn Leu Ser Asp Arg Leu Asn Gln Val Leu Arg Trp Ala 805 810 Leu Gly Ser Val Glu Ile Leu Leu Ser Arg His Cys Pro Ile Trp Tyr 820 825 Gly Tyr Asn Gly Arg Leu Lys Leu Leu Glu Arg Leu Ala Tyr Ile Asn 835 840 Thr Ile Val Tyr Pro Ile Thr Ser Val Pro Leu Ile Ala Tyr Cys Val 855 860 Leu Pro Ala Ile Cys Leu Leu Thr Asn Lys Phe Ile Ile Pro Glu Ile 865 870 875 Ser Asn Tyr Ala Gly Met Phe Phe Ile Leu Leu Phe Ala Ser Ile Phe 890 895 Ala Thr Gly Ile Leu Glu Leu Arg Trp Ser Gly Val Gly Ile Glu Asp 905 Trp Trp Arg Asn Glu Gln Phe Trp Val Ile Gly Gly Thr Ser Ala His 920 Leu Phe Ala Val Phe Gln Gly Leu Leu Lys Val Leu Ala Gly Ile Asp 935 Thr Asn Phe Thr Val Thr Ser Lys Ala Ser Asp Glu Asp Gly Asp Phe 955 950 Ala Glu Leu Tyr Val Phe Lys Trp Thr Ser Leu Leu Ile Pro Pro Thr 965 970 Thr Val Leu Val Ile Asn Leu Val Gly Met Val Ala Gly Ile Ser Tyr 980 985 Ala Ile Asn Ser Gly Tyr Gln Ser Trp Gly Pro Leu Phe Gly Lys Leu 1000 1005 Phe Phe Ser Ile Trp Val Ile Leu His Leu Tyr Pro Phe Leu Lys Gly 1010 1015 1020 Leu Met Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Ile Val Trp Ser 1030 1035 1040 Ile Leu Leu Ala Ser Ile Phe Ser Leu Leu Trp Val Lys Ile Asp Pro 1050 1045 Phe Ile Ser Pro Thr Gln Lys Ala Ala Ala Leu Gly Gln Cys Gly Val 1065 Asn Cys

<210> 23

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<212> DNA

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<400> 23

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<210> 24

<211> 25

<212> DNA

<213> Zea mays

667

<400> 24 tcagcagttc acaccacatt gcccc 25

<210> 25
<211> 3813
<212> DNA
<213> Zea mays
<220>
<221> CDS
<222> (215)...(3494)

agc cca	cagc acaa gagg	agg (	tata ccac ggag	ccgc gact	ca ca ac gi	agcc: tgcai	gegte	g cgo	catgr gtgc ag a M	tgag cgcc tg g	ag g gcc: cgg	tege geggg et a	ege g ggt ge g er A	ggcc: tcgt: cg g:	gegeca gggaga ge etg gy Leu	1	60 .20 .80 :35
gtg Val	gcc Ala	ggc Gly 10	tcg Ser	cat His	aac Asn	cgg Arg	aac Asn 15	gag Glu	ctg Leu	gtg Val	gtg Val	atc Ile 20	cgc Arg	cgc Arg	gac Asp	2	83
cgc Arg	gag Glu 25	tcg Ser	gga Gly	gcc Ala	gcg Ala	ggc Gly 30	Gly	ggc	gcg Ala	gcg Ala	cgc Arg 35	cgg Arg	gcg Ala	gag Glu	gcg Ala	3	31
ecg Pro 40	tgc Cys	cag Gln	ata Ile	tgc Cys	ggc Gly 45	gac Asp	gag Glu	gtc Val	gjå aaa	gtg Val 50	ggc Gly	ttc Phe	gac Asp	Gly aaa	gag Glu 55	3	79
ccc Pro	ttc Phe	gtg Val	gcg Ala	tgc Cys 60	aac Asn	gag Glu	tgc Cys	gcc Ala	ttc Phe 65	ccc Pro	gtc Val	tgc Cys	cgc Arg	gcc Ala 70	tgc Cys	4	27
tac Tyr	gag Glu	tac Tyr	gag Glu 75	cgc Arg	cgc Arg	gag Glu	ggc Gly	tcg Ser 80	caa Gln	gcg Ala	tgc Cys	ccg Pro	cag Gln 85	tgc Cys	agg Arg	4	75
acc	cgc Arg	tac Tyr 90	aag Lys	cgc Arg	ctc Leu	aag Lys	ggc Gly 95	tgc Cys	ccg Pro	cgg Arg	gtg Val	gcc Ala 100	gly	gac Asp	gag Glu	5	23
gag Glu	gag Glu 105	gac Asp	ggc Gly	gtc Val	gac Asp	gac Asp 110	ctg Leu	gag Glu	ggc	gag Glu	ttc Phe 115	ggc Gly	ctg Leu	cag Gln	gac Asp	5	71
ggc Gly 120	gcc Ala	gcc Ala	cac His	gag Glu	gac Asp 125	gac Asp	ccg Pro	cag Gln	tac Tyr	gtc Val 130	gcc Ala	gag Glu	tcc Ser	atg Met	ctc Leu 135	6	19

agg gcg cag atg agc tac ggc cgc ggc ggc gac gcg cac ccc ggc ttc

Arg Ala Gln Met Ser Tyr Gly Arg Gly Gly Asp Ala His Pro Gly Phe

140

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_	ccc Pro	_				_						_	-	-	-	715
-	atc Ile	_	_		_				_	_			_	_		763
	ggc Gly 185				-		_		_				_	_		811
	ctt Leu				_	-		_	_	_		_	-		-	859
-	tac Tyr				_		_		_		_	-				907
-	cag Gln	-	_		_	_	_		_		_					955
_	tgg Trp	-		-	_	_	_	_			-	_	_	_		1003
_	cca Pro 265	_		_		_					_					1051
	agg Arg	_			-			_		_	_					1099
	tac Tyr	_		_		_			-	-		-	_		ctc Leu	1147
	tct Ser	-		_	-					_					-	1195
															cgt Arg	1243
_					-	_	_					_		-	cca Pro	1291
	Asp			_	-	-	-	_			Lys	-			ttg Leu 375	1339
-					-							-		_	gtt Val	1387

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								- ,	0 -							
				380					385					390		
		-										gca Ala			_	1435
												aaa Lys 420				1483
	_		_						_	-		gag Glu				1531
	_	_		_		_						gct Ala				1579
												ttc Phe				1627
												gag Glu				1675
												gta Val 500				1723
												ggt Gly				1771
-	Gly			_		_	_	_		_	_	aga Arg				1819
			Asn		His	Lys	Lys	Ala		Ala		aat Asn			Val	1867
-	_		_	Val				_	Ala			ttg Leu		Leu		1915
			Tyr					Lys				gag Glu 580	Ala			1963
		Met					Gly					Tyr			ttc Phe	2011
	Gln					Ile					Arg				agg Arg 615	2059

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aac Asn	gtt Val	gtc Val	ttt Phe	ttt Phe 620	gac Asp	atc Ile	aac Asn	atg Met	aaa Lys 625	ggt Gly	ttg Leu	gac Asp	ggt Gly	att Ile 630	caa Gln	2107
gga Gly	ccc Pro	att Ile	tat Tyr 635	gtg Val	ggt Gly	act Thr	gga Gly	tgt Cys 640	gtt Val	ttc Phe	aga Arg	cgg Arg	cag Gln 645	gca Ala	ctg Leu	2155
tat Tyr	ggt Gly	tat Tyr 650	gat Asp	gct Ala	cct Pro	aaa Lys	acg Thr 655	aag Lys	aag Lys	cca Pro	cca Pro	tca Ser 660	aga Arg	act Thr	tgc Cys	2203
aac Asn	tgc Cys 665	tgg Trp	ccc Pro	aag Lys	tgg Trp	tgc Cys 670	ctc Leu	tct Ser	tgc Cys	tgc Cys	tgc Cys 675	agc Ser	agg Arg	aac Asn	aag Lys	2251
aat Asn 680	aaa Lys	aag Lys	aag Lys	act Thr	aca Thr 685	aaa Lys	cca Pro	aag Lys	acg Thr	gag Glu 690	aag Lys	aag Lys	aaa Lys	aga Arg	tta Leu 695	2299
ttt Phe	ttc Phe	aag Lys	aaa Lys	gca Ala 700	gaa Glu	aac Asn	cca Pro	tct Ser	cct Pro 705	gca Ala	tat Tyr	gct Ala	ttg Leu	ggt Gly 710	gaa Glu	2347
att Ile	gat Asp	gaa Glu	ggt Gly 715	gct Ala	cca Pro	ggt Gly	gct Ala	gat Asp 720	atc Ile	gag Glu	aag Lys	gcc Ala	gga Gly 725	atc Ile	gta Val	2395
aat Asn	caa Gln	cag Gln 730	aaa Lys	cta Leu	gag Glu	aag Lys	aaa Lys 735	ttt Phe	gly aaa	cag Gln	tct Ser	tct Ser 740	gtt Val	ttt Phe	gtc Val	2443
gca Ala	tca Ser 745	aca Thr	ctt Leu	ctt Leu	gag Glu	aac Asn 750	gga Gly	gly aaa	acc Thr	ctg Leu	aag Lys 755	agc Ser	gca Ala	agt Ser	cca Pro	2491
gct Ala 760	tct Ser	ctt Leu	ctg Leu	aag Lys	gaa Glu 765	gct Ala	ata Ile	cat His	gtt Val	atc Ile 770	agc Ser	tgc Cys	ggc	tac Tyr	gaa Glu 775	2539
gac Asp	aag Lys	acc Thr	gac Asp	tgg Trp 780	gga Gly	aaa Lys	gag Glu	att Ile	ggc Gly 785	tgg Trp	att Ile	tac Tyr	gga Gly	tcg Ser 790	atc Ile	2587
aca Thr	gag Glu	gat Asp	atc Ile 795	ttg Leu	act Thr	gga Gly	ttt Phe	aag Lys 800	atg Met	cac His	tgc Cys	cat His	ggc Gly 805	tgg Trp	cgg Arg	2635
tct Ser	att Ile	tac Tyr 810	tgc Cys	atc Ile	ccg Pro	aag Lys	cgg Arg 815	cct Pro	gca Ala	ttc Phe	aaa Lys	ggt Gly 820	tct Ser	gcg Ala	cct Pro	2683
ctg Leu	aac Asn 825	ctt Leu	tcc Ser	gac Asp	cgt Arg	ctt Leu 830	cac His	cag Gln	gtc Val	ctt Leu	cgc Arg 835	tgg Trp	gcc Ala	ctt Leu	ggg ggg	2731
tcc Ser	gtc Val	gaa Glu	att Ile	ttc Phe	ttc Phe	agc Ser	aag Lys	cac His	tgc Cys	cca Pro	ctt Leu	tgg Trp	tac Tyr	gga Gly	tac Tyr	2779

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840	845	850	855
ggc ggc ggg cta aaa Gly Gly Gly Leu Lys 860	ttc ctg gaa agg ttt Phe Leu Glu Arg Phe 865	tct tat atc aac tcc Ser Tyr Ile Asn Ser 870	Ile
gtt tat ccc tgg acg Val Tyr Pro Trp Thr 875	tcc att cct ctc ctg Ser Ile Pro Leu Leu 880	gct tac tgt acc ttg Ala Tyr Cys Thr Leu 885	cct 2875 Pro
gcc atc tgc ctg ctc Ala Ile Cys Leu Leu 890	acg ggg aag ttt atc Thr Gly Lys Phe Ile 895	aca cca gag ctt acc Thr Pro Glu Leu Thr 900	aat 2923 Asn
gtc gcc agt atc tgg Val Ala Ser Ile Trp 905	ttc atg gca ctt ttc Phe Met Ala Leu Phe 910	atc tgc atc tcc gtg Ile Cys Ile Ser Val 915	acc 2971 Thr
ggc atc ctg gaa atg Gly Ile Leu Glu Met 920	agg tgg agt ggc gtg Arg Trp Ser Gly Val 925	gcc atc gac gac tgg Ala Ile Asp Asp Trp 930	tgg 3019 Trp 935
agg aac gag cag ttc Arg Asn Glu Gln Phe 940	tgg gtc atc gga ggc Trp Val Ile Gly Gly 945	gtt tcg gcg cat ctg Val Ser Ala His Leu 950	Phe
gcg gtg ttc cag ggc Ala Val Phe Gln Gly 955	ctg ctg aag gtg ttc Leu Leu Lys Val Phe 960	gcc ggc atc gac acg Ala Gly Ile Asp Thr 965	agc 3115 Ser
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tac acg ttc aag tgg Tyr Thr Phe Lys Trp 985	acc acc ctg ctg ata Thr Thr Leu Leu Ile 990	ccc ccg acc acg ctc Pro Pro Thr Thr Leu 995	ctc 3211 Leu
ctg ctg aac ttc atc Leu Leu Asn Phe Ile 1000	ggg gtg gtg gcc ggg Gly Val Val Ala Gly 1005	atc tcg aac gcg atc Ile Ser Asn Ala Ile 1010	aac 3259 Asn 1015
aac ggg tac gag tcg Asn Gly Tyr Glu Ser 1020	Trp Gly Pro Leu Phe	Gly Lys Leu Phe Phe	Ala
ttc tgg gtg atc gtc Phe Trp Val Ile Val 1035	cac ctg tac ccg ttc His Leu Tyr Pro Phe 1040	ctc aag ggt ctg gtg Leu Lys Gly Leu Val 1045	Gly ggg 3355
agg cag aac agg acg Arg Gln Asn Arg Thr 1050	ccg acg atc gtc atc Pro Thr Ile Val Ile 1055	gtc tgg tcc atc ctg Val Trp Ser Ile Leu 1060	ctg 3403 Leu
gcc tcg atc ttc tcg Ala Ser Ile Phe Ser 1065	ctc ctg tgg gtc cgc Leu Leu Trp Val Arg 1070	gtc gac ccg ttc ctc Val Asp Pro Phe Leu 1075	gcc 3451 Ala

3494

3554

3614

3674

3734

3794

3813

aag age aac gge ceg ete etg gag gag tgt gge etg gae tge a Lys Ser Asn Gly Pro Leu Leu Glu Glu Cys Gly Leu Asp Cys 1085 1080 1090 actgaagtgg gggccccctg tcactcgaag ttctgtcacg ggcgaattac gcctgatttt ttgttgttgt tgttgttgga attctttgct gtagatagaa accacatgtc cacggcatct ctgctgtgtc cattggagca ggagagaggt gcctgctgct gtttgttgag taaattaaaa gttttaaagt tatacagtga tgcacattcc agtgcccagt gtattccctt tttacagtct gtatattagc gacaaaggac atattggtta ggagtttgat tcttttgtaa aaaaaaaaa aaaaaaaaa aaaaaaaaa <210> 26 <211> 1094 <212> PRT <213> Zea mays <400> 26 Met Glu Ala Ser Ala Gly Leu Val Ala Gly Ser His Asn Arg Asn Glu 10 Leu Val Val Ile Arg Arg Asp Arg Glu Ser Gly Ala Ala Gly Gly 25 Ala Ala Arg Arg Ala Glu Ala Pro Cys Gln Ile Cys Gly Asp Glu Val 40 Gly Val Gly Phe Asp Gly Glu Pro Phe Val Ala Cys Asn Glu Cys Ala 55 Phe Pro Val Cys Arg Ala Cys Tyr Glu Tyr Glu Arg Arg Glu Gly Ser Gln Ala Cys Pro Gln Cys Arg Thr Arg Tyr Lys Arg Leu Lys Gly Cys 90 Pro Arg Val Ala Gly Asp Glu Glu Glu Asp Gly Val Asp Asp Leu Glu 105 110 Gly Glu Phe Gly Leu Gln Asp Gly Ala Ala His Glu Asp Asp Pro Gln 120 Tyr Val Ala Glu Ser Met Leu Arg Ala Gln Met Ser Tyr Gly Arg Gly 135 140 Gly Asp Ala His Pro Gly Phe Ser Pro Val Pro Asn Val Pro Leu Leu 150 155 Thr Asn Gly Gln Met Val Asp Asp Ile Pro Pro Glu Gln His Ala Leu 165 170 Val Pro Ser Tyr Met Ser Gly Gly Gly Gly Gly Lys Arg Ile His 180 185 Pro Leu Pro Phe Ala Asp Pro Asn Leu Pro Val Gln Pro Arg Ser Met 200 Asp Pro Ser Lys Asp Leu Ala Ala Tyr Gly Tyr Gly Ser Val Ala Trp 215 220 Lys Glu Arg Met Glu Gly Trp Lys Gln Lys Gln Glu Arg Leu Gln His 230 235 Val Arg Ser Glu Gly Gly Asp Trp Asp Gly Asp Asp Ala Asp Leu 245 Pro Leu Met Asp Glu Ala Arg Gln Pro Leu Ser Arg Lys Val Pro Ile 260 265 270 Ser Ser Ser Arg Ile Asn Pro Tyr Arg Met Ile Ile Val Ile Arg Leu 280 Val Val Leu Gly Phe Phe Phe His Tyr Arg Val Met His Pro Ala Lys 295 Asp Ala Phe Ala Leu Trp Leu Ile Ser Val Ile Cys Glu Ile Trp Phe 315 Ala Met Ser Trp Ile Leu Asp Gln Phe Pro Lys Trp Leu Pro Ile Glu

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	325				330					335	
Arg Glu Thi				Ser 345	Leu .	Arg	Phe	Asp	Lys 350	Glu	Gly
Gln Pro Ser		Ala Pro	360	Asp	Phe	Phe	Val	Ser 365	Thr	Val	Asp
Pro Thr Lys		375	5				380				
Leu Ser Val		390				395					400
Asp Gly Ala	405	,			410					415	
Phe Ala Ly	420			425					430		
Arg Ala Pro	5	_	440					445			
Lys Val Al 450		45	5				460				
Tyr Glu Gl 465		470				475					480
Lys Val Pr	489	5			490					495	
Gly Asn As	500			505					510		
Gln Ser Gl	5		520					525			
Tyr Val Se		53	5				540				
Gly Ala Me 545 Ala Tyr Le		550				555					560
Ala Tyr Le	.56	5			570					575	
Lys Val Cy	580			585					590		
59 Asn Asp Ar	5		600					605			
610 Lys Gly Le		61	5				620				
625 Val Phe Ar		630				635					640
Lys Pro Pi	64	5			650					655	
Cys Cys Cy	660			665					670	ļ.	
67 Thr Glu Ly	15		680					685			
690 Pro Ala Ty		69	5				700				
705		710				715					720
Ile Glu Ly	72	5			730					735	i
Gly Gln Se	740			745					750	)	
	55		760	)				765	;		
Val Ile So 770		7	75				780	)			
Gly Trp I	re Tyr Gi	y ser 1.	le Inr	. GIV	ı Asp		. ner	Y TITE	. 31)	, Elle	. Lys

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790
Met His Cys His Gly Trp Arg Ser Ile Tyr Cys Ile Pro Lys Arg Pro
           805 810 815
Ala Phe Lys Gly Ser Ala Pro Leu Asn Leu Ser Asp Arg Leu His Gln
 820 825
Val Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Phe Phe Ser Lys His
     835 840
Cys Pro Leu Trp Tyr Gly Tyr Gly Gly Gly Leu Lys Phe Leu Glu Arg
 850 855 860
Phe Ser Tyr Ile Asn Ser Ile Val Tyr Pro Trp Thr Ser Ile Pro Leu
            870 875 880
Leu Ala Tyr Cys Thr Leu Pro Ala Ile Cys Leu Leu Thr Gly Lys Phe
           885 890 895
Ile Thr Pro Glu Leu Thr Asn Val Ala Ser Ile Trp Phe Met Ala Leu
        900 905
Phe Ile Cys Ile Ser Val Thr Gly Ile Leu Glu Met Arg Trp Ser Gly
     915 920
Val Ala Ile Asp Asp Trp Trp Arg Asn Glu Gln Phe Trp Val Ile Gly
                 935
                                940
Gly Val Ser Ala His Leu Phe Ala Val Phe Gln Gly Leu Leu Lys Val
    950 955
Phe Ala Gly Ile Asp Thr Ser Phe Thr Val Thr Ser Lys Ala Gly Asp
         965 970 975
Asp Glu Glu Phe Ser Glu Leu Tyr Thr Phe Lys Trp Thr Thr Leu Leu
        980 985 990
Ile Pro Pro Thr Thr Leu Leu Leu Asn Phe Ile Gly Val Val Ala
     995 1000 1005
Gly Ile Ser Asn Ala Ile Asn Asn Gly Tyr Glu Ser Trp Gly Pro Leu
  1010 1015 1020
Phe Gly Lys Leu Phe Phe Ala Phe Trp Val Ile Val His Leu Tyr Pro
1025 1030 1035 1040
Phe Leu Lys Gly Leu Val Gly Arg Gln Asn Arg Thr Pro Thr Ile Val
          1045 1050 1055
Ile Val Trp Ser Ile Leu Leu Ala Ser Ile Phe Ser Leu Leu Trp Val
        1060 1065 1070
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                                   1085
Cys Gly Leu Asp Cys Asn
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25

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      <221> CDS
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                                                                    60
cgacagcgac agcggaacca actcacgttg ccgcggcttc ctccatcggt gcggtgccct
                                                                   120
180
actagcagca gcagcgctct cgcagcggga gatgcggtgt tgatccgtgc cccgctcgga
                                                                   240
tetegggaet ggtgeegget etgeeeagge eccaggetee aggeeagete cetegaegtt
                                                                   300
teteggegag etegettgee atg gag gge gae geg gae gge gtg aag teg ggg
                                                                   353
                      Met Glu Gly Asp Ala Asp Gly Val Lys Ser Gly
agg cgc ggt ggc gga cag gtg tgc cag atc tgc ggc gac ggc gtg ggc
                                                                   401
Arg Arg Gly Gly Gln Val Cys Gln Ile Cys Gly Asp Gly Val Gly
acc acg gcg gag ggg gac gtc ttc gcc gcc tgc gac gtc tgc ggg ttt
                                                                   449
Thr Thr Ala Glu Gly Asp Val Phe Ala Ala Cys Asp Val Cys Gly Phe
                            35
ceg gtg tgc ege eee tgc tac gag tac gag ege aag gac ggc acg cag
                                                                   497
Pro Val Cys Arg Pro Cys Tyr Glu Tyr Glu Arg Lys Asp Gly Thr Gln
     45
geg tge cee cag tge aag ace aag tae aag cge cae aag ggg age ceg
                                                                   545
Ala Cys Pro Gln Cys Lys Thr Lys Tyr Lys Arg His Lys Gly Ser Pro
60
gcg atc cgt ggg gag gaa gga gac gac act gat gcc qat aqc qac ttc
                                                                   593
Ala Ile Arg Gly Glu Glu Gly Asp Asp Thr Asp Ala Asp Ser Asp Phe
                80
aat tac ctt gca tct ggc aat gag gac cag aag cag aag att gcc gac
                                                                   641
Asn Tyr Leu Ala Ser Gly Asn Glu Asp Gln Lys Gln Lys Ile Ala Asp
            95
aga atg cgc agc tgg cgc atg aac gtt ggg ggc agc ggg gat gtt ggt
                                                                   689
Arg Met Arg Ser Trp Arg Met Asn Val Gly Gly Ser Gly Asp Val Gly
       110
cgc ccc aag tat gac agt ggc gag atc ggg ctt acc aag tat gac agt
                                                                   737
Arg Pro Lys Tyr Asp Ser Gly Glu Ile Gly Leu Thr Lys Tyr Asp Ser
    125
                       130
ggc gag att cct cgg gga tac atc cca tca gtc act aac agc cag atc
                                                                   785
Gly Glu Ile Pro Arg Gly Tyr Ile Pro Ser Val Thr Asn Ser Gln Ile
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                                       150
tca gga gaa atc cct ggt gct tcc cct gac cat cat atg atg tcc cca
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Ser Gly Glu Ile Pro Gly Ala Ser Pro Asp His His Met Met Ser Pro
               160
                                   165
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			att Ile 175													881
			tca Ser													929
			gtt Val													977
			aat Asn													1025
			gat Asp													1073
			act Thr 255													1121
tcc Ser	agg Arg	ata Ile 270	aat Asn	cca Pro	tac Tyr	agg Arg	atg Met 275	gtc Val	att Ile	gtg Val	ctg Leu	cga Arg 280	ttg Leu	att Ile	gtt Val	1169
Leu		Ile	ttc Phe				Arg					Val				1217
	Pro		tgg Trp								Ile					1265
			ttg Leu		Gln					Phe					Glu	1313
				Arg					Туг					glu	cca Pro	1361
			ı Ala					Phe					Asp		atg Met	1409
		Pro					: Ala					ı Sei			gct Ala	1457
gtg Va] 380	L Asp	tac Ty	c cct	gtg Val	gat L Asp 385	Lys	g gto S Val	c tct L Sei	t tgo	tat Tyi	c Vai	a tci l Sei	c gat	gat As <u>r</u>	gga Gly 395	1505
gct	gog	g ato	g cts	g aca	a ttt	ga!	gca	a cta	a gci	t gag	g ac	t tca	a gag	g tt1	gct	1553

Ala	Ala	Met	Leu	Thr 400	Phe	Asp	Ala	Leu	Ala 405	Glu	Thr	Ser	Glu	Phe 410	Ala	
aga Arg	aaa Lys	tgg Trp	gta Val 415	cca Pro	ttt Phe	gtt Val	aag Lys	aag Lys 420	tac Tyr	aac Asn	att Ile	gaa Glu	cct Pro 425	aga Arg	gct Ala	1601
cct Pro	gaa Glu	tgg Trp 430	tac Tyr	ttc Phe	tcc Ser	cag Gln	aaa Lys 435	att Ile	gat Asp	tac Tyr	ttg Leu	aag Lys 440	gac Asp	aaa Lys	gtg Val	16 <b>4</b> 9
cac His	cct Pro 445	tca Ser	ttt Phe	gtt Val	aaa Lys	gac Asp 450	cgc Arg	cgg Arg	gcc Ala	atg Met	aag Lys 455	aga Arg	gaa Glu	tat Tyr	gaa Glu	1697
gaa Glu 460	ttc Phe	aaa Lys	gtt Val	agg Arg	gta Val 465	aat Asn	ggc	ctt Leu	gtt Val	gct Ala 470	aag Lys	gca Ala	cag Gln	aaa Lys	gtt Val 475	1745
cct Pro	gag Glu	gaa Glu	gga Gly	tgg Trp 480	atc Ile	atg Met	caa Gln	gat Asp	ggc Gly 485	aca Thr	cca Pro	tgg Trp	cca Pro	gga Gly 490	aac Asn	1793
aat Asn	acc Thr	mgg Xaa	gac Asp 495	cat His	cct Pro	gga Gly	atg Met	att Ile 500	cag Gln	gtt Val	ttc Phe	ctt Leu	ggt Gly 505	cac His	agt Ser	1841
ggt Gly	ggc Gly	ctt Leu 510	gat Asp	act Thr	gag Glu	ggc Gly	aat Asn 515	gag Glu	cta Leu	ccc Pro	cgt Arg	ttg Leu 520	gtc Val	tat Tyr	gtt Val	1889
tct Ser	cgt Arg 525	gaa Glu	aag Lys	cgt Arg	cct Pro	gga Gly 530	ttc Phe	cag Gln	cat His	cac His	aag Lys 535	aaa Lys	gct Ala	ggt Gly	gcc Ala	1937
atg Met 540	aat Asn	gct Ala	ctt Leu	gtt Val	cgt Arg 545	gtc Val	tca Ser	gct Ala	gtg Val	ctt Leu 550	acc Thr	aat Asn	gga Gly	caa Gln	tac Tyr 555	1985
atg Met	ttg Leu	aat Asn	ctt Leu	gat Asp 560	tgt Cys	gat Asp	cac His	tac Tyr	att Ile 565	aac Asn	aac Asn	agt Ser	aag Lys	gct Ala 570	ctc Leu	2033
agg Arg	gaa Glu	gct Ala	atg Met 575	tgc Cys	ttc Phe	ctt Leu	atg Met	gac Asp 580	cct Pro	aac Asn	cta Leu	gga Gly	agg Arg 585	agt Ser	gtc Val	2081
tgc Cys	tac Tyr	gtc Val 590	cag Gln	ttt Phe	ccc Pro	cag Gln	aga Arg 595	ttc Phe	gat Asp	ggc Gly	att Ile	gac Asp 600	agg Arg	aat Asn	gat Asp	2129
cga Arg	tat Tyr 605	gcc Ala	aac Asn	agg Arg	aac Asn	acc Thr 610	gtg Val	ttt Phe	ttc Phe	gat Asp	att Ile 615	aac Asn	ttg Leu	aga Arg	ggt Gly	2177
ctt Leu 620	gat Asp	ggc Gly	atc Ile	caa Gln	gga Gly 625	cca Pro	gtt Val	tat Tyr	gtc Val	gga Gly 630	act Thr	ggc Gly	tgt Cys	gtt Val	ttc Phe 635	2225

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	c cga n Arg		_									_	_	_	-	2273
	ggt Gly		_				_				_	_	_	-		2321
	a aag r Lys															2369
	t gtg r Val 685							-							-	2417
	t gga a Gly O															2465
	g aag u Lys															2513
	g tat u Tyr															2561
-	a gct u Ala			_		_	_				_	_		_		2609
	a act y Thr 765											_	_			2657
	c gga r Gly 0															2705
	c aag o Lys															2753
	t ctg g Leu															2801
	c agc e Ser														_	2849
	c ctg e Leu 845	Glu	_										_		_	2897
to	c atc	ccg	ctt	ctc	atc	tac	tgc	atc	ctg	CCC	gcc	atc	tgt	ctg	ctc	2945

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Ser Ile Pro Leu Leu Ile Tyr Cys Ile Leu Pro Ala Ile Cys Leu Leu 860 865 870 875	
acc gga aag ttc atc att cca gag atc agc aac ttc gcc agc atc tgg Thr Gly Lys Phe Ile Ile Pro Glu Ile Ser Asn Phe Ala Ser Ile Trp 880 885 890	2993
Phe Ile Ser Leu Phe Ile Ser Ile Phe Ala Thr Gly Ile Leu Glu Met 895 900 905	3041
agg tgg agc ggg gtg ggc atc gac gag tgg tgg agg aac gag cag ttc Arg Trp Ser Gly Val Gly Ile Asp Glu Trp Trp Arg Asn Glu Gln Phe 910 915 920	3089
tgg gtg atc ggg ggc atc tcc gcg cac ctc ttc gcc gtg ttc cag ggc Trp Val Ile Gly Gly Ile Ser Ala His Leu Phe Ala Val Phe Gln Gly 925 930 935	3137
ctg ctc aag gtg ctg gcc ggc atc gac acc aac ttc acc gtc acc tcc Leu Leu Lys Val Leu Ala Gly Ile Asp Thr Asn Phe Thr Val Thr Ser 940 945 950 955	3185
aag gcc tcg gac gag gac ggc gac ttc gcg gag ctg tac atg ttc aag Lys Ala Ser Asp Glu Asp Gly Asp Phe Ala Glu Leu Tyr Met Phe Lys 960 965 970	3233
tgg acg acg ctc ctg atc ccg ccc acc acc atc ctg atc atc aac ctg Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Ile Leu Ile Ile Asn Leu 975 980 985	3281
gtc ggc gtc gtc gcc ggc atc tcc tac gcc atc aac agc gga tac cag Val Gly Val Val Ala Gly Ile Ser Tyr Ala Ile Asn Ser Gly Tyr Gln 990 995 1000	3329
tcg tgg ggc ccg ctc ttc ggc aag ctc ttc ttc gcc ttc tgg gtc atc Ser Trp Gly Pro Leu Phe Gly Lys Leu Phe Phe Ala Phe Trp Val Ile 1005 1010 1015	3377
gtc cac ctg tac ccg ttc ctc aag ggc ctc atg ggc agg cag aac cgc Val His Leu Tyr Pro Phe Leu Lys Gly Leu Met Gly Arg Gln Asn Arg 1020 1025 1030 1035	3425
acc deg acc atc gtc gtc gtc tgg gcc atc ctg ctg gcg tcc atc ttc Thr Pro Thr Ile Val Val Trp Ala Ile Leu Leu Ala Ser Ile Phe 1040 1045 1050	3473
tee ttg etg tgg gtt ege ate gae eee tte ace ace ege gte act gge Ser Leu Leu Trp Val Arg Ile Asp Pro Phe Thr Thr Arg Val Thr Gly 1055 1060 1065	3521
ccg gat acc cag acg tgt ggc atc aac t gctagggaag tggaaggttt Pro Asp Thr Gln Thr Cys Gly Ile Asn 1070 1075	3569
gtactttgta gaaacggagg aataccacgt gccatctgtt gtctgttaag ttatatatat ataagcagca agtggcgtta tttacagcta cgtacagacc agtggatatt gtttaccaca aagttttact tgtgttaata tgcattcttt tgttgatata aaaaaaaaa aaaaaaa	3629 3689 3746

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410
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Ser Gln Lys Ile Asp Tyr Leu Lys Asp Lys Val His Pro Ser Phe Val
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Lys Asp Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg
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Val Asn Gly Leu Val Ala Lys Ala Gln Lys Val Pro Glu Glu Gly Trp
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Glu Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg
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Pro Gly Phe Gln His His Lys Lys Ala Gly Ala Met Asn Ala Leu Val
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Pro Gln Ser Ala Thr Pro Glu Ser Leu Leu Lys Glu Ala Ile His Val
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His Ala Arg Gly Trp Arg Ser Ile Tyr Cys Met Pro Lys Arg Pro Ala
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                                 795
Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp Arg Leu Asn Gln Val
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             805
Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Leu Phe Ser Arg His Cys
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Pro Leu Trp Tyr Gly Tyr Gly Gly Arg Leu Lys Phe Leu Glu Arg Phe
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Ala Tyr Ile Asn Thr Thr Ile Tyr Pro Leu Thr Ser Ile Pro Leu Leu
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Ile Tyr Cys Ile Leu Pro Ala Ile Cys Leu Leu Thr Gly Lys Phe Ile
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875

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870

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                      890
Ile Ser Ile Phe Ala Thr Gly Ile Leu Glu Met Arg Trp Ser Gly Val
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                        920
Ile Ser Ala His Leu Phe Ala Val Phe Gln Gly Leu Leu Lys Val Leu
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Ala Gly Ile Asp Thr Asn Phe Thr Val Thr Ser Lys Ala Ser Asp Glu
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Asp Gly Asp Phe Ala Glu Leu Tyr Met Phe Lys Trp Thr Thr Leu Leu
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60

120

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gag	atg Met 1	gcg Ala	gcc Ala	aac Asn	aag Lys 5	Gly ggg	atg Met	gtg Val	gcg Ala	ggc Gly 10	tcg Ser	cac His	aac Asn	cgc Arg	aac Asn 15	228
												ggc Gly				276
ccc Pro	aca Thr	aag Lys	agt Ser 35	gcg Ala	aat Asn	gga Gly	cag Gln	gtc Val 40	tgc Cys	cag Gln	att Ile	tgc Cys	ggt Gly 45	gac Asp	tct Ser	324
gtg Val	ggt Gly	gtt Val 50	tca Ser	gcc Ala	act Thr	ggt Gly	gat Asp 55	gtc Val	ttt Phe	gtt Val	gcc Ala	tgc Cys 60	aat Asn	gag Glu	tgt Cys	372
gcc Ala	ttc Phe 65	cct Pro	gtc Val	tgc Cys	cgc Arg	cca Pro 70	tgc Cys	tat Tyr	gag Glu	tat Tyr	gag Glu 75	cgc Arg	aag Lys	gag Glu	gjå aaa	420
												aga Arg				468
												gtt Val				516
gac Asp	aat Asn	gaa Glu	ttc Phe 115	aac Asn	tac Tyr	aag Lys	caa Gln	ggc Gly 120	agt Ser	Gly aaa	aaa Lys	ggc Gly	cca Pro 125	gag Glu	tgg Trp	564
												gct Ala 140				612
												cag Gln				660
												cgc Arg				708
												agg Arg				756
												gtt Val				804
												aat Asn 220				852
gtg	act	aat	aaa	tat	cca	gag	gct	aga	gga	gga	gac	atg	gag	<b>a</b> aa	act	900

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Val	Thr 225	Asn	Lys	Tyr	Pro	Glu 230	Ala	Arg	Gly	Gly	Asp 235	Met	Glu	Gly	Thr	
					nat Xaa 245											948
_	-	_			cca Pro					_						996
_					cgt Arg				_	_				_		1044
					gtg Val	_	-	_						_		1092
-		_		_	tgg Trp		_	_					_	_		1140
					atc Ile 325		_					_			_	1188
	-				gag Glu					_	_	-			-	1236
					gtg Val			-	-	-			_			1284
					tcc Ser											1332
		_		-	tct Ser	_	_			_		_				1380
	Leu				gca Ala 405											1428
					gaa Glu		_	_		-				_		1476
		-		_	aag Lys	-							-	_	-	1524
			_		agg Arg			_	_							1572

gcc Ala	ctt Leu 465	gtt Val	gcc Ala	aaa Lys	gca Ala	cag Gln 470	aaa Lys	gtg Val	cct Pro	gaa Glu	gag Glu 475	gly aaa	tgg Trp	acc Thr	atg Met	1620
gct Ala 480	gat Asp	gga Gly	act Thr	gca Ala	tgg Trp 485	cct Pro	gly aaa	aat Asn	aat Asn	cct Pro 490	agg Arg	gac Asp	cat His	cct Pro	ggc Gly 495	1668
atg Met	att Ile	cag Gln	gtt Val	ttc Phe 500	ttg Leu	GJA āāā	cac His	agt Ser	ggt Gly 505	gly ggg	ctc Leu	gac Asp	act Thr	gat Asp 510	gga Gly	1716
aat Asn	gag Glu	tta Leu	cca Pro 515	cgt Arg	ctt Leu	gtc Val	tat Tyr	gtc Val 520	tct Ser	cgt Arg	gaa Glu	aag Lys	aga Arg 525	cca Pro	ggc Gly	1764
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tct Ser	gct Ala 545	Val	ctg Leu	aca Thr	aat Asn	ggt Gly 550	gcc Ala	tat Tyr	ctt Leu	ctc Leu	aat Asn 555	gtg Val	gat Asp	tgc Cys	gac Asp	1860
cat His 560	Tyr	ttc Phe	aat Asn	ago Ser	agc Ser 565	Lys	gct Ala	ctt Leu	aga Arg	gaa Glu 570	Ala	atg Met	tgc Cys	ttc Phe	atg Met 575	1908
atg Met	gat Asp	ccg Pro	gct Ala	cta Lev 580	ı Gly	agg Arg	aaa Lys	act Thr	tgt Cys 585	Tyr	gta Val	caa Gln	ttt Phe	cca Pro 590	Gln	1956
aga Arg	ttt Phe	gat Asp	ggc Gly 595	r Ile	gac S Asp	ttg Leu	cac His	gat Asp 600	Arg	tat Tyr	gct Ala	aat Asn	cgg Arg 605	Asn	ata	2004
gtt Val	tto L Phe	tti Pho	e Asp	ato Ile	c aad e Asr	ato n Met	aaa Lys 61!	Gly	ctg Lev	gat 1 Asp	ggc Gly	att / Ile 620	Gln	ggt Gly	cca Pro	2052
gti Va:	tac l Tyr 62!	r Va	g gga l Gl	a ac	a gga r Gly	a tgo y Cys 630	s Cy	t tto	c aat e Asr	aga n Arg	g Glr 635	n Ala	ttg Lev	tat Tyr	gga Gly	2100
ta Ty:	r As	t cc p Pr	t gt o Va	t tt	g actuary the second se	r Gl	agc Ala	t gat a Asj	t cto p Le	g gaq u Gli 650	ı Pro	a aac o Asr	att n Ile	gtt Val	att L Ile 655	2148
aa Ly	g ag s Se	c tg r Cy	c tg s Cy	t gg s Gl 66	y Ar	a ag	g aa g Ly	g aa s Ly	a aag s Ly 66	s As	c aag n Ly	g agt s Sei	tat r Tyi	ate Me	g gat t Asp	2196
ag Se	t ca r Gl	a ag n Se	c cg r Ar 67	g Il	t at e Me	g aa t Ly	g ag s Ar	a ac g Th 68	r Gl	a tc u Se	t tc r Se	a gci r Ala	t ccc a Pro 689	o II	c ttc e Phe	2244
aa	t at	g ga	ia ga	c at	c ga	a ga	g gg	t at	t ga	a gg	t ta	c ga	g ga	t ga	a agg	2292

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Asn	Met	Glu 690	Asp	Ile	Glu	Glu	Gly 695	Ile	Glu	Gly	Tyr	Glu 700	Asp	Glu	Arg	
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												ggc Gly				2388
tca Ser	aca Thr	aac Asn	cca Pro	gct Ala 740	tct Ser	cta Leu	cta Leu	aag Lys	gaa Glu 745	gct Ala	atc Ile	cat His	gtc Val	atc Ile 750	agt Ser	2436
tgt Cys	gga Gly	tat Tyr	gag Glu 755	gac Asp	aaa Lys	act Thr	gaa Glu	tgg Trp 760	gga Gly	aaa Lys	gag Glu	att Ile	ggc Gly 765	tgg Trp	atc Ile	2484
tat Tyr	ggt Gly	tca Ser 770	gta Val	acg Thr	gag Glu	gat Asp	att Ile 775	ctg Leu	act Thr	Gly aaa	ttt Phe	aaa Lys 780	atg Met	cat His	gca Ala	2532
		Trp										cct Pro				2580
	Ser											cag Gln				2628
tgg Trp	gct Ala	ctt Leu	gly	tca Ser 820	Val	gaa Glu	att Ile	ctg Leu	ctt Leu 825	agt Ser	aga Arg	cat His	tgt Cys	cct Pro 830	atc Ile	2676
				Asn					Leu			agg Arg				2724
ato Ile	aac Asn	act Thr	Ile	gta Val	tat Tyr	cca Pro	atc Ile 855	Thr	tcc Ser	att	ccg Pro	ctt Leu 860	Ile	gcc Ala	tat Tyr	2772
		. Leı					Leu					ttt Phe				2820
gag Glu 880	ılle	ago Ser	aat Asn	tat Tyr	gct Ala 885	Gly	atg Met	tto Phe	tto Phe	att 11e 890	e Lei	t ctt 1 Leu	ttc Phe	gcc Ala	tcc Ser 895	2868
					/ Ile					, Tr		t ggt r Gly			, Ile	2916
gaa Glu	a gat ı Ası	tgg p Tr	g tgg P Trp 915	Arg	a aat g Asr	gag Glu	g cag ı Glr	tti Phe 920	e Tr	g gtt p Val	ati	t ggt e Gly	ggo Gl <sub>y</sub> 925	Thi	tct Ser	2964

- 68 -

	cat His															3012
	gat Asp 945					_			_	_		_		_		3060
_	ttt Phe	_						_			_	_				3108
_	acc Thr		_		_			-	_		_		_			3156
	tat Tyr	_		Asn	-				Ser			_		Phe		3204
_	r ctg : Leu		Phe	_				Ile					Pro			3252
	ggt Gly 102	Leu					Asn					Ile				3300
	tcc Ser				_	Ser				_	Leu			_		3348
_	cct Pro				Pro		_		_	Ala	_	_			Cys	3396
	gtc YVal		_	ctga	tcga	g ac	agtg	actc	tta	tttg	aag	aggc	tcaa	tc		3446
gtg tgc aag	gagga etgeg gatgt	tgg gac gaa gtg	attt taag tttt	gcat aatc gaag	ct a ac g tt t	agtt gagc tgtt	atgc cttt atgc	c tc c ta g tg	tgtt cctt cagt	catt ccat ttat	agc gta tgt	ttct gcgc ttta	tcc cag gag	gtgc ccag taaa	ttgtag cggtgc cagcgt ttatca aaaaaa	3506 3566 3626 3686 3746 3753
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 Asn
 Arg
 Asn
 Glu

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 10
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 15

 Phe
 Val
 Met
 Ile
 Arg
 His
 Asp
 Gly
 Asp
 Val
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 Gly
 Ser
 Ala
 Lys
 Pro

 20
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 30
 30
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Thr Lys Ser Ala Asn Gly Gln Val Cys Gln Ile Cys Gly Asp Ser Val Gly Val Ser Ala Thr Gly Asp Val Phe Val Ala Cys Asn Glu Cys Ala 55 60 Phe Pro Val Cys Arg Pro Cys Tyr Glu Tyr Glu Arg Lys Glu Gly Asn 70 Gln Cys Cys Pro Gln Cys Lys Thr Arg Tyr Lys Arg Gln Lys Gly Ser 90 Pro Arg Val His Gly Asp Glu Asp Glu Glu Asp Val Asp Asp Leu Asp 105 Asn Glu Phe Asn Tyr Lys Gln Gly Ser Gly Lys Gly Pro Glu Trp Gln 120 Leu Gln Gly Asp Asp Ala Asp Leu Ser Ser Ser Ala Arg His Glu Pro 135 140 His His Arg Ile Pro Arg Leu Thr Ser Gly Gln Gln Ile Ser Gly Glu 155 150 Ile Pro Asp Ala Ser Pro Asp Arg His Ser Ile Arg Ser Pro Thr Ser 165 170 Ser Tyr Val Asp Pro Ser Val Pro Val Pro Val Arg Ile Val Asp Pro 180 185 Ser Lys Asp Leu Asn Ser Tyr Gly Leu Asn Ser Val Asp Trp Lys Glu 1.95 200 205 Arg Val Glu Ser Trp Arg Val Lys Gln Asp Lys Asn Met Met Gln Val 210 215 Thr Asn Lys Tyr Pro Glu Ala Arg Gly Gly Asp Met Glu Gly Thr Gly 225 230 235 Ser Asn Gly Glu Xaa Met Gln Met Val Asp Asp Ala Arg Leu Pro Leu 245 250 Ser Arg Ile Val Pro Ile Ser Ser Asn Gln Leu Asn Leu Tyr Arg Val 265 Val Ile Ile Leu Arg Leu Ile Ile Leu Cys Phe Phe Phe Gln Tyr Arg 280 Val Ser His Pro Val Arg Asp Ala Tyr Gly Leu Trp Leu Val Ser Val 295 300 Ile Cys Glu Val Trp Phe Ala Leu Ser Trp Leu Leu Asp Gln Phe Pro 310 315 320 Lys Trp Tyr Pro Ile Asn Arg Glu Thr Tyr Leu Asp Arg Leu Ala Leu 330 Arg Tyr Asp Arg Glu Gly Glu Pro Ser Gln Leu Ala Pro Ile Asp Val 345 Phe Val Ser Thr Val Asp Pro Leu Lys Glu Pro Pro Leu Ile Thr Ala · 355 360 Asn Thr Val Leu Ser Ile Leu Ser Val Asp Tyr Pro Val Asp Lys Val 375 380 Ser Cys Tyr Val Ser Asp Asp Gly Ser Ala Met Leu Thr Phe Glu Ser 390 395 Leu Ser Glu Thr Ala Glu Phe Ala Arg Lys Trp Val Pro Phe Cys Lys 405 410 Lys His Asn Ile Glu Pro Arg Ala Pro Glu Phe Tyr Phe Ala Gln Lys 425 Ile Asp Tyr Leu Lys Asp Lys Ile Gln Pro Ser Phe Val Lys Glu Arg 440 445 Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Ile Asn Ala 455 460 Leu Val Ala Lys Ala Gln Lys Val Pro Glu Glu Gly Trp Thr Met Ala 470 475 Asp Gly Thr Ala Trp Pro Gly Asn Asn Pro Arg Asp His Pro Gly Met 485 490

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		835			Arg		840					845			
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                      1015
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geggaagtag aggggaggaa geg atg gag geg age gee ggg etg gte gee gge
                                                                 173
                         Met Glu Ala Ser Ala Gly Leu Val Ala Gly
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                                                                 221
Ser His Asn Arg Asn Glu Leu Val Val Ile Arg Arg Asp Gly Asp Pro
ggg ccg aag ccg ccg cgg gag cag aac ggg cag gtg tgc cag att tgc
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Gly Pro Lys Pro Pro Arg Glu Gln Asn Gly Gln Val Cys Gln Ile Cys
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tct Ser	caa Gln	ctt Leu	gct Ala 350	cca Pro	att Ile	gat Asp	ttc Phe	ttt Phe 355	gtc Val	agt Ser	acg Thr	gtt Val	gat Asp 360	ccc Pro	tta Leu	1229
										gtt Val						1277
gtg Val	gat Asp 380	tat Tyr	cct Pro	gtt Val	gat Asp	aag Lys 385	gtt Val	tct Ser	tgc Cys	tat Tyr	gtt Val 390	tct Ser	gat Asp	gat Asp	ggt Gly	1325
gct Ala 395	gca Ala	atg Met	cta Leu	acg Thr	ttt Phe 400	gaa Glu	gca Ala	tta Leu	tct Ser	gaa Glu 405	aca Thr	tct Ser	gaa Glu	ttt Phe	gca Ala 410	1373
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gaa Glu	ttc Phe 460	aag Lys	gtg Val	aga Arg	atc Ile	aat Asn 465	gcc Ala	tta Leu	gtt Val	gcc Ala	aaa Lys 470	gcc Ala	cag Gln	aaa Lys	gtt Val	1565
cct Pro 475	gaa Glu	gaa Glu	gga Gly	tgg Trp	aca Thr 480	atg Met	caa Gln	gat Asp	gga Gly	acc Thr 485	ccc Pro	tgg Trp	cct Pro	gga Gly	aac Asn 490	1613
										gtc Val						1661

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	aga Arg															1757
	aat Asn 540															1805
_	tta Leu		_	_	_	-						-	_	_		1853
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	gat Asp 620															2045
_	agg Arg	_	_					_	_				_			2093
	tca Ser				Asn											2141
	ttt Phe								Thr					Thr	gag Glu	2189
	aaa Lys		Leu					Lys					Ser		gca Ala	2237
		Leu					Glu					Ala			gaa Glu	2285
	Ala					Gln					Lys				caa Gln 730	2333
tct	tct	gtt	ttt	gtt	aca	. tcc	aca	ctt	cto	gag	aat	ggt	gga	acc	ttg	2381

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Ser	Ser	Val	Phe	Val 735	Thr	Ser	Thr	Leu	Leu 740	Glu	Asn	Gly	Gly	Thr 745	Leu	
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agt Ser	tgt Cys	ggt Gly 765	tat Tyr	gaa Glu	gac Asp	aag Lys	aca Thr 770	gac Asp	tgg Trp	gga Gly	aaa Lys	gag Glu 775	att Ile	ggc	tgg Trp	2477
atc Ile	tat Tyr 780	gga Gly	tca Ser	gtt Val	aca Thr	gaa Glu 785	gat Asp	att Ile	cta Leu	act Thr	ggt Gly 790	ttc Phe	aag Lys	atg Met	cat His	2525
tgt Cys 795	cat His	ggt Gly	tgg Trp	cgg Arg	tca Ser 800	att Ile	tac Tyr	tgc Cys	ata Ile	cct Pro 805	aaa Lys	cgg Arg	gtt Val	gca Ala	ttc Phe 810	2573
aaa Lys	ggt Gly	tct Ser	gca Ala	cct Pro 815	ctg Leu	aat Asn	ctt Leu	tca Ser	gat Asp 820	cgt Arg	ctt Leu	cac His	cag Gln	gtg Val 825	ctt Leu	2621
cgg Arg	tgg Trp	gct Ala	ctt Leu 830	GJÀ āāā	tct Ser	att Ile	gag Glu	atc Ile 835	ttc Phe	ttc Phe	agc Ser	aat Asn	cat His 840	tgc Cys	cct Pro	2669
ctt Leu	tgg Trp	tat Tyr 845	gly aaa	tat Tyr	ggt Gly	ggc Gly	ggt Gly 850	ctg Leu	aaa Lys	ttt Phe	ttg Leu	gaa Glu 855	aga Arg	ttt Phe	tcc Ser	2717
tac Tyr	atc Ile 860	aac Asn	tcc Ser	atc Ile	gtg Val	tat Tyr 865	cct Pro	tgg Trp	aca Thr	tct Ser	att Ile 870	ccc Pro	ctc Leu	ttg Leu	gct Ala	2765
tac Tyr 875	tgt Cys	aca Thr	ttg Leu	cct Pro	gcc Ala 880	atc Ile	tgt Cys	tta Leu	ttg Leu	aca Thr 885	gjå aaa	aaa Lys	ttt Phe	atc Ile	act Thr 890	2813
cca Pro	gag Glu	ctg Leu	aat Asn	aat Asn 895	gtt Val	gcc Ala	agc Ser	ctg Leu	tgg Trp 900	ttc Phe	atg Met	tca Ser	ctt Leu	ttt Phe 905	atc Ile	2861
tgc Cys	att Ile	ttt Phe	gct Ala 910	acg Thr	agc Ser	atc Ile	cta Leu	gaa Glu 915	atg Met	aga Arg	tgg Trp	agt Ser	ggt Gly 920	gtt Val	gga Gly	2909
att Ile	gat Asp	gac Asp 925	tgg Trp	tgg Trp	agg Arg	aat Asn	gag Glu 930	cag Gln	ttc Phe	tgg Trp	gtc Val	att Ile 935	gga Gly	ggt Gly	gtg Val	2957
tcc Ser	tca Ser 940	cac His	ctc Leu	ttt Phe	gct Ala	gtg Val 945	ttc Phe	cag Gln	gga Gly	ctt Leu	ctc Leu 950	aag Lys	gtc Val	ata Ile	gct Ala	3005
ggt Gly 955	gtt Val	gat Asp	aca Thr	agc Ser	ttc Phe 960	acc Thr	gtg Val	aca Thr	tca Ser	aag Lys 965	ggt Gly	gga Gly	gat Asp	gat Asp	gag Glu 970	3053

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cct acc acc ttg ctt cta ttg aac ttc att ggt gtg gtc gct ggc gtt Pro Thr Thr Leu Leu Leu Asn Phe Ile Gly Val Val Ala Gly Val 990 995 1000	3149
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ttg gat tgc a actaggatgt cagtgcatca gctccccaa tctgcatatg Leu Asp Cys 1085	3439
Leu Asp Cys 1085  cttgaagtat attttctggt gtttgtcccc atattcagtg tctgtagata agagacatga aatgtcccaa gtttcttttg atccatggtg aacctactta atatctgaga gatatactgg gggaaaatgg aggctgcggc aatccttgtg cagttgggcc gtggaataca gcatatgcaa gtgtttgatt gtgcagcatt ctttattact tggtcgcaat atagatgggc tgagccgaac agcaaggtat tttgattctg cactgctccc gtgtacaaac ttggttctca ataaggcagg caggaatgca tctgccagtg gaacagagca acctgcacat tatttatgta tgcctgttca ttggagggct tgttcattac atgttcgtct atactagaaa aaacagaata ttagcattaa tctatagtta attaaagtat gtaaatgcgc ctgttttttg ttgtgtactg taatcatctg agttggtttt gtgaaaaaaa aaaaaaaaa aaaaaaaa	3499 3559 3619 3679 3739 3859 3919 3969
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Leu Asp Cys 1085  Cttgaagtat attttctggt gtttgtcccc atattcagtg tctgtagata agagacatga aatgtcccaa gtttctttg atccatggtg aacctactta atatctgaga gatatactgg gggaaaatgg aggctgcggc aatccttgtg cagttgggcc gtggaataca gcatatgcaa gtgttttgatt gtgcagcatt ctttattact tggtcgcaat atagatgggc tgagccgaac agcaaggtat tttgattctg cactgctccc gtgtacaaac ttggttcca ataaggcagg caggaatgca tctgccagtg gaacagagca acctgcacat tatttatgta tgcctgttca ttggagggct tgttcattac atgttcgtct atactagaaa aaacagaata ttagcattaa tctatagtta attaaagtat gtaaatgcgc ctgttttttg ttgtgtactg taatcatctg agttggtttt gtgaaaaaaa aaaaaaaaaa	3499 3559 3619 3679 3739 3799 3859 3919
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Gln 145	Ala	Phe	Gln	Leu	Asn 150	Pro	Asn	Val	Pro	Leu 155	Leu	Thr	Asn	Gly	Gln 160
Met	Val	Asp	Asp	Ile 165	Pro	Pro	Glu	Gln	His 170		Leu	Val	Pro	Ser 175	
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Gln	Leu	Ser	Arg 260	Lys	Ile	Pro	Leu	Pro 265	Ser	Ser	Gln	Ile	Asn 270	Pro	Tyr
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		355					Asp 360					365			
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			420				Pro	425					430		
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			500				Gly	505					510		
		515					Val 520					525			
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	٠			565					570			Ala		575	
			580					585				Val	590		
Gln	Arg	Phe 595	Asp	Gly	Ile	Asp	Arg 600	His	Asp	Arg	Tyr	Ala 605	Asn	Arg	Asn
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			aac Asn													562
			gat Asp													610
			att Ile													658
			gca Ala													706
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		Glu	agc Ser				Lys					Met				850
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					Gln					Ala					agc Ser	946
_				Ile					Leu					Ile	gta Val	994
			ı Arg					ı Cys					туг		atc Ile	1042
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WO 00/09706

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	_	_				gct Ala 375		-			_	_				1330
-		_		_	_	ggc Gly		_	_	_			-			1378
						gct Ala										1426
						gct Ala										1474
_		_	_	_		att Ile					_	_	-	-	_	1522
_	_	_	_			gaa Glu 455	_				_	Ile				1570
						gtg Val										1618
		_				aat Asn				Asp				-	Ile	1666
_			_	Gly		-			Leu	_		_		Asn	gaa Glu	1714
			Leu					Arg					Gly		cag Gln	1762

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cat His	cac His	3 rAs	, aag Lys	gct Ala	ggt Gly	gca Ala 535	Met	, aat : Asr	gca Ala	cto Leu	g att 1 Ile 540	e Arg	gta JVal	tct Ser	gct Ala	1810
gtg Val 545	. Let	aca Thr	aat Asn	ggt Gly	gcc Ala 550	Tyr	ctt Leu	cto Leu	aat Asn	gtg Val	Asp	tgt Cys	gac Asp	cat His	tac Tyr 560	1858
ttc Phe	aat Asn	ago Ser	agc Ser	aaa Lys 565	Ala	ctt Leu	aga Arg	gaa Glu	gca Ala 570	Met	tgc Cys	ttc Phe	atg Met	atg Met	gat	1906
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Asp	GIY	595	Asp	Leu	His	Asp	Arg 600	Tyr	gct Ala	Asn	Arg	Asn 605	Ile	Val	Phe	2002
Pne	610	TTE	Asn	Met	Lys	Gly 615	Leu	Asp	ggc	Ile	Gln 620	Gly	Pro	Val	Tyr	2050
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GIU	690	116	GIU	GIU	GIY	11e 695	GLu	Gly	tat Tyr	Glu	Asp 700	Glu	Arg	Ser	Val	2290
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	cca Pro									_	_		-		-	2626
	GJÀ 333															2674
	tac Tyr															2722
	att Ile 850	_						_	_			_		_		2770
	cct Pro	-		_												2818
_	aat Asn		_		_							_				2866
_	act Thr			_			-		_		-			-	-	2914
	tgg Trp	_			_			_						-		2962
	ttc Phe 930				_		_	_			_	_			_	3010
	aac Asn			_			_	_		_		_		_		3058
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	gtt Val								Met							3154

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Ala	Ile	Asn 999		Gly	Tyr	Gln	Ser 1000		Gly	Pro	Leu	Phe 1005		Lys	Leu	
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	Met					Arg					Val			tgg Trp		3298
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ccc	ctcgt		aata	accto											gatgga	3508
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atca catt	atgga cttt! caca	agc d ta t ega a	agtta ettte eacgt actat	ctaco cggto	et to yt ti	gcttg cattg	gtagt gttt	gct aga	ggco agtaa	cagc aatt		,taaa	att 9			3628 3688
atca catt	atgga ettti eacad	age o ta t ega a 210>	agtta ettte eacgt actat	etaco eggto eatgo	et to yt ti	gcttg cattg	gtagt gttt	gct aga	ggco agtaa	cagc aatt		,taaa	att 9			3628 3688
atca catt	atgga ettti eacad <2 <2	age of the temperature of temperature of the temperature of temperature of temperature of temperature of temperature of temperature of temperature	agtta cacgt actat 42 1074	etaco eggto eatgo	et to yt ti	gcttg cattg	gtagt gttt	gct aga	ggco agtaa	cagc aatt		,taaa	att 9			3628 3688
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atca catt	atgga cttti cacac <2 <2 <2	age of the temperature of temperature of the temperature of temperature of temperature of temperature of temperature of temperature of temperature	agtta ctttc actal 42 1074 PRT Zea	etaco eggto eatgo	et to gt tt gc aa	gcttg cattg	gtagt gttt	gct aga	ggco agtaa	cagc aatt		,taaa	att 9			3628 3688
atca catt atto	atgga cttti cacac <2 <2 <2	age of ta	agtta cttto cacgt actat 42 1074 PRT Zea	etace eggte eatge 1 mays	et togge et togge	gette catte atget	ytagt yttt :gtta	t get t aga t t t	egge eggtas eaaas	cage aatt a	atca	gtaaa	att g	tgagg	gtaact	3628 3688
atca catt atto	atgga ettti cacac <2 <2 <2 <2 <4 Ala	age of the temperature of temperature of the temperature of temperature of temperature of temperature of temperature of temperature of temperature	agtta cattto actal 42 1074 PRT Zea 42 Asn	etace eggte catge f mays Lys 5	ct to	gettg attget	yttagt ytttt cgtta	age ttt	Egged agtaa caaaa Gly 10	cagc aatt a	atca His	ytaaa uttto Asn	Arg	Asn 15	gtaact	3628 3688
atca catt atto	atgga ettti cacac <2 <2 <2 <2 <4 Ala	age of the temperature of temperature of the temperature of temperature of temperature of temperature of tem	agtta cattto actal 42 1074 PRT Zea 42 Asn	etace eggte catge f mays Lys 5	ct to	gettg attget	yttagt ytttt gtta	age ttt	Egged agtaa caaaa Gly 10	cagc aatt a	atca His	ytaaa uttto Asn	Arg	Asn	gtaact	3628 3688
Met 1 Phe	atgga cttti cacac <2 <2 <2 Ala Val	agc of the term of	agtta tacgt actat 42 1074 PRT Zea 42 Asn Ile 20	tace eggte atge mays Lys 5 Arg	Gly His	gettg attget Met Asp	ytagt yttt gtta Val Gly Val	Ala Asp	Gly 10 Ala	ser Pro	His Val	Asn Pro	Arg Ala 30	Asn 15	Glu Pro	3628 3688
Met 1 Phe	atgga cttti cacac	agc of ta the control of the control	agtta catto cacgt actat 42 1074 PRT Zea 42 Asn Ile 20 Ala	tace eggte atge mays Lys 5 Arg	Gly His	Met Asp Gln Asp	Val Gly Val 40	Ala Asp 25 Cys	Gly 10 Ala	Ser Pro	His Val Cys	Asn Pro Gly 45	Arg Ala 30 Asp	Asn 15 Lys	Glu Pro Val	3628 3688
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Met 1 Phe Thr Gly Phe 65 Gln Pro Asn	Atggattttteacac	agc of the term of	agtta ctttc cacgt actal 42 1074 PRT Zea 42 Asn Ile 20 Ala Ala Cys Pro His 100 Asn	Lys Sharp Arg Arg Gln 85 Gly Tyr	Gly His Gly Pro 70 Cys Asp	Met Asp Gln Asp Cys Lys Asp Gln	Val Gly Val Tyr Thr Glu Gly 120	Ala Asp 25 Cys Phe Glu Arg Glu 105 Asn	Gly 10 Ala Gln Val Tyr 90 Glu Gly	Ser Pro Ile Ala Glu 75 Lys Asp	His Val Cys 60 Arg Val	Asn Pro Gly 45 Asn Lys Gln Asp	Arg Ala 30 Asp Glu Lys Asp 110 Glu	Asn 15 Lys Thr Cys Gly 95 Leu	Glu Pro Val Ala Asn 80 Ser Asp Gln	3628 3688
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95

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gat Asp	tgg Trp	gat Asp 250	ggc Gly	gac Asp	gat Asp	gca Ala	gat Asp 255	ctg Leu	cca Pro	cta Leu	atg Met	gat Asp 260	gaa Glu	gct Ala	agg Arg	1003
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ata Ile	tct Ser	gta Val	atc Ile 315	tgt Cys	gaa Glu	atc Ile	Trp	ttt Phe 320	gcg Ala	atg Met	tcc Ser	tgg Trp	att Ile 325	ctt Leu	gat Asp	1195
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Gln	Phe	Pro 330	Lys	Trp	Leu	Pro	Ile 335	Glu	Arg	Glu	Thr	Tyr 340	Leu	Asp	Arg	
_					-	-	_				tct Ser 355	_		-		1291
	-			_	_	_	_	_			aag Lys	_				1339
-					_						gtg Val	-			-	1387
	_	_		_		-		_	-		gct Ala	_	-		_	1435
	_	_	_		_			_		_	aag Lys			_		1483
	_		_						-	_	cct Pro 435					1531
	_	_		_		_		_			gct Ala					1579
					_	_	_	-			gaa Glu					1627
_			_	_					_	-	cct Pro					1675
	_		_		_						aac Asn	_	_	_		1723
		_		_	_					_	ggc Gly 515		_	_		1771
_	Gly			-		Arg	_	_		_		_	_	_	agg Arg 535	1819
						_	_	_		Ala	atg Met		_	_	Val	1867
_	_		_	_				-	Ala			_		Leu	gac Asp	1915

WO 00/09706

tg Cy	t gai	cac His 570	Tyr	atc Ile	aac Asn	aat Asn	agc Ser 575	aag Lys	gcc Ala	ata Ile	aaa Lys	gag Glu 580	gct Ala	atg Met	tgt Cys	1963
t t Ph	c ato e Mei 58!	g atg : Met	gat Asp	cct Pro	ttg Leu	gtg Val 590	Gly aaa	aag Lys	aaa Lys	gtg Val	tgc Cys 595	tat Tyr	gta Val	cag Gln	ttc Phe	2011
ec Pr 60	o Gli	g agg n Arg	ttt Phe	gat Asp	ggt Gly 605	att Ile	gac Asp	aaa Lys	aat Asn	gat Asp 610	cga Arg	tac Tyr	gct Ala	aac Asn	agg Arg 615	2059
aa As	c gtt n Val	gtc Val	ttt Phe	ttt Phe 620	gac Asp	atc Ile	aac Asn	atg Met	aaa Lys 625	ggt Gly	ttg Leu	gac Asp	ggt Gly	att Ile 630	caa Gln	2107
G1 39	a cco y Pro	att Ile	tat Tyr 635	gtg Val	ggt Gly	act Thr	gga Gly	tgt Cys 640	gtt Val	ttc Phe	aga Arg	cgg Arg	cag Gln 645	gca Ala	ctg Leu	2155
ta Ty	t ggt r Gly	tat Tyr 650	gat Asp	gct Ala	cct Pro	aaa Lys	acg Thr 655	Lys	aag Lys	cca Pro	cca Pro	tca Ser 660	aga Arg	act Thr	tgc Cys	2203
aa As	n Cys	tgg Trp	ccc Pro	aag Lys	tgg Trp	tgc Cys 670	ctc Leu	tct Ser	tgc Cys	tgc Cys	tgc Cys 675	agc Ser	agg Arg	aac Asn	aag Lys	2251
aa As 68	n Lys	aag Lys	aag Lys	act Thr	aca Thr 685	aaa Lys	cca Pro	aag Lys	acg Thr	gag Glu 690	aag Lys	aag Lys	aaa Lys	aga Arg	tta Leu 695	2299
tt Ph	t tto e Phe	aag Lys	aaa Lys	gca Ala 700	gaa Glu	aac Asn	cca Pro	tct Ser	cct Pro 705	gca Ala	tat Tyr	gct Ala	ttg Leu	ggt Gly 710	gaa Glu	2347
at Il	t gat e Asp	gaa Glu	ggt Gly 715	gct Ala	cca Pro	ggt Gly	gct Ala	gat Asp 720	atc Ile	gag Glu	aag Lys	gcc Ala	gga Gly 725	atc Ile	gta Val	2395
aa As:	t caa n Gln	cag Gln 730	aaa Lys	cta Leu	gag Glu	aag Lys	aaa Lys 735	ttt Phe	gjå aaa	cag Gln	tct Ser	tct Ser 740	gtt Val	ttt Phe	gtc Val	2443
gc: Al:	a tca a Ser 745	aca Thr	ctt Leu	ctt Leu	gag Glu	aac Asn 750	gga Gly	gly	acc Thr	ctg Leu	aag Lys 755	agc Ser	gca Ala	agt Ser	cca Pro	2491
gci Ala 76	a Ser	ctt Leu	ctg Leu	aag Lys	gaa Glu 765	gct Ala	ata Ile	cat His	gtt Val	atc Ile 770	agc Ser	tgc Cys	ggc Gly	tac Tyr	gaa Glu 775	2539
ga Ası	c aag p Lys	acc Thr	gac Asp	tgg Trp 780	gga Gly	aaa Lys	gag Glu	att Ile	ggc Gly 785	tgg Trp	att Ile	tac Tyr	gga Gly	tcg Ser 790	atc Ile	2587
aca	a gag	gat	atc	ttg	act	gga	ttt	aag	atg	cac	tgc	cat	ggc	tgg	cgg	2635

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Thr	Glu	Asp	Ile 795	Leu	Thr	Gly	Phe	Lys 800	Met	His	Cys	His	Gly 805	Trp	Arg	
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ctg Leu	aac Asn 825	ctt Leu	tcc Ser	gac Asp	cgt Arg	ctt Leu 830	cac His	cag Gln	gtc Val	ctt Leu	cgc Arg 835	tgg Trp	gcc Ala	ctt Leu	gl <sup>A</sup> aaa	2731
tcc Ser 840	gtc Val	gaa Glu	att Ile	ttc Phe	ttc Phe 845	agc Ser	aag Lys	cac His	tgc Cys	cca Pro 850	ctt Leu	tgg Trp	tac Tyr	gga Gly	tac Tyr 855	2779
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gtt Val	tat Tyr	ccc Pro	tgg Trp 875	acg Thr	tcc Ser	att Ile	cct Pro	ctc Leu 880	ctg Leu	gct Ala	tac Tyr	tgt Cys	acc Thr 885	ttg Leu	cct Pro	2875
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tac Tyr	acg Thr 985	Phe	aag Lys	tgg Trp	acc Thr	acc Thr 990	Leu	ctg Leu	ata Ile	ccc Pro	ccg Pro 999	acc Thr	acg Thr	ctc Leu	ctc Leu	3211
ctg Leu 1000	Leu	aac Asn	ttc Phe	atc Ile	999 1005	Val	gtg Val	gcc Ala	gly aaa	atc Ile 1010	Ser	aac Asn	gcg Ala	atc Ile	aac Asn 1015	3259
aac Asn	gly aaa	tac Tyr	gag Glu	tcg Ser 1020	$\mathtt{Trp}$	ggc Gly	ccc Pro	ctg Leu	ttc Phe 1025	Gly	aag Lys	ctc Leu	ttc Phe	ttc Phe 1030	Ala	3307

Phe	tgg	gtg Val	103	e Val	cac His	ctg Leu	tac Tyr	Pro 104	) Phe	cto Lev	aag Lys	ggt Gl	Let Let 104	ı Val	r grå a aaa	3355
agg Arg	Gln	aac Asn 105	ı Arç	g acg g Thr	ccg	acg Thr	ato Ile 105	· Val	ato l Ile	gtc Val	tgg Trp	s tec Ser 106	: Ile	c cto	g ctg 1 Leu	3403
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aag Lys 108	Ser	aac Asn	ggc	ccg Pro	Leu 108	Leu	gag Glu	gag Glu	tgt Cys	ggc Gly 109	Leu	gac Asp	tgo Cys	a s		3494
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/al	Pro	Ser	Tyr 180	Met	Ser	Gly	Gly		170 Gly	Gly	Gly	Lys		175 Ile	His	
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Lys 225	Glu	Arg	Met	Glu	Gly	Trp	Lys	Gln	Lys		Glu	Arg	Leu	Gln	His
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Pro	Len	Met	Δen	245	Ala	7 ra	Cl n	Dro	250		3	<b>.</b>	**- 3	255	
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Asp 305	Ala	Phe	Ala	Leu	Trp 310	Leu	Ile	Ser	Val	Ile 315	Cys	Glu	Ile	Trp	Phe 320
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Arg	Glu	Thr	Tyr 340		Asp	Arg	Leu	Ser 345		Arg	Phe	Asp	Lys 350	Glu	Gly
Gln	Pro	Ser 355	Gln	Leu	Ala	Pro	Ile 360		Phe	Phe	Val	Ser 365		Val	Asp
Pro	Thr 370	Lys	Glu	Pro	Pro	Leu 375		Thr	Ala	Asn	Thr		Leu	Ser	Ile
Leu 385	Ser	Val	Asp	Tyr	Pro 390		Glu	Lys	Val	Ser 395		Tyr	Val	Ser	
	Gly	Ala	Ala	Met 405	Leu	Thr	Phe	Glu			Ser	Glu	Thr		400 Glu
Phe	Ala	Lys	Lys 420		Val	Pro	Phe		410 Lys	Lys	Phe	Asn		415 Glu	Pro
Arg	Ala	Pro		Trp	Tyr	Phe		425 Gln	Lys	Ile	Asp	Tyr	430 Leu	Lys	Asp
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Lys	Val	Pro	Glu	Glu 485	Gly	Trp	Thr	Met	Gln 490	Asp	Gly	Ser	Pro	Trp 495	Pro
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Thr Leu Lys Ser Ala Ser Pro Ala Ser Leu Leu Lys Glu Ala Ile His
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Val Ile Ser Cys Gly Tyr Glu Asp Lys Thr Asp Trp Gly Lys Glu Ile
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Gly Trp Ile Tyr Gly Ser Ile Thr Glu Asp Ile Leu Thr Gly Phe Lys
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              825 830
Val Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Phe Phe Ser Lys His
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Cys Pro Leu Trp Tyr Gly Tyr Gly Gly Gly Leu Lys Phe Leu Glu Arg
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Phe Ser Tyr Ile Asn Ser Ile Val Tyr Pro Trp Thr Ser Ile Pro Leu
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Ile Thr Pro Glu Leu Thr Asn Val Ala Ser Ile Trp Phe Met Ala Leu
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Phe Leu Lys Gly Leu Val Gly Arg Gln Asn Arg Thr Pro Thr Ile Val
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                       Met Glu Gly Asp Ala Asp Gly Val Lys Ser Gly
agg cgc ggt ggc gga cag gtg tgc cag atc tgc ggc gac ggc gtg ggc
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Arg Arg Gly Gly Gln Val Cys Gln Ile Cys Gly Asp Gly Val Gly
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acc acg gcg gag ggg gac gtc ttc gcc gcc tgc gac gtc tgc ggg ttt
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Thr Thr Ala Glu Gly Asp Val Phe Ala Ala Cys Asp Val Cys Gly Phe
ccg gtg tgc cgc ccc tgc tac gag tac gag cgc aag gac ggc acg cag
                                                                      497
Pro Val Cys Arg Pro Cys Tyr Glu Tyr Glu Arg Lys Asp Gly Thr Gln
gcg tgc ccc cag tgc aag acc aag tac aag cgc cac aag ggg agc ccg
                                                                      545
Ala Cys Pro Gln Cys Lys Thr Lys Tyr Lys Arg His Lys Gly Ser Pro
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Ala Ile Arg Gly Glu Glu Gly Asp Asp Thr Asp Ala Asp Ser Asp Phe
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85

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Arg Met Arg Ser Trp Arg Met Asn Val Gly Gly Ser Gly Asp Val Gly

100

95

PCT/US99/18760

641

689

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tca Ser	gga Gly	gaa Glu	atc Ile	cct Pro 160	Gly	gct Ala	tcc Ser	cct Pro	gac Asp 165	cat His	cat His	atg Met	atg Met	tcc Ser 170	Pro	833
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ccc Pro 220	atg Met	acg Thr	aat Asn	ggc Gly	aca Thr 225	agc Ser	att Ile	gct Ala	ccc Pro	tct Ser 230	gag Glu	ggt Gly	cgg Arg	ggt Gly	gtt Val 235	1025
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acg Thr	tac Tyr	ctt Leu	gat Asp 335	agg Arg	ctg Leu	gca Ala	Leu	agg Arg 340	tat Tyr	gac Asp	cgg Arg	gaa Glu	ggt Gly 345	gag Glu	cca Pro	1361

tct Ser	cag Gln	ttg Leu 350	Ala	gct Ala	gtt Val	gac Asp	att Ile 355	Phe	gtc Val	agt Ser	aca Thr	gto Val	Asp	cca Pro	atg Met	1409
aag Lys	gag Glu 365	Pro	cct Pro	ctt Leu	gto Val	act Thr 370	gcc Ala	aat Asn	acc Thr	gtg Val	cta Leu 375	Ser	att	ctt Leu	gct Ala	1457
gtg Val 380	Asp	tac Tyr	cct Pro	gtg Val	gat Asp 385	Lys	gtc Val	tct Ser	tgc Cys	tat Tyr 390	Val	tct Ser	gat Asp	gat Asp	gga Gly 395	1505
gct Ala	gcg Ala	atg Met	ctg Leu	aca Thr 400	ttt Phe	gat Asp	gca Ala	cta Leu	gct Ala 405	gag Glu	act Thr	tca Ser	gag Glu	ttt Phe 410	Ala	1553
aga Arg	aaa Lys	tgg Trp	gta Val 415	cca Pro	ttt Phe	gtt Val	aag Lys	aag Lys 420	tac Tyr	aac Asn	att Ile	gaa Glu	cct Pro 425	aga Arg	gct Ala	1601
cct Pro	gaa Glu	tgg Trp 430	tac Tyr	ttc Phe	tcc Ser	cag Gln	aaa Lys 435	att Ile	gat Asp	tac Tyr	ttg Leu	aag Lys 440	gac Asp	aaa Lys	gtg Val	1649
cac His	cct Pro 445	tca Ser	ttt Phe	gtt Val	aaa Lys	gac Asp 450	cgc Arg	cgg Arg	gcc Ala	atg Met	aag Lys 455	aga Arg	gaa Glu	tat Tyr	gaa Glu	1697
gaa Glu 460	ttc Phe	aaa Lys	gtt Val	agg Arg	gta Val 465	aat Asn	ggc Gly	ctt Leu	gtt Val	gct Ala 470	aag Lys	gca Ala	cag Gln	aaa Lys	gtt Val 475	1745
cct	gag Glu	gaa Glu	gga Gly	tgg Trp 480	atc Ile	atg Met	caa Gln	gat Asp	ggc Gly 485	aca Thr	cca Pro	tgg Trp	cca Pro	gga Gly 490	aac Asn	1793
aat Asn	acc Thr	mgg Xaa	gac Asp 495	cat His	cct Pro	gga Gly	atg Met	att Ile 500	cag Gln	gtt Val	ttc Phe	ctt Leu	ggt Gly 505	cac His	agt Ser	1841
ggt Gly	ggc Gly	ctt Leu 510	gat Asp	act Thr	gag Glu	ggc Gly	aat Asn 515	gag Glu	cta Leu	ccc Pro	cgt Arg	ttg Leu 520	gtc Val	tat Tyr	gtt Val	1889
tct Ser	cgt Arg 525	gaa Glu	aag Lys	cgt Arg	cct Pro	gga Gly 530	ttc Phe	cag Gln	cat His	cac His	aag Lys 535	aaa Lys	gct Ala	ggt Gly	gcc Ala	1937
atg Met 540	aat Asn	gct Ala	ctt Leu	gtt Val	cgt Arg 545	gtc Val	tca Ser	gct Ala	gtg Val	ctt Leu 550	acc Thr	aat Asn	gga Gly	caa Gln	tac Tyr 555	1985
atg Met	ttg Leu	aat Asn	ctt Leu	gat Asp 560	tgt Cys	gat Asp	cac His	tac Tyr	att Ile 565	aac Asn	aac Asn	agt Ser	aag Lys	gct Ala 570	ctc Leu	2033
agg Arg	gaa Glu	gct Ala	atg Met	tgc Cys	ttc Phe	ctt Leu	atg Met	gac Asp	cct Pro	aac Asn	cta Leu	gga Gly	agg Arg	agt Ser	gtc Val	2081

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								_	Ū							
			575					580					585			
tgc Cys	tac Tyr	gtc Val 590	cag Gln	ttt Phe	ccc Pro	cag Gln	aga Arg 595	ttc Phe	gat Asp	gly	att Ile	gac Asp 600	agg Arg	aat Asn	gat Asp	2129
cga Arg	tat Tyr 605	gcc Ala	aac Asn	agg Arg	aac Asn	acc Thr 610	gtg Val	ttt Phe	ttc Phe	gat Asp	att Ile 615	aac Asn	ttg Leu	aga Arg	ggt Gly	2177
ctt Leu 620	gat Asp	ggc	atc Ile	caa Gln	gga Gly 625	cca Pro	gtt Val	tat Tyr	gtc Val	gga Gly 630	act Thr	ggc Gly	tgt Cys	gtt Val	ttc Phe 635	2225
aac Asn	cga Arg	aca Thr	gct Ala	cta Leu 640	tat Tyr	ggt Gly	tat Tyr	gag Glu	ccc Pro 645	cca Pro	att Ile	aag Lys	cag Gln	aag Lys 650	aag Lys	2273
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tca Ser	aag Lys	aag Lys 670	ggc Gly	tcg Ser	gac Asp	aag Lys	aag Lys 675	aag Lys	tcg Ser	cag Gln	aag Lys	cat His 680	gtg Val	gac Asp	agt Ser	2369
tct Ser	gtg Val 685	cca Pro	gta Val	ttc Phe	aac Asn	ctt Leu 690	gaa Glu	gat Asp	ata Ile	gag Glu	gag Glu 695	gga Gly	gtt Val	gaa Glu	ggc ggc	2417
		ttt Phe														2465
gag Glu	aag Lys	aga Arg	ttt Phe	ggc Gly 720	cag Gln	tcc Ser	gca Ala	gcg Ala	ttt Phe 725	gtt Val	gcc Ala	tcc Ser	act Thr	ctg Leu 730	atg Met	2513
gag Glu	tat Tyr	ggt Gly	ggt Gly 735	Val	cct Pro	Gln	Ser	gca Ala 740	Thr	ccg Pro	gag Glu	tct Ser	ctt Leu 745	ctg Leu	aaa Lys	2561
gaa Glu	gct Ala	atc Ile 750	cat His	gtt Val	ata Ile	agc Ser	tgt Cys 755	ggc Gly	tat Tyr	gag Glu	gac Asp	aag Lys 760	act Thr	gaa Glu	tgg Trp	2609
gga Gly	act Thr 765	gag Glu	atc Ile	Gly 999	tgg Trp	atc Ile 770	tac Tyr	ggt Gly	tct Ser	gtg Val	aca Thr 775	gaa Glu	gac Asp	att Ile	ctc Leu	2657
acc Thr 780	gga Gly	ttc Phe	aag Lys	atg Met	cac His 785	gcg Ala	cga Arg	ggc Gly	tgg Trp	cgg Arg 790	tcg Ser	atc Ile	tac Tyr	tgc Cys	atg Met 795	2705
ccc Pro	aag Lys	cgg Arg	cca Pro	gct Ala 800	ttc Phe	aag Lys	gly ggg	tct Ser	gcc Ala 805	ccc Pro	atc Ile	aat Asn	ctt Leu	tcg Ser 810	gac Asp	2753

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cgt Arg	ctg Leu	aac Asn	cag Gln 815	vai	cto Leu	cgg Arg	tgg Trp	gct Ala 820	Leu	gly ggg	tcc Ser	gtg Val	gag Glu 825	Ile	ctc Leu	2801
tto Phe	agc Ser	cgg Arg 830	His	tgc Cys	Pro	ctg Leu	tgg Trp 835	tac Tyr	Gly	tac Tyr	gga Gly	ggg Gly 840	Arg	Ctc	aag Lys	2849
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tcc Ser 860	TTE	ccg Pro	ctt Leu	ctc Leu	atc Ile 865	tac Tyr	tgc Cys	atc Ile	ctg Leu	ccc Pro 870	Ala	atc Ile	tgt Cys	ctg Leu	ctc Leu 875	2945
acc Thr	gga Gly	aag Lys	ttc Phe	atc Ile 880	att Ile	cca Pro	gag Glu	atc Ile	agc Ser 885	aac Asn	ttc Phe	gcc Ala	agc Ser	atc Ile 890	tgg Trp	2993
ttc Phe	atc Ile	tcc Ser	ctc Leu 895	ttc Phe	atc Ile	tcg Ser	atc Ile	ttc Phe 900	gcc Ala	acg Thr	ggc Gly	atc Ile	ctg Leu 905	gag Glu	atg Met	3041
agg Arg	tgg Trp	agc Ser 910	gl <sup>à</sup> aaa	gtg Val	ggc	atc Ile	gac Asp 915	gag Glu	tgg Trp	tgg Trp	agg Arg	aac Asn 920	gag Glu	cag Gln	ttc Phe	3089
tgg Trp	gtg Val 925	atc Ile	gjà aaa	ggc Gly	atc Ile	tcc Ser 930	gcg Ala	cac His	ctc Leu	ttc Phe	gcc Ala 935	gtg Val	ttc Phe	cag Gln	ggc Gly	3137
ctg Leu 940	ctc Leu	aag Lys	gtg Val	ctg Leu	gcc Ala 945	ggc	atc Ile	gac Asp	acc Thr	aac Asn 950	ttc Phe	acc Thr	gtc Val	acc Thr	tcc Ser 955	3185
aag Lys	gcc Ala	tcg Ser	gac Asp	gag Glu 960	gac Asp	ggc Gly	gac Asp	ttc Phe	gcg Ala 965	gag Glu	ctg Leu	tac Tyr	atg Met	ttc Phe 970	aag Lys	3233
tgg Trp	acg Thr	acg Thr	ctc Leu 975	ctg Leu	atc Ile	ccg Pro	ccc Pro	acc Thr 980	acc Thr	atc Ile	ctg Leu	atc Ile	atc Ile 985	aac Asn	ctg Leu	3281
gtc Val	ggc	gtc Val 990	Val	gcc Ala	ggc Gly	atc Ile	tcc Ser 995	Tyr	gcc Ala	atc Ile	aac Asn	agc Ser 1000	Gly	tac Tyr	cag Gln	3329
tcg Ser	tgg Trp 1005	GIA	ccg Pro	ctc Leu	ttc Phe	ggc Gly 1010	Lys	ctc Leu	ttc Phe	ttc Phe	gcc Ala 1015	Phe	tgg Trp	gtc Val	atc Ile	3377
gtc Val 1020	HIS	ctg Leu	tac Tyr	ccg Pro	ttc Phe 1025	ctc Leu	aag Lys	ggc Gly	Leu	atg Met 1030	Gly	agg Arg	cag Gln	Asn	cgc Arg 1035	3425
acc Thr	ccg Pro	acc Thr	atc Ile	gtc Val	gtc Val	gtc Val	tgg Trp	gcca	teet	gc t	ggcg	tcca	t ct	tctc	cttg	3479

## 1040

gg:	catca catca caca	get gecc	geta	iggga gtta ggata	aag t aag t att g	iggaa itata yttta	aggtt atata accad	it gi at ai	tacti	ttgta cagca	a gaa	acgg	gagg	aata	gacgtgt accacgt acagcta attcttt	3539 3599 3659 3719 3746
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	<	212:	PR1													
	<	213:	> Zea	may	/S											
	<	400>	> 50													
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			20					25					30		Gly	
Asp	Val	Phe 35	Ala	Ala	Cys	Asp	Val	Cys	Gly	Phe	Pro	Val 45	Cys	Arg	Pro	
Cys	Tyr 50	Glu	Tyr	Glu	Arg	Lys	Asp	Gly	Thr	Gln	Ala	Cys	Pro	Gln	Cys	
Lys 65	Thr	Lys	Tyr	Lys	Arg		Lys	Gly	Ser		Ala	Ile	Arg	Gly	Glu	
	Gly	Asp	Asp	Thr		Ala	Asp	Ser	Asp	75 Phe	Asn	Tyr	Leu	Ala	80 Ser	
				85					90					95		
			Asp 100					105					110		-	
		TTO					120					125				
Ser	Gly 130	Glu	Ile	Gly	Leu	Thr 135	Lys	Tyr	Asp	Ser	Gly 140	Glu	Ile	Pro	Arg	
Gly	Tyr	Ile	Pro	Ser	Val	Thr	Asn	Ser	Gln	Ile	Ser	Gly	Glu	Ile	Pro	
145 Gly		Ser	Pro	Asp	150 His	Hie	Met	Mot	C0.20	155	m\	<b>~</b> 1	_		160	
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			Pro 180					185					190			
Glu	Phe	Ser	Gly	Ser	Ile	Gly	Asn 200	Val	Ala	Trp	Lys		Arg	Val	Asp	
Gly	Trp 210	Lys	Met	Lys	Gln	Asp 215		Gly	Thr	Ile		205 Met	Thr	Asn	Gly	
Thr		Ile	Ala	Pro	Ser		Gly	Arg	Gly	Val	220 Gly	Asp	Ile	Asp	Ala	
225					230					235					240	
501	1111	Asp	Tyr	245	wet	GIU	Asp	Ala	Leu 250	Leu	Asn	Asp	Glu		Arg	
Gln	Pro	Leu	Ser 260	Arg	Lys	Val	Pro	Leu 265		Ser	Ser	Arg		255 Asn	Pro	
Tyr	Arg	Met 275	Val	Ile	Val	Leu	Arg 280		Ile	Val	Leu		270 Ile	Phe	Leu	
His	Tyr 290		Ile	Thr	Asn	Pro	Val	Arg	Asn	Ala		285 Pro	Leu	Trp	Leu	
Leu 305		Val	Ile	Cys	Glu	295 Ile	Trp	Phe	Ala	Leu	300 Ser	Trp	Ile	Leu	Asp	
505			Lys		310					315					220	
				325					330					225		
Leu	Ala	Leu	Arg 340	Tyr	Asp	Arg	Glu	Gly 345	Glu	Pro	Ser	Gln	Leu 350	Ala	Ala	

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Val Asp Ile Phe Val Ser Thr Val Asp Pro Met Lys Glu Pro Pro Leu
                           360
Val Thr Ala Asn Thr Val Leu Ser Ile Leu Ala Val Asp Tyr Pro Val
                      375
                                          380
 Asp Lys Val Ser Cys Tyr Val Ser Asp Asp Gly Ala Ala Met Leu Thr
                   390
                                      395
Phe Asp Ala Leu Ala Glu Thr Ser Glu Phe Ala Arg Lys Trp Val Pro
               405
                                  410
Phe Val Lys Lys Tyr Asn Ile Glu Pro Arg Ala Pro Glu Trp Tyr Phe
           420
                              425
Ser Gln Lys Ile Asp Tyr Leu Lys Asp Lys Val His Pro Ser Phe Val
                           440
Lys Asp Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg
                       455
Val Asn Gly Leu Val Ala Lys Ala Gln Lys Val Pro Glu Glu Gly Trp
                   470
                           475
Ile Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn Thr Xaa Asp His
               485
                                  490
Pro Gly Met Ile Gln Val Phe Leu Gly His Ser Gly Gly Leu Asp Thr
           500
                              505
Glu Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg
                          520
Pro Gly Phe Gln His His Lys Lys Ala Gly Ala Met Asn Ala Leu Val
                       535
Arg Val Ser Ala Val Leu Thr Asn Gly Gln Tyr Met Leu Asn Leu Asp
                   550
                                      555
Cys Asp His Tyr Ile Asn Asn Ser Lys Ala Leu Arg Glu Ala Met Cys
                                  570
Phe Leu Met Asp Pro Asn Leu Gly Arg Ser Val Cys Tyr Val Gln Phe
                               585
Pro Gln Arg Phe Asp Gly Ile Asp Arg Asn Asp Arg Tyr Ala Asn Arg
                           600
                                              605
Asn Thr Val Phe Phe Asp Ile Asn Leu Arg Gly Leu Asp Gly Ile Gln
                      615
                                          620
Gly Pro Val Tyr Val Gly Thr Gly Cys Val Phe Asn Arg Thr Ala Leu
                                     635
Tyr Gly Tyr Glu Pro Pro Ile Lys Gln Lys Lys Gly Gly Phe Leu Ser
                                 650
Ser Leu Cys Gly Gly Arg Lys Lys Ala Ser Lys Ser Lys Gly Ser
                             665
Asp Lys Lys Ser Gln Lys His Val Asp Ser Ser Val Pro Val Phe
                          680
Asn Leu Glu Asp Ile Glu Glu Gly Val Glu Gly Ala Gly Phe Asp Asp
                      695
                                          700
Glu Lys Ser Leu Leu Met Ser Gln Met Ser Leu Glu Lys Arg Phe Gly
        710
                                      715
Gln Ser Ala Ala Phe Val Ala Ser Thr Leu Met Glu Tyr Gly Gly Val
                                  730
Pro Gln Ser Ala Thr Pro Glu Ser Leu Leu Lys Glu Ala Ile His Val
                              745
Ile Ser Cys Gly Tyr Glu Asp Lys Thr Glu Trp Gly Thr Glu Ile Gly
                          760
Trp Ile Tyr Gly Ser Val Thr Glu Asp Ile Leu Thr Gly Phe Lys Met
                       775
                                         780
His Ala Arg Gly Trp Arg Ser Ile Tyr Cys Met Pro Lys Arg Pro Ala
                  790
                                     795
Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp Arg Leu Asn Gln Val
                                  810
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Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Leu Phe Ser Arg His Cys
 Pro Leu Trp Tyr Gly Tyr Gly Gly Arg Leu Lys Phe Leu Glu Arg Phe
                         840
 Ala Tyr Ile Asn Thr Thr Ile Tyr Pro Leu Thr Ser Ile Pro Leu Leu
             855
 Ile Tyr Cys Ile Leu Pro Ala Ile Cys Leu Leu Thr Gly Lys Phe Ile
                  870
                           875
 Ile Pro Glu Ile Ser Asn Phe Ala Ser Ile Trp Phe Ile Ser Leu Phe
              885
                       890
 Ile Ser Ile Phe Ala Thr Gly Ile Leu Glu Met Arg Trp Ser Gly Val
           900
                             905
 Gly Ile Asp Glu Trp Trp Arg Asn Glu Gln Phe Trp Val Ile Gly Gly
                         920
 Ile Ser Ala His Leu Phe Ala Val Phe Gln Gly Leu Leu Lys Val Leu
           935
 Ala Gly Ile Asp Thr Asn Phe Thr Val Thr Ser Lys Ala Ser Asp Glu
         950
 Asp Gly Asp Phe Ala Glu Leu Tyr Met Phe Lys Trp Thr Thr Leu Leu
              965
 Ile Pro Pro Thr Thr Ile Leu Ile Ile Asn Leu Val Gly Val Val Ala
                              985
 Gly Ile Ser Tyr Ala Ile Asn Ser Gly Tyr Gln Ser Trp Gly Pro Leu
                          1000
 Phe Gly Lys Leu Phe Phe Ala Phe Trp Val Ile Val His Leu Tyr Pro
   1010 1015
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                                     1035
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gto	ggrg	ıccg	gtgt	cgtt	gg c	gtgt	ggag	c cq	tctc	aata	ффа	acao	caa	ggag	agaaca	180
gag	atg	gc9	gcc	aac	aag	agg	atg	gtg	gcq	ggc	tca	cac	aac	: cac	aac	228
	Met	: Ala	ı Ala	Asn	Lys	Gly	Met	Val	Ala	Gly	Ser	His	Asn	Aro	Asn	220
	1				5					10				3	15	
gag	ttc	gto	atg	atc	cgc	cac	gac	ggc	gat	gtg	ccg	gge	tcq	qct	aag	276
Glu	Phe	: Val	Met	Ile	Arg	His	Asp	Gly	Asp	Val	Pro	Gly	Ser	Ala	Lys	7.0
				20					25			•		30		
ccc	aca	aag	agt	gcg	aat	gga	cag	gtc	tgc	cag	att	tgc	ggt	gac	tct	324
Pro	Thr	Lys	Ser	Ala	Asn	Gly	Gln	Val	Cys	Gln	Ile	Cys	Gly	Asp	Ser	
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grg	ggt	gtt	tca	gcc	act	ggt	gat	gtc	ttt	gtt	gcc	tgc	aat	gag	tgt	372
vaı	GIY	vai	ser	Ala	Thr	Gly	Asp	Val	Phe	Val	Ala	Cys	Asn	Glu	Cys	
		50					55					60				
aaa	++~	-a		<b>.</b>			_									
Ala	Dho	משם	ycc v.1	tgc	cgc	cca	tgc	tat	gag	tat	gag	cgc	aag	gag	aaa	420
nia	65	PIU	vai	Cys	Arg		Cys	Tyr	GLu	Tyr		Arg	Lys	Glu	Gly	
	0.5					70					75					
aac	caa	tac	tac	ccc	cac	+~~	225									
Asn	Gln	Cvs	Cvs	ccc Pro	Gln	Cyc	aag	act The	aga	tac	aag	aga	cag	aaa	ggt	468
80		<b>-</b> 1-2	<b>475</b>		85	Cys	ьуѕ	TIIL	Arg		гÀг	Arg	GIn	Lys		
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age	cct	cga	att	cat	aat	ra t	<b>~</b>	ant.	~~~	~~~						
Ser	Pro	Ara	Val	His	Glv	Asn	Glu	Nan	Glu	Clu	gat	gtt	gat	gac	cta -	516
				100	0-1	Mop	Gru	vəħ	105	GIU	ASD	vai	Asp		Leu	
									103					110		
gac	aat	gaa	ttc	aac	tac	aaσ	caa	aac	agt	aaa	222	<b>777</b>		~~~	•	F.C.4
Asp	Asn	Glu	Phe	Asn	Tvr	Lvs	Gln	Glv	Ser	233	Lve	Glv	Dro	gag	rgg Tom	564
			115		•	4		120		<b>-</b> - <i>y</i>	Lys	GIY	125	GIU	тъ	
													12.7			
caa	ctg	caa	gga	gat	gat	gct	gat	ctq	tct	tca	tct	act	cac	cat	gag	612
Gln	Leu	Gln	Gly	Asp	Asp	Ala	Asp	Leu	Ser	Ser	Ser	Ala	Ara	His	Glu	012
		130					135					140	3			
cca	cat	cat	cgg	att	cca	cgc	ctg	aca	agc	ggt	caa	cag	ata	tct	qqa	660
Pro	His	His	Arg	Ile	Pro	Arg	Leu	Thr	Ser	Gly	Gln	Gln	Ile	Ser	Gly	
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GIU	TIE	Pro	Asp	Ala	Ser	Pro	Asp	Arg	His	Ser	Ile	Arg	Ser	Pro	Thr	
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ccg	agc	tat	gtt	gat	cca -	agc	gtc	cca	gtt	cct	gtg	agg	att	gtg	gac	756
ser	ser	Tyr	Val	Asp	Pro	Ser	Val	Pro	Val	Pro	Val	Arg	Ile	Val	Asp	
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	<b>.</b>															
D~~	cog	aag	gac	ttg	aat	tcc	tat -	ggg	ctt	aat	agt	gtt	gac	tgg	aag	804
PT.O	ser	ьys	Asp	Leu	Asn	Ser	Tyr	Gly	Leu	Asn	Ser	Val	Asp	Trp	Lys	
			195					200					205			
ma a	2012	~++	~~~	20-	<b>-</b>											
Glu	Ara	yct Val	gag	agc	rgg	agg	gtt	aaa	cag	gac	aaa	aat	atg	atg	caa	852
	9	, ur	JIU	Ser	тър	мгд	val	гуѕ	Gin .	Asp	ьys	Asn	Met	Met	Gln	

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210		215	220	
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	atc gtg cca att Ile Val Pro Ile 260			
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	cat cca gtg cgt His Pro Val Arg			•
	gag gtc tgg ttt Glu Val Trp Phe 310			_
	tat cca atc aac Tyr Pro Ile Asn 325	Arg Glu Thr		
	gat aga gag gga Asp Arg Glu Gly 340			_
	agt aca gtg gat Ser Thr Val Asp 355			
	gtt ttg tcc att Val Leu Ser Ile			
	tat gtt tct gat Tyr Val Ser Asp 390			
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	aat att gaa cca Asn Ile Glu Pro 420			_
	tac ctg aag gac Tyr Leu Lys Asp 435			

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gcc Ala	Leu 465	vai	gcc Ala	aaa Lys	gca Ala	cag Gln 470	aaa Lys	gtg Val	g cct Pro	gaa Glu	gag Glu 475	Gly	tgg	acc Thr	atg Met	1620
gct Ala 480	Asp	gga Gly	act Thr	gca Ala	tgg Trp 485	Pro	GJ A aaa	aat Asn	aat Asn	cct Pro 490	Arg	gac Asp	cat	cct Pro	ggc Gly 495	1 <u>6</u> 68
atg Met	att Ile	cag Gln	gtt Val	ttc Phe 500	Leu	Gly	cac	agt Ser	ggt Gly 505	gly	ctc Leu	gac Asp	act Thr	gat Asp 510	Gly	1716
aat Asn	gag Glu	tta Leu	cca Pro 515	cgt Arg	ctt Leu	gtc Val	tat Tyr	gto Val 520	tct Ser	cgt Arg	gaa Glu	aag Lys	aga Arg 525	cca Pro	ggc Gly	1764
ttt Phe	cag Gln	cat His 530	His	aag Lys	aag Lys	gct Ala	ggt Gly 535	gca Ala	atg Met	aat Asn	gcg Ala	ctg Leu 540	att Ile	cgt Arg	gta Val	1812
tct Ser	gct Ala 545	gtg Val	ctg Leu	aca Thr	aat Asn	ggt Gly 550	gcc Ala	tat Tyr	ctt Leu	ctc Leu	aat Asn 555	gtg Val	gat Asp	tgc Cys	gac Asp	1860
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aga Arg	ttt Phe	gat Asp	ggc Gly 595	att Ile	gac Asp	ttg Leu	cac His	gat Asp 600	cga Arg	tat Tyr	gct Ala	aat Asn	cgg Arg 605	aac Asn	ata Ile	2004
gtt Val	ttc Phe	ttt Phe 610	gat Asp	atc Ile	aac Asn	atg Met	aaa Lys 615	ggt Gly	ctg Leu	gat Asp	ggc	att Ile 620	cag Gln	ggt Gly	cca Pro	2052
gtt Val	tac Tyr 625	gtg Val	gga Gly	aca Thr	gga Gly	tgc Cys 630	tgt Cys	ttc Phe	aat Asn	aga Arg	cag Gln 635	gct Ala	ttg Leu	tat Tyr	gga Gly	2100
tac Tyr 640	gat Asp	cct Pro	gtt Val	ttg Leu	act Thr 645	gaa Glu	gct Ala	gat Asp	ctg Leu	gag Glu 650	cca Pro	aac Asn	att Ile	gtt Val	att Ile 655	2148
aag Lys	agc Ser	tgc Cys	Cys	ggt Gly 660	aga Arg	agg Arg	aag Lys	aaa Lys	aag Lys 665	aac Asn	aag Lys	agt Ser	tat Tyr	atg Met 670	gat Asp	2196
agt Ser	caa Gln	agc Ser	cgt Arg	att Ile	atg Met	aag Lys	aga Arg	aca Thr	gaa Glu	tct Ser	tca Ser	gct Ala	ccc Pro	atc Ile	ttc Phe	2244

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				agg Arg 710							_	2340
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				cta Leu								2436
				act Thr								2484
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				tac Tyr 790								2580
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				ttg Leu								2916

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gaa Glu	gat Asp	tgg Trp	tgg Trp 915	aga Arg	aat Asn	gag Glu	cag Gln	ttt Phe 920	tgg Trp	gtt Val	att Ile	ggt Gly	ggc Gly 925	acc Thr	tct Ser	2964
gcc Ala	cat His	ctc Leu 930	ttc Phe	gca Ala	gtg Val	ttc Phe	cag Gln 935	ggt Gly	ctg Leu	ctg Leu	aaa Lys	gtg Val 940	ttg Leu	gct Ala	gjå aaa	3012
att Ile	gat Asp 945	acc Thr	aac Asn	ttc Phe	aca Thr	gtt Val 950	acc Thr	tca Ser	aag Lys	gca Ala	tct Ser 955	gat Asp	gag Glu	gat Asp	gly ggc	30,60
gac Asp 960	ttt Phe	gct Ala	gag Glu	cta Leu	tat Tyr 965	gtg Val	ttc Phe	aag Lys	tgg Trp	acc Thr 970	agt Ser	ttg Leu	ctc Leu	att Ile	cct Pro 975	3108
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tgg Trp 1040	Ser	atc Ile	ctt Leu	ctt Leu	gca Ala 1045	Ser	atc Ile	ttc Phe	tcc Ser	ttg Leu 1050	Leu	tgg Trp	gtg Val	aag Lys	atc Ile 1055	3348
gat Asp	cct Pro	ttc Phe	atc Ile	tcc Ser 1060	Pro	aca Thr	cag Gln	aaa Lys	gct Ala 1065	Ala	gcc Ala	ttg Leu	gly aaa	caa Gln 1070	Cys	3396
ggc	gtc Val	aac Asn	t go	tgat:	cgag	, aca	ıgtga	ictc	ttat	ttga	ıag a	.ggct	caat	c		3446
gtga tgct aaga	ggat gcgg tgtg tttg	gg a ac t aa t	tttg aaga tttg	rcato latca laagt	t aa c gg t tt	igtta Iagco Igtta	itged :ttte itgeg	tet tac tgc	gtto ctto agtt	att cat tat	agct gtag tgtt	tctt cgcc ttag	cc g ag c ag t	tgcc cagc aaat	tgtag ggtgc agcgt tatca aaaaa	3506 3566 3626 3686 3746 3753

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Phe 65	Pro	Val	Cys	Arg	Pro 70	Суз	Tyr	Glu	Tyr	Glu 75	Arg	Lys	Glu	Gly	Asn 80
Gln	Cys	Cys	Pro	Gln 85	Cys	Lys	Thr	Arg	Tyr 90	Lys	Arg	Gln	Lys	Gly 95	
Pro	Arg	Val	His 100	Gly	Asp	Glu	Asp	Glu 105	Glu	Asp	Val	Asp	Asp		Asp
Asn	Glu	Phe 115	Asn	Tyr	Lys	Gln	Gly 120	Ser	Gly	Lys	Gly	Pro 125		Trp	Gln
Leu	Gln 130	Gly	Asp	Asp	Ala	Asp 135	Leu	Ser	Ser	Ser	Ala 140	Arg	His	Glu	Pro
His 145	His	Arg	Ile	Pro	Arg 150	Leu	Thr	Ser	Gly	Gln 155	Gln	Ile	Ser	Gly	Glu 160
				165	Pro				170					175	
			180		Ser			185					190		
		195			Ser		200					205			
	210				Arg	215					220				
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				245	Met				250					255	
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Phe	Val	Ser 355	Thr	Val	Asp	Pro	Leu 360		Glu	Pro	Pro	Leu 365		Thr	Ala
Asn	Thr 370	Val	Leu	Ser	Ile	Leu 375		Val	Asp	Tyr	Pro 380		Asp	Lys	Val
Ser	Cys	Tyr	Val	Ser	Asp	Asp	Gly	Ser	Ala	Met		Thr	Phe	Glu	Ser
385					390					395					400
				405	Glu				410					415	
			420		Pro			425					430		
		435			Asp		440					445			-
	450				Glu	455					460				
ьeu	val	Ата	гÀа	Ala	Gln	Lys	Val	Pro	Glu	Glu	Gly	Trp	Thr	Met	Ala

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465					470					475					480
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Ile	Gln	Val	Phe 500	Leu	Gly	His	Ser	Gly 505	Gly	Leu	Asp	Thr	Asp	Gly	Asn
Glu	Leu	Pro 515	Arg	Leu	Val	Tyr	Val 520		Arg	Glu	Lys	Arg	Pro	Gly	Phe
Gln	His 530	His	Lys	Lys	Ala	Gly 535	Ala	Met	Asn	Ala	Leu 540	Ile	Arg	Val	Ser
Ala 545	Val	Leu	Thr	Asn	Gly 550	Ala	Tyr	Leu	Leu	Asn 555			Cys	Asp	His 560
Tyr	Phe	Asn	Ser	Ser 565	Lys	Ala	Leu	Arg	Glu 570	Ala	Met	Cys	Phe	Met 575	Met
Asp	Pro	Ala	Leu 580	Gly	Arg	Lys	Thr	Cys 585	Tyr	Val	Gln	Phe	Pro	Gln	Arg
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				645		Ala			650					655	
			660			Lys		665					670		
		675				Arg	680					685			
	690					Gly 695					700			-	
705					710	Lys				715					720
				725		Phe			730					735	
			740			Leu		745					750		
		755				Glu	760					765			
	770					Ile 775 Cys					780				
785					790	Ser				795					800
				805		Ile			810					815	
			820			Leu		825					830		
		835				Ile	840					845			
	850					855 Leu					860				
865					870					875					880
				885		Met			890					895	
			900			Glu Gln		905					910		
		915				Gln	920					925			
							y	u	⊔eu	пåя	val	Leu	мта	σтλ	тте

WO 00/09706 PCT/US99/18760

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Asp Thr Asn Phe Thr Val Thr Ser Lys Ala Ser Asp Glu Asp Gly Asp
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Phe Ala Glu Leu Tyr Val Phe Lys Trp Thr Ser Leu Leu Ile Pro Pro
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Thr Thr Val Leu Val Ile Asn Leu Val Gly Met Val Ala Gly Ile Ser
             980
                                985
Tyr Ala Ile Asn Ser Gly Tyr Gln Ser Trp Gly Pro Leu Phe Gly Lys
                            1000
Leu Phe Phe Ser Ile Trp Val Ile Leu His Leu Tyr Pro Phe Leu Lys
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Gly Leu Met Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Ile Val Trp
                    1030
                                        1035
Ser Ile Leu Leu Ala Ser Ile Phe Ser Leu Leu Trp Val Lys Ile Asp
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aat tog ggg aag cat gtg goo ggg cag gtg tgc cag atc tgc ggc gac
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Asn Ser Gly Lys His Val Ala Gly Gln Val Cys Gln Ile Cys Gly Asp
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ggc gtg gg Gly Val Gl 25	c acc gco y Thr Ala	g gcg gac a Ala Asp 30	ggc gac ct Gly Asp Le	c ttc acc gcc u Phe Thr Ala 35	tgc gac gtc Cys Asp Val	388
tgc ggc tt Cys Gly Ph 40	c ccc gtg e Pro Va]	tgc cgc ( Cys Arg ) 45	cca tgc ta Pro Cys Ty	c gag tac gag r Glu Tyr Glu 50	cgc aag gac Arg Lys Asp 55	)
ggc acc ca Gly Thr Gl	g geg tge n Ala Cys 60	Pro Gln	Cys Lys Th	t aag tac aag r Lys Tyr Lys 5	cgc cac aaa Arg His Lys 70	484
ggg agc cc Gly Ser Pr	a cca gta o Pro Val 75	cac ggt o	gag gaa aa Glu Glu As 80	t gag gat gtg n Glu Asp Val	gat gct gac Asp Ala Asp 85	532
gat gtg ag Asp Val Se 9	r Asp Tyr	aac tac o	caa gca tc Gln Ala Se 95	t ggc aac cag r Gly Asn Gln 100	gat cag aag Asp Gln Lys	580
caa aag at Gln Lys Il 105	t gct gag e Ala Glu	aga atg o Arg Met I 110	ctc act tg Leu Thr Tr	g cgg aca aac p Arg Thr Asn 115	tca cgt ggc Ser Arg Gly	628
agt gat at Ser Asp Il 120	t ggc ctg e Gly Leu	gct aag t Ala Lys 1 125	tat gac ag Tyr Asp Se	c ggt gaa att r Gly Glu Ile 130	ggg cat ggg Gly His Gly 135	676
aag tat ga Lys Tyr As	e agt ggt p Ser Gly 140	Glu Ile E	ect cgt gg Pro Arg Gl 14	a tat atc ccg y Tyr Ile Pro 5	tca cta act Ser Leu Thr 150	724
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ggt gca att Gly Ala Ile	cct atg Pro Met 220	acc aat g Thr Asn G	ga aca ago Sly Thr Sen 225	e att gct cca r Ile Ala Pro	tca gaa ggg Ser Glu Gly 230	964
cgt gga gtt Arg Gly Val	gct gat Ala Asp 235	att gat g Ile Asp A	gct tct act la Ser Thi 240	gat tat aac Asp Tyr Asn	atg gaa gat Met Glu Asp 245	1012

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			tcc Ser	_			_		_	_	_				-	1108
_	_	_	cta Leu	_			_	-		_	_	_				1156
			tat Tyr													1204
			tcc Ser 315													1252
	_	_	aca Thr			_	-	_	_				_	_	_	1300
			tct Ser													1348
			aag Lys													1396
			gtc Val													1444
_	_		gct Ala 395	_	_	_										1492
		_	aga Arg			_	_		_	_	_				gag Glu	1540
		Ala					Phe					Asp			aaa Lys	1588
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										cac His			:	1876
										ctt Leu			:	1924
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										aac Asn			:	2020
										ggt Gly			:	2068
										gat Asp			:	2116
							-			gga Gly				2164
		Arg	Ala	Leu	Tyr	Gly	Tyr	Glu	Pro	cca Pro 645	Val			2212
					-		_			agg Arg		-		2260
										cat His				2308
										gag Glu				2356
										atg Met				2404

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act Thr	ctg Leu	atg Met 730	gaa Glu	tat Tyr	ggt Gly	ggt Gly	gtt Val 735	cca Pro	caa Gln	tct Ser	gca Ala	act Thr 740	cca Pro	gag Glu	tct Ser	2500
							gtc Val									2548
act Thr 760	gac Asp	tgg Trp	gga Gly	act Thr	gag Glu 765	att Ile	Gly aaa	tgg Trp	atc Ile	tat Tyr 770	ggt Gly	tct Ser	gtt Val	aca Thr	gaa Glu 775	2596
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Leu	Ser	Asp 810	Arg	Leu	Asn	Gln	gtg Val 815	Leu	Arg	Trp	Ala	Leu 820	Gly	Ser	Ile	2740
Glu	Ile 825	Leu	Phe	Ser	Arg	His 830	tgt Cys	Pro	Ile	Trp	Tyr 835	Gly	Tyr	Gly	Gly	2788
Arg 840	Leu	Lys	Phe	Leu	Glu 845	Arg	ttt Phe	Ala	Tyr	Ile 850	Asn	Thr	Thr	Ile	Tyr 855	2836
Pro	Leu	Thr	Ser	Ile 860	Pro	Leu		Leu	865	Cys	Ile	Leu	Pro	Ala 870	Val	2884
Cys	Leu	Leu	875	Gly	. TÀa	Phe	: Ile	11e	Pro	Lys	Ile	Ser	Asn 885	Leu i	gag Glu	2932
Ser	· Val	890	Phe	: Ile	: Ser	: Leu	Phe 895	Ile	e Ser	: Ile	Phe	900	Thr	Gly	atc Ile	2980
ctt Leu	gag Glu 905	Met	agg : Arg	r tgg Trp	g agt Ser	ggt Gly 910	v Val	ggo Gly	att / Ile	: gat : Asp	gaa Glu 915	Trp	tgg Trp	agg Arg	g aac g Asn	3028
Glu 920	ı Glr	n Phe	e Tr	Va]	925	e Gly	/ Gly	/ Ile	e Sei	930	a His	: Lev	ı Phe	e Ala	gtc Val 935	3076
tto Phe	cag e Glr	g ggt n Gly	cto Lev	cto Lev	g aag 1 Lys	g gtg s Val	g ctt L Lei	gci Ala	ggt a Gly	ato 7 Ile	e gad e Asp	ace Thi	g ago	tto Phe	c act e Thr	3124

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		Tyr Pro Phe L	tc aag ggc ctc atg eu Lys Gly Leu Met 025					
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	Ser Leu Met		tc gat cca ttc acc le Asp Pro Phe Thr 1060					
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Gly 145					150					155					160
Gly	Ala	Ser	Pro	Asp 165	His	Met	Met	Ser	Pro 170	Val	Gly	Asn	Ile	Gly 175	Arg
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		195	Gly				200					205			
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Ser 225	Ile	Ala	Pro	Ser	Glu 230	Gly	Arg	Gly	Val	Ala 235	Asp	Ile	Asp	Ala	Ser 240
	Asp	Tyr	Asn	Met 245		Asp	Ala	Leu	Leu 250		Asp	Glu	Thr	Arg 255	
			Arg 260					265					270		
		275	Ile				280					285			
	290		Thr			295					300				•
	Val	Ile	Cys	Glu		Trp	Phe	Ala	Leu	Ser 315	Trp	lle	Leu	Asp	320
305 Phe	Pro	Lys	Trp	Ser 325	310 Pro	Ile	Asn	Arg	Glu 330		Tyr	Leu	Asp	Arg 335	
Ala	Leu	Arg	Tyr 340	Asp	Arg	Glu	Gly	Glu 345	Pro	Ser	Gln	Leu	Ala 350	Pro	Val
		355					360					365			
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Lys 385	Val	Ser	Cys	Tyr	Val 390	Ser	Asp	Asp	Gly	Ala 395		Met	Leu	Thr	Phe 400
	Ala	Leu	Ser	Glu 405	Thr	Ser	Glu	Phe	Ala 410	Arg		Trp	Val	Pro	
Суѕ	Lys	Lys	Tyr 420		Ile	Glu	Pro	Xaa 425	Ala	Pro	Glu	Trp	Tyr 430		Ala
		435					440					445			Lys
Glu	Arg 450		Ala	Met	Lys	Arg 455		Tyr	Glu	Glu	Phe 460		Val	Arg	Ile
	Gly	Leu	val	Ala			Gln	Lys	Val			Glu	Gly	Trp	Ile
465 Met	Gln	Asr	Gly	Thr	470 Pro		Pro	Gly	Asn	475 Asn		Arg	Asp	His	480 Pro
				485	,				490	)				495	,
			500	)				505					510	)	Glu
_		515	5				520	+				525	;		Pro
_	530	)				535	,				540	<b>)</b>			Arg
Val 545		Ala	a Val	Let	1 Thr 550		ı Gly	Gln	Туг	Met 555		Asn	Lev	Asp	Cys 560
		Туг	: Ile	e Asr			: Lys	Ala	Lev			Ala	Met	Суз	Phe

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		595					600					605		Arg	
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